Package 'MSstats'

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Title Protein Significance Analysis in DDA, SRM and DIA for Label-free
or Label-based Proteomics Experiments

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Description A set of tools for statistical relative protein significance analy-
sis in DDA, SRM and DIA experiments.

License Artistic-2.0

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URL http://msstats.org

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R **topics documented:**

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checkRepeatedDesign   *Check if data represents repeated measurements design*

Description

Check if data represents repeated measurements design

Usage

```
checkRepeatedDesign(summarization_output)
```

Arguments

summarization_output
                output of the dataProcess function

Details

This extracts information required by the group comparison workflow

Value

logical, TRUE if data represent repeated measurements design

Examples

```
QuantData1 <- dataProcess(SRMRawData, use_log_file = FALSE)
checkRepeatedDesign(QuantData1)
```

dataProcess	*Process MS data: clean, normalize and summarize before differential analysis*

Description

Process MS data: clean, normalize and summarize before differential analysis

Usage

```
dataProcess(
  raw,
  logTrans = 2,
  normalization = "equalizeMedians",
  nameStandards = NULL,
  featureSubset = "all",
  remove_uninformative_feature_outlier = FALSE,
  min_feature_count = 2,
  n_top_feature = 3,
  summaryMethod = "TMP",
  equalFeatureVar = TRUE,
  censoredInt = "NA",
  MBimpute = TRUE,
  remove50missing = FALSE,
  fix_missing = NULL,
  maxQuantileforCensored = 0.999,
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL
)
```

Arguments

raw	name of the raw (input) data set.
logTrans	base of logarithm transformation: 2 (default) or 10.
normalization	normalization to remove systematic bias between MS runs. There are three different normalizations supported: 'equalizeMedians' (default) represents constant normalization (equalizing the medians) based on reference signals is performed. 'quantile' represents quantile normalization based on reference signals 'globalStandards' represents normalization with global standards proteins. If FALSE, no normalization is performed.
nameStandards	optional vector of global standard peptide names. Required only for normalization with global standard peptides.

featureSubset	"all" (default) uses all features that the data set has. "top3" uses top 3 features which have highest average of log-intensity across runs. "topN" uses top N features which has highest average of log-intensity across runs. It needs the input for n_top_feature option. "highQuality" flags uninformative feature and outliers.
remove_uninformative_feature_outlier	
	optional. Only required if featureSubset = "highQuality". TRUE allows to remove 1) noisy features (flagged in the column feature_quality with "Uninformative"), 2) outliers (flagged in the column, is_outlier with TRUE, before run-level summarization. FALSE (default) uses all features and intensities for run-level summarization.
min_feature_count	
	optional. Only required if featureSubset = "highQuality". Defines a minimum number of informative features a protein needs to be considered in the feature selection algorithm.
n_top_feature	optional. Only required if featureSubset = 'topN'. It that case, it specifies number of top features that will be used. Default is 3, which means to use top 3 features.
summaryMethod	"TMP" (default) means Tukey's median polish, which is robust estimation method. "linear" uses linear mixed model.
equalFeatureVar	
	only for summaryMethod = "linear". default is TRUE. Logical variable for whether the model should account for heterogeneous variation among intensities from different features. Default is TRUE, which assume equal variance among intensities from features. FALSE means that we cannot assume equal variance among intensities from features, then we will account for heterogeneous variation from different features.
censoredInt	Missing values are censored or at random. 'NA' (default) assumes that all 'NA's in 'Intensity' column are censored. '0' uses zero intensities as censored intensity. In this case, NA intensities are missing at random. The output from Skyline should use '0'. Null assumes that all NA intensites are randomly missing.
MBimpute	only for summaryMethod = "TMP" and censoredInt = 'NA' or '0'. TRUE (default) imputes 'NA' or '0' (depending on censoredInt option) by Accelated failure model. FALSE uses the values assigned by cutoffCensored.
remove50missing	
	only for summaryMethod = "TMP". TRUE removes the runs which have more than 50% missing values. FALSE is default.
fix_missing	Optional, same as the 'fix_missing' parameter in MSstatsConvert::MSstatsBalancedDesign function
maxQuantileforCensored	
	Maximum quantile for deciding censored missing values, default is 0.999
use_log_file	logical. If TRUE, information about data processing will be saved to a file.
append	logical. If TRUE, information about data processing will be added to an existing log file.
verbose	logical. If TRUE, information about data processing wil be printed to the console.

log_file_path   character. Path to a file to which information about data processing will be
                 saved. If not provided, such a file will be created automatically. If 'append =
                 TRUE', has to be a valid path to a file.

Examples

```
# Consider a raw data (i.e. SRMRawData) for a label-based SRM experiment from a yeast study
# with ten time points (T1-T10) of interests and three biological replicates.
# It is a time course experiment. The goal is to detect protein abundance changes
# across time points.
head(SRMRawData)
# Log2 transformation and normalization are applied (default)
QuantData<-dataProcess(SRMRawData, use_log_file = FALSE)
head(QuantData$FeatureLevelData)
# Log10 transformation and normalization are applied
QuantData1<-dataProcess(SRMRawData, logTrans=10, use_log_file = FALSE)
head(QuantData1$FeatureLevelData)
# Log2 transformation and no normalization are applied
QuantData2<-dataProcess(SRMRawData,normalization=FALSE, use_log_file = FALSE)
head(QuantData2$FeatureLevelData)
```

---

dataProcessPlots   *Visualization for explanatory data analysis*

Description

To illustrate the quantitative data after data-preprocessing and quality control of MS runs, dat-
aProcessPlots takes the quantitative data from function ([dataProcess](#)) as input and automatically
generate three types of figures in pdf files as output : (1) profile plot (specify "ProfilePlot" in option
type), to identify the potential sources of variation for each protein; (2) quality control plot (specify
"QCPlot" in option type), to evaluate the systematic bias between MS runs; (3) mean plot for con-
ditions (specify "ConditionPlot" in option type), to illustrate mean and variability of each condition
per protein.

Usage

```
dataProcessPlots(
  data,
  type,
  featureName = "Transition",
  ylimUp = FALSE,
  ylimDown = FALSE,
  scale = FALSE,
  interval = "CI",
  x.axis.size = 10,
  y.axis.size = 10,
  text.size = 4,
```

```
    text.angle = 0,
    legend.size = 7,
    dot.size.profile = 2,
    dot.size.condition = 3,
    width = 10,
    height = 10,
    which.Protein = "all",
    originalPlot = TRUE,
    summaryPlot = TRUE,
    save_condition_plot_result = FALSE,
    remove_uninformative_feature_outlier = FALSE,
    address = ""
)
```

Arguments

`data`	name of the (output of dataProcess function) data set.
`type`	choice of visualization. "ProfilePlot" represents profile plot of log intensities across MS runs. "QCPlot" represents quality control plot of log intensities across MS runs. "ConditionPlot" represents mean plot of log ratios (Light/Heavy) across conditions.
`featureName`	for "ProfilePlot" only, "Transition" (default) means printing feature legend in transition-level; "Peptide" means printing feature legend in peptide-level; "NA" means no feature legend printing.
`ylimUp`	upper limit for y-axis in the log scale. FALSE(Default) for Profile Plot and QC Plot use the upper limit as rounded off maximum of log2(intensities) after normalization + 3. FALSE(Default) for Condition Plot is maximum of log ratio + SD or CI.
`ylimDown`	lower limit for y-axis in the log scale. FALSE(Default) for Profile Plot and QC Plot is 0. FALSE(Default) for Condition Plot is minumum of log ratio - SD or CI.
`scale`	for "ConditionPlot" only, FALSE(default) means each conditional level is not scaled at x-axis according to its actual value (equal space at x-axis). TRUE means each conditional level is scaled at x-axis according to its actual value (unequal space at x-axis).
`interval`	for "ConditionPlot" only, "CI"(default) uses confidence interval with 0.95 significant level for the width of error bar. "SD" uses standard deviation for the width of error bar.
`x.axis.size`	size of x-axis labeling for "Run" in Profile Plot and QC Plot, and "Condition" in Condition Plot. Default is 10.
`y.axis.size`	size of y-axis labels. Default is 10.
`text.size`	size of labels represented each condition at the top of graph in Profile Plot and QC plot. Default is 4.
`text.angle`	angle of labels represented each condition at the top of graph in Profile Plot and QC plot or x-axis labeling in Condition plot. Default is 0.

legend.size	size of feature legend (transition-level or peptide-level) above graph in Profile Plot. Default is 7.
dot.size.profile	
	size of dots in profile plot. Default is 2.
dot.size.condition	
	size of dots in condition plot. Default is 3.
width	width of the saved file. Default is 10.
height	height of the saved file. Default is 10.
which.Protein	Protein list to draw plots. List can be names of Proteins or order numbers of Proteins from levels(data$FeatureLevelData$PROTEIN). Default is "all", which generates all plots for each protein. For QC plot, "allonly" will generate one QC plot with all proteins.
originalPlot	TRUE(default) draws original profile plots.
summaryPlot	TRUE(default) draws profile plots with summarization for run levels.
save_condition_plot_result	
	TRUE saves the table with values using condition plots. Default is FALSE.
remove_uninformative_feature_outlier	
	It only works after users used featureSubset="highQuality" in dataProcess. TRUE allows to remove 1) the features are flagged in the column, feature_quality="Uninformative" which are features with bad quality, 2) outliers that are flagged in the column, is_outlier=TRUE in Profile plots. FALSE (default) shows all features and intensities in profile plots.
address	the name of folder that will store the results. Default folder is the current working directory. The other assigned folder has to be existed under the current working directory. An output pdf file is automatically created with the default name of "ProfilePlot.pdf" or "QCplot.pdf" or "ConditionPlot.pdf" or "Condition-Plot_value.csv". The command address can help to specify where to store the file as well as how to modify the beginning of the file name. If address=FALSE, plot will be not saved as pdf file but showed in window.

Details

- Profile Plot : identify the potential sources of variation of each protein. QuantData$FeatureLevelData is used for plots. X-axis is run. Y-axis is log-intensities of transitions. Reference/endogenous signals are in the left/right panel. Line colors indicate peptides and line types indicate transitions. In summarization plots, gray dots and lines are the same as original profile plots with QuantData$FeatureLevelData. Dark dots and lines are for summarized intensities from QuantData$ProteinLevelData.

- QC Plot : illustrate the systematic bias between MS runs. After normalization, the reference signals for all proteins should be stable across MS runs. QuantData$FeatureLevelData is used for plots. X-axis is run. Y-axis is log-intensities of transition. Reference/endogenous signals are in the left/right panel. The pdf file contains (1) QC plot for all proteins and (2) QC plots for each protein separately.

- Condition Plot : illustrate the systematic difference between conditions. Summarized intensnties from QuantData$ProteinLevelData are used for plots. X-axis is condition. Y-axis is summarized log transformed intensity. If scale is TRUE, the levels of conditions is scaled

according to its actual values at x-axis. Red points indicate the mean for each condition. If interval is "CI", blue error bars indicate the confidence interval with 0.95 significant level for each condition. If interval is "SD", blue error bars indicate the standard deviation for each condition.The interval is not related with model-based analysis.

The input of this function is the quantitative data from function `dataProcess`.

Examples

```
# Consider quantitative data (i.e. QuantData) from a yeast study with ten time points of interests,
# three biological replicates, and no technical replicates which is a time-course experiment.
# The goal is to provide pre-analysis visualization by automatically generate two types of figures
# in two separate pdf files.
# Protein IDHC (gene name IDP2) is differentially expressed in time point 1 and time point 7,
# whereas, Protein PMG2 (gene name GPM2) is not.

QuantData<-dataProcess(SRMRawData, use_log_file = FALSE)
head(QuantData$FeatureLevelData)
# Profile plot
dataProcessPlots(data=QuantData,type="ProfilePlot")
# Quality control plot
dataProcessPlots(data=QuantData,type="QCPlot")
# Quantification plot for conditions
dataProcessPlots(data=QuantData,type="ConditionPlot")
```

DDARawData   *Example dataset from a label-free DDA, a controlled spike-in experiment.*

Description

This is a data set obtained from a published study (Mueller, et. al, 2007). A controlled spike-in experiment, where 6 proteins, (horse myoglobin, bovine carbonic anhydrase, horse Cytochrome C, chicken lysozyme, yeast alcohol dehydrogenase, rabbit aldolase A) were spiked into a complex background in known concentrations in a latin square design. The experiment contained 6 mixtures, and each mixture was analyzed in label-free LC-MS mode with 3 technical replicates (resulting in the total of 18 runs). Each protein was represented by 7-21 peptides, and each peptide was represented by 1-5 transition.

Usage

```
DDARawData
```

Format

data.frame

Details

The raw data (input data for MSstats) is required to contain variable of ProteinName, PeptideSequence, PrecursorCharge, FragmentIon, ProductCharge, IsotopeLabelType, Condition, BioReplicate, Run, Intensity. The variable names should be fixed.

If the information of one or more columns is not available for the original raw data, please retain the column variables and type in fixed value. For example, the original raw data does not contain the information of PrecursorCharge and ProductCharge, we retain the column PrecursorCharge and ProductCharge and then type in NA for all transitions in RawData.

Variable Intensity is required to be original signal without any log transformation and can be specified as the peak of height or the peak of area under curve.

Value

data.frame with the required format of MSstats.

Author(s)

Meena Choi, Olga Vitek.

Maintainer: Meena Choi (<mnchoi67@gmail.com>)

References

Meena Choi, Ching-Yun Chang, Timothy Clough, Daniel Broudy, Trevor Killeen, Brendan MacLean and Olga Vitek. "MSstats: an R package for statistical analysis of quantitative mass spectrometry-based proteomic experiments" *Bioinformatics*, 30(17):1514-1526, 2014.

Timothy Clough, Safia Thaminy, Susanne Ragg, Ruedi Aebersold, Olga Vitek. "Statistical protein quantification and significance analysis in label-free LC-M experiments with complex designs" *BMC Bioinformatics*, 13:S16, 2012.

Mueller, L. N., Rinner, O., Schmidt, A., Letarte, S., Bodenmiller, B., Brusniak, M., Vitek, O., Aebersold, R., and Muller, M. (2007). SuperHirn - a novel tool for high resolution LC-MS based peptide/protein profiling. Proteomics, 7, 3470-3480. 3, 34

Examples

```
head(DDARawData)
```

DDARawData.Skyline   *Example dataset from a label-free DDA, a controlled spike-in experiment, processed by Skyline.*

Description

This is a data set obtained from a published study (Mueller, et. al, 2007). A controlled spike-in experiment, where 6 proteins, (horse myoglobin, bovine carbonic anhydrase, horse Cytochrome C, chicken lysozyme, yeast alcohol dehydrogenase, rabbit aldolase A) were spiked into a complex background in known concentrations in a latin square design. The experiment contained 6 mixtures, and each mixture was analyzed in label-free LC-MS mode with 3 technical replicates (resulting in the total of 18 runs). Each protein was represented by 7-21 peptides, and each peptide was represented by 1-5 transition. Skyline is used for processing.

Usage

```
DDARawData.Skyline
```

Format

data.frame

Details

The raw data (input data for MSstats) is required to contain variable of ProteinName, PeptideSequence, PrecursorCharge, FragmentIon, ProductCharge, IsotopeLabelType, Condition, BioReplicate, Run, Intensity. The variable names should be fixed.

This is 'MSstats input' format from Skyline used by 'MSstats_report.skyr'. The column names, 'FileName' and 'Area', should be changed to 'Run' and 'Intensity'. There are two extra columns called 'StandardType' and 'Truncated'.'StandardType' column can be used for normalization='globalStandard' in `dataProcess`. 'Truncated' columns can be used to remove the truncated peaks with skylineReport=TRUE in `dataProcess`.

If the information of one or more columns is not available for the original raw data, please retain the column variables and type in fixed value. For example, the original raw data does not contain the information of PrecursorCharge and ProductCharge, we retain the column PrecursorCharge and ProductCharge and then type in NA for all transitions in RawData.

Variable Intensity is required to be original signal without any log transformation and can be specified as the peak of height or the peak of area under curve.

Value

data.frame with the required format of MSstats.

Author(s)

Meena Choi, Olga Vitek.

Maintainer: Meena Choi (<mnchoi67@gmail.com>)

References

Meena Choi, Ching-Yun Chang, Timothy Clough, Daniel Broudy, Trevor Killeen, Brendan MacLean and Olga Vitek. "MSstats: an R package for statistical analysis of quantitative mass spectrometry-based proteomic experiments" *Bioinformatics*, 30(17):1514-1526, 2014.

Timothy Clough, Safia Thaminy, Susanne Ragg, Ruedi Aebersold, Olga Vitek. "Statistical protein quantification and significance analysis in label-free LC-M experiments with complex designs" *BMC Bioinformatics*, 13:S16, 2012.

Examples

```
head(DDARawData.Skyline)
```

---

designSampleSize	*Planning future experimental designs of Selected Reaction Monitoring (SRM), Data-Dependent Acquisition (DDA or shotgun), and Data-Independent Acquisition (DIA or SWATH-MS) experiments in sample size calculation*

Description

Calculate sample size for future experiments of a Selected Reaction Monitoring (SRM), Data-Dependent Acquisition (DDA or shotgun), and Data-Independent Acquisition (DIA or SWATH-MS) experiment based on intensity-based linear model. Two options of the calculation: (1) number of biological replicates per condition, (2) power.

Usage

```
designSampleSize(
  data,
  desiredFC,
  FDR = 0.05,
  numSample = TRUE,
  power = 0.9,
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL
)
```

Arguments

data	'FittedModel' in testing output from function groupComparison.
desiredFC	the range of a desired fold change which includes the lower and upper values of the desired fold change.
FDR	a pre-specified false discovery ratio (FDR) to control the overall false positive rate. Default is 0.05
numSample	minimal number of biological replicates per condition. TRUE represents you require to calculate the sample size for this category, else you should input the exact number of biological replicates.

power	a pre-specified statistical power which defined as the probability of detecting a true fold change. TRUE represent you require to calculate the power for this category, else you should input the average of power you expect. Default is 0.9
use_log_file	logical. If TRUE, information about data processing will be saved to a file.
append	logical. If TRUE, information about data processing will be added to an existing log file.
verbose	logical. If TRUE, information about data processing wil be printed to the console.
log_file_path	character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If 'append = TRUE', has to be a valid path to a file.

Details

The function fits the model and uses variance components to calculate sample size. The underlying model fitting with intensity-based linear model with technical MS run replication. Estimated sample size is rounded to 0 decimal. The function can only obtain either one of the categories of the sample size calculation (numSample, numPep, numTran, power) at the same time.

Value

data.frame - sample size calculation results including varibles: desiredFC, numSample, FDR, and power.

Author(s)

Meena Choi, Ching-Yun Chang, Olga Vitek.

Examples

```
# Consider quantitative data (i.e. QuantData) from yeast study.
# A time course study with ten time points of interests and three biological replicates.
QuantData <- dataProcess(SRMRawData)
head(QuantData$FeatureLevelData)
## based on multiple comparisons  (T1 vs T3; T1 vs T7; T1 vs T9)
comparison1<-matrix(c(-1,0,1,0,0,0,0,0,0,0),nrow=1)
comparison2<-matrix(c(-1,0,0,0,0,0,1,0,0,0),nrow=1)
comparison3<-matrix(c(-1,0,0,0,0,0,0,0,1,0),nrow=1)
comparison<-rbind(comparison1,comparison2, comparison3)
row.names(comparison)<-c("T3-T1","T7-T1","T9-T1")
colnames(comparison)<-unique(QuantData$ProteinLevelData$GROUP)

testResultMultiComparisons<-groupComparison(contrast.matrix=comparison,data=QuantData)

## Calculate sample size for future experiments:
#(1) Minimal number of biological replicates per condition
designSampleSize(data=testResultMultiComparisons$FittedModel, numSample=TRUE,
                desiredFC=c(1.25,1.75), FDR=0.05, power=0.8)
#(2) Power calculation
designSampleSize(data=testResultMultiComparisons$FittedModel, numSample=2,
```

```
                    desiredFC=c(1.25,1.75), FDR=0.05, power=TRUE)
```

---

designSampleSizePlots   *Visualization for sample size calculation*

Description

To illustrate the relationship of desired fold change and the calculated minimal number sample size which are (1) number of biological replicates per condition, (2) number of peptides per protein, (3) number of transitions per peptide, and (4) power. The input is the result from function (designSampleSize.

Usage

```
designSampleSizePlots(data)
```

Arguments

data   output from function designSampleSize.

Details

Data in the example is based on the results of sample size calculation from function designSampleSize

Value

Plot for estimated sample size with assigned variable.

Author(s)

Meena Choi, Ching-Yun Chang, Olga Vitek.

Examples

```
# Based on the results of sample size calculation from function designSampleSize,
# we generate a series of sample size plots for number of biological replicates, or peptides,
# or transitions or power plot.
QuantData<-dataProcess(SRMRawData)
head(QuantData$ProcessedData)
## based on multiple comparisons  (T1 vs T3; T1 vs T7; T1 vs T9)
comparison1<-matrix(c(-1,0,1,0,0,0,0,0,0,0),nrow=1)
comparison2<-matrix(c(-1,0,0,0,0,0,1,0,0,0),nrow=1)
comparison3<-matrix(c(-1,0,0,0,0,0,0,0,1,0),nrow=1)
comparison<-rbind(comparison1,comparison2, comparison3)
row.names(comparison)<-c("T3-T1","T7-T1","T9-T1")
colnames(comparison)<-unique(QuantData$ProteinLevelData$GROUP)

testResultMultiComparisons<-groupComparison(contrast.matrix=comparison, data=QuantData)
```

```
# plot the calculated sample sizes for future experiments:
# (1) Minimal number of biological replicates per condition
result.sample<-designSampleSize(data=testResultMultiComparisons$FittedModel, numSample=TRUE,
                                desiredFC=c(1.25,1.75), FDR=0.05, power=0.8)
designSampleSizePlots(data=result.sample)
# (2) Power
result.power<-designSampleSize(data=testResultMultiComparisons$FittedModel, numSample=2,
                               desiredFC=c(1.25,1.75), FDR=0.05, power=TRUE)
designSampleSizePlots(data=result.power)
```

DIARawData	*Example dataset from a label-free DIA, a group comparison study of S.Pyogenes.*

Description

This example dataset was obtained from a group comparison study of S. Pyogenes. Two conditions, S. Pyogenes with 0% and 10% of human plasma added (denoted Strep 0% and Strep 10%), were profiled in two replicates, in the label-free mode, with a SWATH-MS-enabled AB SCIEX TripleTOF 5600 System. The identification and quantification of spectral peaks was assisted by a spectral library, and was performed using OpenSWATH software (http: //proteomics.ethz.ch/openswath.html). For reasons of space, the example dataset only contains two proteins from this study. Protein FabG shows strong evidence of differential abundance, while protein Probable RNA helicase exp9 only shows moderate evidence of dif- ferential abundance between conditions.

Usage

```
DIARawData
```

Format

data.frame

Details

The raw data (input data for MSstats) is required to contain variable of ProteinName, PeptideSequence, PrecursorCharge, FragmentIon, ProductCharge, IsotopeLabelType, Condition, BioReplicate, Run, Intensity. The variable names should be fixed.

If the information of one or more columns is not available for the original raw data, please retain the column variables and type in fixed value. For example, the original raw data does not contain the information of PrecursorCharge and ProductCharge, we retain the column PrecursorCharge and ProductCharge and then type in NA for all transitions in RawData.

Variable Intensity is required to be original signal without any log transformation and can be specified as the peak of height or the peak of area under curve.

Value

data.frame with the required format of MSstats.

Author(s)

Meena Choi, Olga Vitek.

Maintainer: Meena Choi (<mnchoi67@gmail.com>)

Examples

```
head(DIARawData)
```

DIAUmpiretoMSstatsFormat

Import DIA-Umpire files

Description

Import DIA-Umpire files

Usage

```
DIAUmpiretoMSstatsFormat(
  raw.frag,
  raw.pep,
  raw.pro,
  annotation,
  useSelectedFrag = TRUE,
  useSelectedPep = TRUE,
  removeFewMeasurements = TRUE,
  removeProtein_with1Feature = FALSE,
  summaryforMultipleRows = max,
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL,
  ...
)
```

Arguments

raw.frag	name of FragSummary_date.xls data, which includes feature-level data.
raw.pep	name of PeptideSummary_date.xls data, which includes selected fragments information.
raw.pro	name of ProteinSummary_date.xls data, which includes selected peptides information.

annotation   name of annotation data which includes Condition, BioReplicate, Run informa-
                   tion.

useSelectedFrag

                   TRUE will use the selected fragment for each peptide. 'Selected_fragments'
                   column is required.

useSelectedPep   TRUE will use the selected peptide for each protein. 'Selected_peptides' col-
                   umn is required.

removeFewMeasurements

                   TRUE (default) will remove the features that have 1 or 2 measurements across
                   runs.

removeProtein_with1Feature

                   TRUE will remove the proteins which have only 1 feature, which is the combi-
                   nation of peptide, precursor charge, fragment and charge. FALSE is default.

summaryforMultipleRows

                   max(default) or sum - when there are multiple measurements for certain feature
                   and certain run, use highest or sum of multiple intensities.

use_log_file   logical. If TRUE, information about data processing will be saved to a file.

append   logical. If TRUE, information about data processing will be added to an existing
                   log file.

verbose   logical. If TRUE, information about data processing wil be printed to the con-
                   sole.

log_file_path   character. Path to a file to which information about data processing will be
                   saved. If not provided, such a file will be created automatically. If 'append =
                   TRUE', has to be a valid path to a file.

...   additional parameters to 'data.table::fread'.

Value

data.frame in the MSstats required format.

Author(s)

Meena Choi, Olga Vitek

Examples

```
diau_frag = system.file("tinytest/raw_data/DIAUmpire/dia_frag.csv",
                        package = "MSstatsConvert")
diau_pept = system.file("tinytest/raw_data/DIAUmpire/dia_pept.csv",
                        package = "MSstatsConvert")
diau_prot = system.file("tinytest/raw_data/DIAUmpire/dia_prot.csv",
                        package = "MSstatsConvert")
annot = system.file("tinytest/annotations/annot_diau.csv",
                    package = "MSstats")
diau_frag = data.table::fread(diau_frag)
diau_pept = data.table::fread(diau_pept)
diau_prot = data.table::fread(diau_prot)
annot = data.table::fread(annot)
```

```
diau_frag = diau_frag[, lapply(.SD, function(x) if (is.integer(x)) as.numeric(x) else x)]
# In case numeric columns are not interpreted correctly

diau_imported = DIAUmpiretoMSstatsFormat(diau_frag, diau_pept, diau_prot,
                                         annot, use_log_file = FALSE)
head(diau_imported)
```

---

getProcessed   *Get feature-level data to be used in the MSstatsSummarizationOutput*
                             function

Description

Get feature-level data to be used in the MSstatsSummarizationOutput function

Usage

```
getProcessed(input)
```

Arguments

input   data.table processed by dataProcess subfunctions

Value

data.table processed by dataProcess subfunctions

Examples

```
raw = DDARawData
method = "TMP"
cens = "NA"
impute = TRUE
MSstatsConvert::MSstatsLogsSettings(FALSE)
input = MSstatsPrepareForDataProcess(raw, 2, NULL)
input = MSstatsNormalize(input, "EQUALIZEMEDIANS")
input = MSstatsMergeFractions(input)
input = MSstatsHandleMissing(input, "TMP", TRUE, "NA", 0.999)
input_all = MSstatsSelectFeatures(input, "all") # all features
input_5 = MSstatsSelectFeatures(data.table::copy(input),
"topN", top_n = 5) # top 5 features

proc1 = getProcessed(input_all)
proc2 = getProcessed(input_5)

proc1
proc2
```

---

getSamplesInfo   *Get information about number of measurements for each group*

Description

Get information about number of measurements for each group

Usage

```
getSamplesInfo(summarization_output)
```

Arguments

summarization_output

output of the dataProcess function

Details

This function extracts information required to compute percentages of missing and imputed values in group comparison.

Value

data.table

Examples

```
QuantData <- dataProcess(DDARawData, use_log_file = FALSE)
samples_info <- getSamplesInfo(QuantData)
samples_info
```

---

getSelectedProteins   *Get proteins based on names or integer IDs*

Description

Get proteins based on names or integer IDs

Usage

```
getSelectedProteins(chosen_proteins, all_proteins)
```

Arguments

chosen_proteins

protein names or integers IDs

all_proteins   all unique proteins

Value

character

---

groupComparison   *Whole plot testing*

Description

Whole plot testing

Usage

```
groupComparison(
  contrast.matrix,
  data,
  save_fitted_models = TRUE,
  log_base = 2,
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL
)
```

Arguments

contrast.matrix

               comparison between conditions of interests.

data   name of the (output of dataProcess function) data set.

save_fitted_models

               logical, if TRUE, fitted models will be added to the output.

log_base   base of the logarithm used in dataProcess.

use_log_file logical. If TRUE, information about data processing will be saved to a file.

append   logical. If TRUE, information about data processing will be added to an existing log file.

verbose   logical. If TRUE, information about data processing wil be printed to the console.

log_file_path character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If 'append = TRUE', has to be a valid path to a file.

Details

contrast.matrix : comparison of interest. Based on the levels of conditions, specify 1 or -1 to the conditions of interests and 0 otherwise. The levels of conditions are sorted alphabetically. Command levels(QuantData$FeatureLevelData$GROUP_ORIGINAL) can illustrate the actual order of the levels of conditions. The underlying model fitting functions are lm and lmer for the fixed effects model and mixed effects model, respectively. The input of this function is the quantitative data from function (dataProcess).

Value

list that consists of three elements: "ComparisonResult" - data.frame with results of statistical testing, "ModelQC" - data.frame with data used to fit models for group comparison and "FittedModel" - list of fitted models.

Examples

```
# Consider quantitative data (i.e. QuantData) from yeast study with ten time points of interests,
# three biological replicates, and no technical replicates.
# It is a time-course experiment and we attempt to compare differential abundance
# between time 1 and 7 in a set of targeted proteins.
# In this label-based SRM experiment, MSstats uses the fitted model with expanded scope of
# Biological replication.
QuantData <- dataProcess(SRMRawData, use_log_file = FALSE)
head(QuantData$FeatureLevelData)
levels(QuantData$ProteinLevelData$GROUP)
comparison <- matrix(c(-1,0,0,0,0,0,1,0,0,0),nrow=1)
row.names(comparison) <- "T7-T1"
groups = levels(QuantData$ProteinLevelData$GROUP)
colnames(comparison) <- groups[order(as.numeric(groups))]
# Tests for differentially abundant proteins with models:
# label-based SRM experiment with expanded scope of biological replication.
testResultOneComparison <- groupComparison(contrast.matrix=comparison, data=QuantData,
                                            use_log_file = FALSE)
# table for result
testResultOneComparison$ComparisonResult
```

---

groupComparisonPlots   *Visualization for model-based analysis and summarizing differentially abundant proteins*

Description

To summarize the results of log-fold changes and adjusted p-values for differentially abundant proteins, groupComparisonPlots takes testing results from function ([groupComparison](#)) as input and automatically generate three types of figures in pdf files as output : (1) volcano plot (specify "VolcanoPlot" in option type) for each comparison separately; (2) heatmap (specify "Heatmap" in option type) for multiple comparisons ; (3) comparison plot (specify "ComparisonPlot" in option type) for multiple comparisons per protein.

Usage

```
groupComparisonPlots(
  data,
  type,
  sig = 0.05,
  FCcutoff = FALSE,
  logBase.pvalue = 10,
  ylimUp = FALSE,
  ylimDown = FALSE,
  xlimUp = FALSE,
  x.axis.size = 10,
  y.axis.size = 10,
  dot.size = 3,
  text.size = 4,
  text.angle = 0,
  legend.size = 13,
  ProteinName = TRUE,
  colorkey = TRUE,
  numProtein = 100,
  clustering = "both",
  width = 10,
  height = 10,
  which.Comparison = "all",
  which.Protein = "all",
  address = ""
)
```

Arguments

data	'ComparisonResult' in testing output from function groupComparison.
type	choice of visualization. "VolcanoPlot" represents volcano plot of log fold changes and adjusted p-values for each comparison separately. "Heatmap" represents heatmap of adjusted p-values for multiple comparisons. "ComparisonPlot" represents comparison plot of log fold changes for multiple comparisons per protein.
sig	FDR cutoff for the adjusted p-values in heatmap and volcano plot. level of significance for comparison plot. 100(1-sig)% confidence interval will be drawn. sig=0.05 is default.
FCcutoff	for volcano plot or heatmap, whether involve fold change cutoff or not. FALSE (default) means no fold change cutoff is applied for significance analysis. FCcutoff = specific value means specific fold change cutoff is applied.
logBase.pvalue	for volcano plot or heatmap, (-) logarithm transformation of adjusted p-value with base 2 or 10(default).
ylimUp	for all three plots, upper limit for y-axis. FALSE (default) for volcano plot/heatmap use maximum of -log2 (adjusted p-value) or -log10 (adjusted p-value). FALSE (default) for comparison plot uses maximum of log-fold change + CI.

ylimDown	for all three plots, lower limit for y-axis. FALSE (default) for volcano plot/heatmap use minimum of -log2 (adjusted p-value) or -log10 (adjusted p-value). FALSE (default) for comparison plot uses minimum of log-fold change - CI.
xlimUp	for Volcano plot, the limit for x-axis. FALSE (default) for use maximum for absolute value of log-fold change or 3 as default if maximum for absolute value of log-fold change is less than 3.
x.axis.size	size of axes labels, e.g. name of the comparisons in heatmap, and in comparison plot. Default is 10.
y.axis.size	size of axes labels, e.g. name of targeted proteins in heatmap. Default is 10.
dot.size	size of dots in volcano plot and comparison plot. Default is 3.
text.size	size of ProteinName label in the graph for Volcano Plot. Default is 4.
text.angle	angle of x-axis labels represented each comparison at the bottom of graph in comparison plot. Default is 0.
legend.size	size of legend for color at the bottom of volcano plot. Default is 7.
ProteinName	for volcano plot only, whether display protein names or not. TRUE (default) means protein names, which are significant, are displayed next to the points. FALSE means no protein names are displayed.
colorkey	TRUE(default) shows colorkey.
numProtein	The number of proteins which will be presented in each heatmap. Default is 100. Maximum possible number of protein for one heatmap is 180.
clustering	Determines how to order proteins and comparisons. Hierarchical cluster analysis with Ward method(minimum variance) is performed. 'protein' means that protein dendrogram is computed and reordered based on protein means (the order of row is changed). 'comparison' means comparison dendrogram is computed and reordered based on comparison means (the order of comparison is changed). 'both' means to reorder both protein and comparison. Default is 'protein'.
width	width of the saved file. Default is 10.
height	height of the saved file. Default is 10.
which.Comparison	
	list of comparisons to draw plots. List can be labels of comparisons or order numbers of comparisons from levels(data$Label), such as levels(testResultMultiComparisons$Compariso Default is "all", which generates all plots for each protein.
which.Protein	Protein list to draw comparison plots. List can be names of Proteins or order numbers of Proteins from levels(testResultMultiComparisons$ComparisonResult$Protein). Default is "all", which generates all comparison plots for each protein.
address	the name of folder that will store the results. Default folder is the current working directory. The other assigned folder has to be existed under the current working directory. An output pdf file is automatically created with the default name of "VolcanoPlot.pdf" or "Heatmap.pdf" or "ComparisonPlot.pdf". The command address can help to specify where to store the file as well as how to modify the beginning of the file name. If address=FALSE, plot will be not saved as pdf file but showed in window.

Details

- Volcano plot : illustrate actual log-fold changes and adjusted p-values for each comparison separately with all proteins. The x-axis is the log fold change. The base of logarithm transformation is the same as specified in "logTrans" from [dataProcess](). The y-axis is the negative log2 or log10 adjusted p-values. The horizontal dashed line represents the FDR cutoff. The points below the FDR cutoff line are non-significantly abundant proteins (colored in black). The points above the FDR cutoff line are significantly abundant proteins (colored in red/blue for up-/down-regulated). If fold change cutoff is specified (FCcutoff = specific value), the points above the FDR cutoff line but within the FC cutoff line are non-significantly abundant proteins (colored in black)/

- Heatmap : illustrate up-/down-regulated proteins for multiple comparisons with all proteins. Each column represents each comparison of interest. Each row represents each protein. Color red/blue represents proteins in that specific comparison are significantly up-regulated/down-regulated proteins with FDR cutoff and/or FC cutoff. The color scheme shows the evidences of significance. The darker color it is, the stronger evidence of significance it has. Color gold represents proteins are not significantly different in abundance.

- Comparison plot : illustrate log-fold change and its variation of multiple comparisons for single protein. X-axis is comparison of interest. Y-axis is the log fold change. The red points are the estimated log fold change from the model. The blue error bars are the confidence interval with 0.95 significant level for log fold change. This interval is only based on the standard error, which is estimated from the model.

Examples

```
QuantData<-dataProcess(SRMRawData, use_log_file = FALSE)
head(QuantData$FeatureLevelData)
## based on multiple comparisons  (T1 vs T3; T1 vs T7; T1 vs T9)
comparison1<-matrix(c(-1,0,1,0,0,0,0,0,0,0),nrow=1)
comparison2<-matrix(c(-1,0,0,0,0,0,1,0,0,0),nrow=1)
comparison3<-matrix(c(-1,0,0,0,0,0,0,0,1,0),nrow=1)
comparison<-rbind(comparison1,comparison2, comparison3)
row.names(comparison)<-c("T3-T1","T7-T1","T9-T1")
groups = levels(QuantData$ProteinLevelData$GROUP)
colnames(comparison) <- groups[order(as.numeric(groups))]
testResultMultiComparisons<-groupComparison(contrast.matrix=comparison,
data=QuantData,
use_log_file = FALSE)
testResultMultiComparisons$ComparisonResult
# Volcano plot with FDR cutoff = 0.05 and no FC cutoff
groupComparisonPlots(data=testResultMultiComparisons$ComparisonResult, type="VolcanoPlot",
logBase.pvalue=2, address="Ex1_")
# Volcano plot with FDR cutoff = 0.05, FC cutoff = 70, upper y-axis limit = 100,
# and no protein name displayed
# FCcutoff=70 is for demonstration purpose
groupComparisonPlots(data=testResultMultiComparisons$ComparisonResult, type="VolcanoPlot",
FCcutoff=70, logBase.pvalue=2, ylimUp=100, ProteinName=FALSE,address="Ex2_")
# Heatmap with FDR cutoff = 0.05
groupComparisonPlots(data=testResultMultiComparisons$ComparisonResult, type="Heatmap",
logBase.pvalue=2, address="Ex1_")
# Heatmap with FDR cutoff = 0.05 and FC cutoff = 70
```

```
# FCcutoff=70 is for demonstration purpose
groupComparisonPlots(data=testResultMultiComparisons$ComparisonResult, type="Heatmap",
FCcutoff=70, logBase.pvalue=2, address="Ex2_")
# Comparison Plot
groupComparisonPlots(data=testResultMultiComparisons$ComparisonResult, type="ComparisonPlot",
address="Ex1_")
# Comparison Plot
groupComparisonPlots(data=testResultMultiComparisons$ComparisonResult, type="ComparisonPlot",
ylimUp=8, ylimDown=-1, address="Ex2_")
```

---

makePeptidesDictionary

Prepare a peptides dictionary for global standards normalization

---

Description

Prepare a peptides dictionary for global standards normalization

Usage

```
makePeptidesDictionary(input, normalization)
```

Arguments

input   'data.table' in MSstats standard format

normalization normalization method

Details

This function extracts information required to perform normalization with global standards. It is useful for running the summarization workflow outside of the dataProcess function.

Examples

```
input = data.table::as.data.table(DDARawData)
peptides_dict = makePeptidesDictionary(input, "GLOBALSTANDARDS")
head(peptides_dict) # ready to be passed to the MSstatsNormalize function
```

MaxQtoMSstatsFormat   *Import MaxQuant files*

Description

Import MaxQuant files

Usage

```
MaxQtoMSstatsFormat(
  evidence,
  annotation,
  proteinGroups,
  proteinID = "Proteins",
  useUniquePeptide = TRUE,
  summaryforMultipleRows = max,
  removeFewMeasurements = TRUE,
  removeMpeptides = FALSE,
  removeOxidationMpeptides = FALSE,
  removeProtein_with1Peptide = FALSE,
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL,
  ...
)
```

Arguments

evidence	name of 'evidence.txt' data, which includes feature-level data.
annotation	name of 'annotation.txt' data which includes Raw.file, Condition, BioReplicate, Run, IsotopeLabelType information.
proteinGroups	name of 'proteinGroups.txt' data. It needs to matching protein group ID. If proteinGroups=NULL, use 'Proteins' column in 'evidence.txt'.
proteinID	'Proteins'(default) or 'Leading.razor.protein' for Protein ID.
useUniquePeptide	
	TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.
summaryforMultipleRows	
	max(default) or sum - when there are multiple measurements for certain feature and certain run, use highest or sum of multiple intensities.
removeFewMeasurements	
	TRUE (default) will remove the features that have 1 or 2 measurements across runs.
removeMpeptides	
	TRUE will remove the peptides including 'M' sequence. FALSE is default.

removeOxidationMpeptides

> TRUE will remove the peptides including 'oxidation (M)' in modification. FALSE is default.

removeProtein_with1Peptide

> TRUE will remove the proteins which have only 1 peptide and charge. FALSE is default.

use_log_file   logical. If TRUE, information about data processing will be saved to a file.

append   logical. If TRUE, information about data processing will be added to an existing log file.

verbose   logical. If TRUE, information about data processing wil be printed to the console.

log_file_path   character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If 'append = TRUE', has to be a valid path to a file.

...   additional parameters to 'data.table::fread'.

Value

data.frame in the MSstats required format.

Note

Warning: MSstats does not support for metabolic labeling or iTRAQ experiments.

Author(s)

Meena Choi, Olga Vitek.

Examples

```
mq_ev = data.table::fread(system.file("tinytest/raw_data/MaxQuant/mq_ev.csv",
                                      package = "MSstatsConvert"))
mq_pg = data.table::fread(system.file("tinytest/raw_data/MaxQuant/mq_pg.csv",
                                      package = "MSstatsConvert"))
annot = data.table::fread(system.file("tinytest/raw_data/MaxQuant/annotation.csv",
                                      package = "MSstatsConvert"))
maxq_imported = MaxQtoMSstatsFormat(mq_ev, annot, mq_pg, use_log_file = FALSE)
head(maxq_imported)
```

---

modelBasedQCPlots   *Visualization for model-based quality control in fitting model*

Description

To check the assumption of linear model for whole plot inference, modelBasedQCPlots takes the results after fitting models from function ([groupComparison](#)) as input and automatically generate two types of figures in pdf files as output: (1) normal quantile-quantile plot (specify "QQPlot" in option type) for checking normally distributed errors.; (2) residual plot (specify "ResidualPlot" in option type).

Usage

```
modelBasedQCPlots(
  data,
  type,
  axis.size = 10,
  dot.size = 3,
  width = 10,
  height = 10,
  which.Protein = "all",
  address = ""
)
```

Arguments

data	output from function groupComparison.
type	choice of visualization. "QQPlots" represents normal quantile-quantile plot for each protein after fitting models. "ResidualPlots" represents a plot of residuals versus fitted values for each protein in the dataset.
axis.size	size of axes labels. Default is 10.
dot.size	size of points in the graph for residual plots and QQ plots. Default is 3.
width	width of the saved file. Default is 10.
height	height of the saved file. Default is 10.
which.Protein	Protein list to draw plots. List can be names of Proteins or order numbers of Proteins from levels(testResultOneComparison$ComparisonResult$Protein). Default is "all", which generates all plots for each protein.
address	name that will serve as a prefix to the name of output file.

Details

Results based on statistical models for whole plot level inference are accurate as long as the assumptions of the model are met. The model assumes that the measurement errors are normally distributed with mean 0 and constant variance. The assumption of a constant variance can be checked by examining the residuals from the model.

- QQPlots : a normal quantile-quantile plot for each protein is generated in order to check whether the errors are well approximated by a normal distribution. If points fall approximately along a straight line, then the assumption is appropriate for that protein. Only large deviations from the line are problematic.
- ResidualPlots : The plots of residuals against predicted(fitted) values. If it shows a random scatter, then the assumption is appropriate.

Value

produce a pdf file

Examples

```
QuantData <- dataProcess(SRMRawData, use_log_file = FALSE)
head(QuantData$FeatureLevelData)
levels(QuantData$FeatureLevelData$GROUP)
comparison <- matrix(c(-1,0,0,0,0,0,1,0,0,0),nrow=1)
row.names(comparison) <- "T7-T1"
colnames(comparison) <- unique(QuantData$ProteinLevelData$GROUP)
# Tests for differentially abundant proteins with models:
# label-based SRM experiment with expanded scope of biological replication.
testResultOneComparison <- groupComparison(contrast.matrix=comparison, data=QuantData,
use_log_file = FALSE)
# normal quantile-quantile plots
modelBasedQCPlots(data=testResultOneComparison, type="QQPlots", address="")
# residual plots
modelBasedQCPlots(data=testResultOneComparison, type="ResidualPlots", address="")
```

MSstatsContrastMatrix   *Create a contrast matrix for groupComparison function*

Description

Create a contrast matrix for groupComparison function

Usage

```
MSstatsContrastMatrix(contrasts, conditions, labels = NULL)
```

Arguments

contrasts   One of the following: i) list of lists. Each sub-list consists of two vectors that name conditions that will be compared. See the details section for more information ii) matrix. In this case, it's correctness will be checked iii) "pairwise". In this case, pairwise comparison matrix will be generated iv) data.frame. In this case, input will be converted to matrix

conditions   unique condition labels

labels   labels for contrasts (row.names of the contrast matrix)

MSstatsGroupComparison

Group comparison

Description

Group comparison

Usage

```
MSstatsGroupComparison(
  summarized_list,
  contrast_matrix,
  save_fitted_models,
  repeated,
  samples_info
)
```

Arguments

summarized_list

output of MSstatsPrepareForGroupComparison

contrast_matrix

contrast matrix

save_fitted_models

if TRUE, fitted models will be included in the output

repeated   logical, output of checkRepeatedDesign function

samples_info   data.table, output of getSamplesInfo function

Examples

```
QuantData <- dataProcess(SRMRawData, use_log_file = FALSE)
group_comparison_input = MSstatsPrepareForGroupComparison(QuantData)
levels(QuantData$ProteinLevelData$GROUP)
comparison <- matrix(c(-1,0,0,0,0,0,1,0,0,0),nrow=1)
row.names(comparison) <- "T7-T1"
groups = levels(QuantData$ProteinLevelData$GROUP)
colnames(comparison) <- groups[order(as.numeric(groups))]
samples_info = getSamplesInfo(QuantData)
repeated = checkRepeatedDesign(QuantData)
group_comparison = MSstatsGroupComparison(group_comparison_input, comparison,
                                          FALSE, repeated, samples_info)
length(group_comparison) # list of length equal to number of proteins
group_comparison[[1]][[1]] # data used to fit linear model
group_comparison[[1]][[2]] # comparison result
group_comparison[[2]][[3]] # NULL, because we set save_fitted_models to FALSE
```

MSstatsGroupComparisonOutput

Create output of group comparison based on results for individual proteins

Description

Create output of group comparison based on results for individual proteins

Usage

```
MSstatsGroupComparisonOutput(input, summarization_output, log_base = 2)
```

Arguments

input   output of MSstatsGroupComparison function

summarization_output
                      output of dataProcess function

log_base   base of the logarithm used in fold-change calculation

Value

list, same as the output of 'groupComparison'

Examples

```
QuantData <- dataProcess(SRMRawData, use_log_file = FALSE)
group_comparison_input = MSstatsPrepareForGroupComparison(QuantData)
levels(QuantData$ProteinLevelData$GROUP)
comparison <- matrix(c(-1,0,0,0,0,0,1,0,0,0),nrow=1)
row.names(comparison) <- "T7-T1"
groups = levels(QuantData$ProteinLevelData$GROUP)
colnames(comparison) <- groups[order(as.numeric(groups))]
samples_info = getSamplesInfo(QuantData)
repeated = checkRepeatedDesign(QuantData)
group_comparison = MSstatsGroupComparison(group_comparison_input, comparison,
                                          FALSE, repeated, samples_info)
group_comparison_final = MSstatsGroupComparisonOutput(group_comparison,
                                                      QuantData)
group_comparison_final[["ComparisonResult"]]
```

MSstatsGroupComparisonSingleProtein

Group comparison for a single protein

Description

Group comparison for a single protein

Usage

```
MSstatsGroupComparisonSingleProtein(
  single_protein,
  contrast_matrix,
  repeated,
  groups,
  samples_info,
  save_fitted_models,
  has_imputed
)
```

Arguments

single_protein	data.table with summarized data for a single protein
contrast_matrix	
	contrast matrix
repeated	if TRUE, repeated measurements will be modeled
groups	unique labels of experimental conditions
samples_info	number of runs per group
save_fitted_models	
	if TRUE, fitted model will be saved. If not, it will be replaced with NULL
has_imputed	TRUE if missing values have been imputed

Examples

```
QuantData <- dataProcess(SRMRawData, use_log_file = FALSE)
group_comparison_input <- MSstatsPrepareForGroupComparison(QuantData)
levels(QuantData$ProteinLevelData$GROUP)
comparison <- matrix(c(-1,0,0,0,0,0,1,0,0,0),nrow=1)
row.names(comparison) <- "T7-T1"
groups = levels(QuantData$ProteinLevelData$GROUP)
colnames(comparison) <- groups[order(as.numeric(groups))]
samples_info <- getSamplesInfo(QuantData)
repeated <- checkRepeatedDesign(QuantData)
single_output <- MSstatsGroupComparisonSingleProtein(
  group_comparison_input[[1]], comparison, repeated, groups, samples_info,
  FALSE, TRUE)
single_output # same as a single element of MSstatsGroupComparison output
```

MSstatsHandleMissing   *Handle censored missing values*

Description

Handle censored missing values

Usage

```
MSstatsHandleMissing(
  input,
  summary_method,
  impute,
  missing_symbol,
  censored_cutoff
)
```

Arguments

input	'data.table' in MSstats data format
summary_method	summarization method ('summaryMethod' parameter to 'dataProcess')
impute	if TRUE, missing values are supposed to be imputed ('MBimpute' parameter to 'dataProcess')
missing_symbol	'censoredInt' parameter to 'dataProcess'
censored_cutoff	
	'maxQuantileforCensored' parameter to 'dataProcess'

Value

data.table

Examples

```
raw = DDARawData
method = "TMP"
cens = "NA"
impute = TRUE
MSstatsConvert::MSstatsLogsSettings(FALSE)
input = MSstatsPrepareForDataProcess(raw, 2, NULL)
input = MSstatsNormalize(input, "EQUALIZEMEDIANS")
input = MSstatsMergeFractions(input)
input = MSstatsHandleMissing(input, "TMP", TRUE, "NA", 0.999)
head(input)
```

MSstatsMergeFractions   *Re-format the data before feature selection*

Description

Re-format the data before feature selection

Usage

```
MSstatsMergeFractions(input)
```

Arguments

input   'data.table' in MSstats format

Value

data.table

Examples

```
raw = DDARawData
method = "TMP"
cens = "NA"
impute = TRUE
MSstatsConvert::MSstatsLogsSettings(FALSE)
input = MSstatsPrepareForDataProcess(raw, 2, NULL)
input = MSstatsNormalize(input, "EQUALIZEMEDIANS")
input = MSstatsMergeFractions(input)
head(input)
```

MSstatsNormalize   *Normalize MS data*

Description

Normalize MS data

Usage

```
MSstatsNormalize(
  input,
  normalization_method,
  peptides_dict = NULL,
  standards = NULL
)
```

Arguments

input	data.table in MSstats format
normalization_method	
	name of a chosen normalization method: "NONE" or "FALSE" for no normalization, "EQUALIZEMEDIANS" for median normalization, "QUANTILE" normalization for quantile normalization from 'preprocessCore' package, "GLOBALSTANDARDS" for normalization based on selected peptides or proteins.
peptides_dict	'data.table' of names of peptides and their corresponding features.
standards	character vector with names of standards, required if "GLOBALSTANDARDS" method was selected.

Value

data.table

Examples

```
raw = DDARawData
method = "TMP"
cens = "NA"
impute = TRUE
MSstatsConvert::MSstatsLogsSettings(FALSE)
input = MSstatsPrepareForDataProcess(raw, 2, NULL)
input = MSstatsNormalize(input, "EQUALIZEMEDIANS") # median normalization
head(input)
```

MSstatsPrepareForDataProcess
Prepare data for processing by 'dataProcess' function

Description

Prepare data for processing by 'dataProcess' function

Usage

```
MSstatsPrepareForDataProcess(input, log_base, fix_missing)
```

Arguments

input	'data.table' in MSstats format
log_base	base of the logarithm to transform intensities
fix_missing	str, optional. Defaults to NULL, which means no action. If not NULL, must be one of the options: "zero_to_na" or "na_to_zero". If "zero_to_na", Intensity values equal exactly to 0 will be converted to NA. If "na_to_zero", missing values will be replaced by zeros.

Value

data.table

Examples

```
raw = DDARawData
method = "TMP"
cens = "NA"
impute = TRUE
MSstatsConvert::MSstatsLogsSettings(FALSE)
input = MSstatsPrepareForDataProcess(raw, 2, NULL)
head(input)
```

MSstatsPrepareForGroupComparison

Prepare output for dataProcess for group comparison

Description

Prepare output for dataProcess for group comparison

Usage

```
MSstatsPrepareForGroupComparison(summarization_output)
```

Arguments

summarization_output

output of dataProcess

Value

list of run-level data for each protein in the input. This list has a "has_imputed" attribute that indicates if missing values were imputed in the input dataset.

Examples

```
QuantData <- dataProcess(SRMRawData, use_log_file = FALSE)
group_comparison_input = MSstatsPrepareForGroupComparison(QuantData)
length(group_comparison_input) # list of length equal to number of proteins
# in protein-level data of QuantData
head(group_comparison_input[[1]])
```

```
MSstatsPrepareForSummarization
```
Prepare feature-level data for protein-level summarization

Description

Prepare feature-level data for protein-level summarization

Usage

```
MSstatsPrepareForSummarization(
  input,
  method,
  impute,
  censored_symbol,
  remove_uninformative_feature_outlier
)
```

Arguments

input	feature-level data processed by dataProcess subfunctions
method	summarization method - 'summaryMethod' parameter of the dataProcess function
impute	if TRUE, censored missing values will be imputed - 'MBimpute' parameter of the dataProcess function
censored_symbol	
	censored missing value indicator - 'censoredInt' parameter of the dataProcess function
remove_uninformative_feature_outlier	
	if TRUE, features labeled as outlier of uninformative by the MSstatsSelectFeatures function will not be used in summarization

Value

data.table

Examples

```
raw = DDARawData
method = "TMP"
cens = "NA"
impute = TRUE
MSstatsConvert::MSstatsLogsSettings(FALSE)
input = MSstatsPrepareForDataProcess(raw, 2, NULL)
head(input)
```

MSstatsSelectFeatures   *Feature selection before feature-level data summarization*

Description

Feature selection before feature-level data summarization

Usage

```
MSstatsSelectFeatures(input, method, top_n = 3, min_feature_count = 2)
```

Arguments

input   data.table

method   "all" / "highQuality", "topN"

top_n   number of features to use for "topN" method

min_feature_count
                  number of quality features for "highQuality" method

Value

data.table

Examples

```
raw = DDARawData
method = "TMP"
cens = "NA"
impute = TRUE
MSstatsConvert::MSstatsLogsSettings(FALSE)
input = MSstatsPrepareForDataProcess(raw, 2, NULL)
input = MSstatsNormalize(input, "EQUALIZEMEDIANS")
input = MSstatsMergeFractions(input)
input = MSstatsHandleMissing(input, "TMP", TRUE, "NA", 0.999)
input_all = MSstatsSelectFeatures(input, "all") # all features
input_5 = MSstatsSelectFeatures(data.table::copy(input), "topN", top_n = 5) # top 5 features
input_informative = MSstatsSelectFeatures(input, "highQuality") # feature selection

head(input_all)
head(input_5)
head(input_informative)
```

```
MSstatsSummarizationOutput
```
Post-processing output from MSstats summarization

Description

Post-processing output from MSstats summarization

Usage

```
MSstatsSummarizationOutput(
  input,
  summarized,
  processed,
  method,
  impute,
  censored_symbol
)
```

Arguments

input	'data.table' in MSstats format
summarized	output of the 'MSstatsSummarize' function
processed	output of MSstatsSelectFeatures
method	name of the summarization method ('summaryMethod' parameter to 'dataProcess')
impute	if TRUE, censored missing values were imputed ('MBimpute' parameter to 'dataProcess')
censored_symbol	censored missing value indicator ('censoredInt' parameter to 'dataProcess')

Value

list that consists of the following elements:

- FeatureLevelData - feature-level data after processing
- ProteinLevelData - protein-level (summarized) data
- SummaryMethod (string) - name of summarization method that was used

Examples

```
raw = DDARawData
method = "TMP"
cens = "NA"
impute = TRUE
MSstatsConvert::MSstatsLogsSettings(FALSE)
```

```
input = MSstatsPrepareForDataProcess(raw, 2, NULL)
input = MSstatsNormalize(input, "EQUALIZEMEDIANS")
input = MSstatsMergeFractions(input)
input = MSstatsHandleMissing(input, "TMP", TRUE, "NA", 0.999)
input = MSstatsSelectFeatures(input, "all")
processed = getProcessed(input)
input = MSstatsPrepareForSummarization(input, method, impute, cens, FALSE)
input_split = split(input, input$PROTEIN)
summarized = MSstatsSummarize(input_split, method, impute, cens, FALSE, TRUE)
output = output = MSstatsSummarizationOutput(input, summarized, processed,
method, impute, cens)
```

MSstatsSummarize   *Feature-level data summarization*

Description

Feature-level data summarization

Usage

```
MSstatsSummarize(
  proteins_list,
  method,
  impute,
  censored_symbol,
  remove50missing,
  equal_variance
)
```

Arguments

proteins_list   list of processed feature-level data

method   summarization method: "linear" or "TMP"

impute   only for summaryMethod = "TMP" and censoredInt = 'NA' or '0'. TRUE (default) imputes 'NA' or '0' (depending on censoredInt option) by Accelated failure model. FALSE uses the values assigned by cutoffCensored

censored_symbol

                Missing values are censored or at random. 'NA' (default) assumes that all 'NA's in 'Intensity' column are censored. '0' uses zero intensities as censored intensity. In this case, NA intensities are missing at random. The output from Skyline should use '0'. Null assumes that all NA intensites are randomly missing.

remove50missing

                only for summaryMethod = "TMP". TRUE removes the runs which have more than 50% missing values. FALSE is default.

equal_variance only for summaryMethod = "linear". Default is TRUE. Logical variable for whether the model should account for heterogeneous variation among intensities from different features. Default is TRUE, which assume equal variance among intensities from features. FALSE means that we cannot assume equal variance among intensities from features, then we will account for heterogeneous variation from different features.

Value

list of length one with run-level data.

Examples

```
raw = DDARawData
method = "TMP"
cens = "NA"
impute = TRUE
MSstatsConvert::MSstatsLogsSettings(FALSE)
input = MSstatsPrepareForDataProcess(raw, 2, NULL)
input = MSstatsNormalize(input, "EQUALIZEMEDIANS")
input = MSstatsMergeFractions(input)
input = MSstatsHandleMissing(input, "TMP", TRUE, "NA", 0.999)
input = MSstatsSelectFeatures(input, "all")
processed = getProcessed(input)
input = MSstatsPrepareForSummarization(input, method, impute, cens, FALSE)
input_split = split(input, input$PROTEIN)
summarized = MSstatsSummarize(input_split, method, impute, cens, FALSE, TRUE)
length(summarized) # list of summarization outputs for each protein
head(summarized[[1]][[1]]) # run-level summary
```

MSstatsSummarizeSingleLinear

Linear model-based summarization for a single protein

Description

Linear model-based summarization for a single protein

Usage

```
MSstatsSummarizeSingleLinear(single_protein, equal_variances = TRUE)
```

Arguments

single_protein feature-level data for a single protein

equal_variances

if TRUE, observation are assumed to be homoskedastic

Value

list with protein-level data

Examples

```
raw = DDARawData
method = "linear"
cens = NULL
impute = FALSE
# currently, MSstats only supports MBimpute = FALSE for linear summarization
MSstatsConvert::MSstatsLogsSettings(FALSE)
input = MSstatsPrepareForDataProcess(raw, 2, NULL)
input = MSstatsNormalize(input, "EQUALIZEMEDIANS")
input = MSstatsMergeFractions(input)
input = MSstatsHandleMissing(input, "TMP", TRUE, "NA", 0.999)
input = MSstatsSelectFeatures(input, "all")
input = MSstatsPrepareForSummarization(input, method, impute, cens, FALSE)
input_split = split(input, input$PROTEIN)
single_protein_summary = MSstatsSummarizeSingleLinear(input_split[[1]])
head(single_protein_summary[[1]])
```

MSstatsSummarizeSingleTMP

Tukey Median Polish summarization for a single protein

Description

Tukey Median Polish summarization for a single protein

Usage

```
MSstatsSummarizeSingleTMP(
  single_protein,
  impute,
  censored_symbol,
  remove50missing
)
```

Arguments

single_protein   feature-level data for a single protein

impute   only for summaryMethod = "TMP" and censoredInt = 'NA' or '0'. TRUE (default) imputes 'NA' or '0' (depending on censoredInt option) by Accelated failure model. FALSE uses the values assigned by cutoffCensored

censored_symbol

>Missing values are censored or at random. 'NA' (default) assumes that all 'NA's in 'Intensity' column are censored. '0' uses zero intensities as censored intensity. In this case, NA intensities are missing at random. The output from Skyline should use '0'. Null assumes that all NA intensites are randomly missing.

remove50missing

>only for summaryMethod = "TMP". TRUE removes the runs which have more than 50% missing values. FALSE is default.

Value

list of two data.tables: one with fitted survival model, the other with protein-level data

Examples

```
raw = DDARawData
method = "TMP"
cens = "NA"
impute = TRUE
# currently, MSstats only supports MBimpute = FALSE for linear summarization
MSstatsConvert::MSstatsLogsSettings(FALSE)
input = MSstatsPrepareForDataProcess(raw, 2, NULL)
input = MSstatsNormalize(input, "EQUALIZEMEDIANS")
input = MSstatsMergeFractions(input)
input = MSstatsHandleMissing(input, "TMP", TRUE, "NA", 0.999)
input = MSstatsSelectFeatures(input, "all")
input = MSstatsPrepareForSummarization(input, method, impute, cens, FALSE)
input_split = split(input, input$PROTEIN)
single_protein_summary = MSstatsSummarizeSingleTMP(input_split[[1]],
                                                   impute, cens, FALSE)
head(single_protein_summary[[1]])
```

OpenMStoMSstatsFormat *Import OpenMS files*

Description

Import OpenMS files

Usage

```
OpenMStoMSstatsFormat(
  input,
  annotation = NULL,
  useUniquePeptide = TRUE,
  removeFewMeasurements = TRUE,
  removeProtein_with1Feature = FALSE,
  summaryforMultipleRows = max,
```

```
    use_log_file = TRUE,
    append = FALSE,
    verbose = TRUE,
    log_file_path = NULL,
    ...
)
```

Arguments

input	name of MSstats input report from OpenMS, which includes feature(peptide ion)-level data.
annotation	name of 'annotation.txt' data which includes Condition, BioReplicate, Run. Run should be the same as filename.
useUniquePeptide	
	TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.
removeFewMeasurements	
	TRUE (default) will remove the features that have 1 or 2 measurements across runs.
removeProtein_with1Feature	
	TRUE will remove the proteins which have only 1 feature, which is the combination of peptide, precursor charge, fragment and charge. FALSE is default.
summaryforMultipleRows	
	max(default) or sum - when there are multiple measurements for certain feature and certain run, use highest or sum of multiple intensities.
use_log_file	logical. If TRUE, information about data processing will be saved to a file.
append	logical. If TRUE, information about data processing will be added to an existing log file.
verbose	logical. If TRUE, information about data processing wil be printed to the console.
log_file_path	character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If 'append = TRUE', has to be a valid path to a file.
...	additional parameters to 'data.table::fread'.

Value

data.frame in the MSstats required format.

Author(s)

Meena Choi, Olga Vitek.

Examples

```
openms_raw = data.table::fread(system.file("tinytest/raw_data/OpenMS/openms_input.csv",
                                package = "MSstatsConvert"))
openms_imported = OpenMStoMSstatsFormat(openms_raw, use_log_file = FALSE)
head(openms_imported)
```

OpenSWATHtoMSstatsFormat

Import OpenSWATH files

Description

Import OpenSWATH files

Usage

```
OpenSWATHtoMSstatsFormat(
  input,
  annotation,
  filter_with_mscore = TRUE,
  mscore_cutoff = 0.01,
  useUniquePeptide = TRUE,
  removeFewMeasurements = TRUE,
  removeProtein_with1Feature = FALSE,
  summaryforMultipleRows = max,
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL,
  ...
)
```

Arguments

input	name of MSstats input report from OpenSWATH, which includes feature-level data.
annotation	name of 'annotation.txt' data which includes Condition, BioReplicate, Run. Run should be the same as filename.
filter_with_mscore	
	TRUE(default) will filter out the features that have greater than mscore_cutoff in m_score column. Those features will be removed.
mscore_cutoff	Cutoff for m_score. Default is 0.01.
useUniquePeptide	
	TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.

removeFewMeasurements

> TRUE (default) will remove the features that have 1 or 2 measurements across runs.

removeProtein_with1Feature

> TRUE will remove the proteins which have only 1 feature, which is the combination of peptide, precursor charge, fragment and charge. FALSE is default.

summaryforMultipleRows

> max(default) or sum - when there are multiple measurements for certain feature and certain run, use highest or sum of multiple intensities.

use_log_file   logical. If TRUE, information about data processing will be saved to a file.

append   logical. If TRUE, information about data processing will be added to an existing log file.

verbose   logical. If TRUE, information about data processing wil be printed to the console.

log_file_path character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If 'append = TRUE', has to be a valid path to a file.

...   additional parameters to 'data.table::fread'.

Value

data.frame in the MSstats required format.

Author(s)

Meena Choi, Olga Vitek.

Examples

```
os_raw = system.file("tinytest/raw_data/OpenSWATH/openswath_input.csv",
                            package = "MSstatsConvert")
annot = system.file("tinytest/annotations/annot_os.csv",
                        package = "MSstats")
os_raw = data.table::fread(os_raw)
annot = data.table::fread(annot)

os_imported = OpenSWATHtoMSstatsFormat(os_raw, annot, use_log_file = FALSE)
head(os_imported)
```

PDtoMSstatsFormat   *Import Proteome Discoverer files*

Description

Import Proteome Discoverer files

Usage

```
PDtoMSstatsFormat(
  input,
  annotation,
  useNumProteinsColumn = FALSE,
  useUniquePeptide = TRUE,
  summaryforMultipleRows = max,
  removeFewMeasurements = TRUE,
  removeOxidationMpeptides = FALSE,
  removeProtein_with1Peptide = FALSE,
  which.quantification = "Precursor.Area",
  which.proteinid = "Protein.Group.Accessions",
  which.sequence = "Sequence",
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL,
  ...
)
```

Arguments

input	PD report or a path to it.
annotation	name of 'annotation.txt' or 'annotation.csv' data which includes Condition, BioReplicate, Run information. 'Run' will be matched with 'Spectrum.File'.
useNumProteinsColumn	
	TRUE removes peptides which have more than 1 in # Proteins column of PD output.
useUniquePeptide	
	TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.
summaryforMultipleRows	
	max(default) or sum - when there are multiple measurements for certain feature and certain run, use highest or sum of multiple intensities.
removeFewMeasurements	
	TRUE (default) will remove the features that have 1 or 2 measurements across runs.
removeOxidationMpeptides	
	TRUE will remove the peptides including 'oxidation (M)' in modification. FALSE is default.
removeProtein_with1Peptide	
	TRUE will remove the proteins which have only 1 peptide and charge. FALSE is default.
which.quantification	
	Use 'Precursor.Area'(default) column for quantified intensities. 'Intensity' or 'Area' can be used instead.

which.proteinid
> Use 'Protein.Accessions'(default) column for protein name. 'Master.Protein.Accessions' can be used instead.

which.sequence Use 'Sequence'(default) column for peptide sequence. 'Annotated.Sequence' can be used instead.

use_log_file logical. If TRUE, information about data processing will be saved to a file.

append logical. If TRUE, information about data processing will be added to an existing log file.

verbose logical. If TRUE, information about data processing wil be printed to the console.

log_file_path character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If 'append = TRUE', has to be a valid path to a file.

... additional parameters to 'data.table::fread'.

Value

data.frame in the MSstats required format.

Author(s)

Meena Choi, Olga Vitek

Examples

```
pd_raw = system.file("tinytest/raw_data/PD/pd_input.csv",
                      package = "MSstatsConvert")
annot = system.file("tinytest/annotations/annot_pd.csv", package = "MSstats")
pd_raw = data.table::fread(pd_raw)
annot = data.table::fread(annot)

pd_imported = PDtoMSstatsFormat(pd_raw, annot, use_log_file = FALSE)
head(pd_imported)
```

ProgenesistoMSstatsFormat
Import Progenesis files

Description

Import Progenesis files

Usage

```
ProgenesistoMSstatsFormat(
  input,
  annotation,
  useUniquePeptide = TRUE,
  summaryforMultipleRows = max,
  removeFewMeasurements = TRUE,
  removeOxidationMpeptides = FALSE,
  removeProtein_with1Peptide = FALSE,
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL,
  ...
)
```

Arguments

input	name of Progenesis output, which is wide-format. 'Accession', 'Sequence', 'Modification', 'Charge' and one column for each run are required.
annotation	name of 'annotation.txt' or 'annotation.csv' data which includes Condition, BioReplicate, Run information. It will be matched with the column name of input for MS runs.
useUniquePeptide	
	TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.
summaryforMultipleRows	
	max(default) or sum - when there are multiple measurements for certain feature and certain run, use highest or sum of multiple intensities.
removeFewMeasurements	
	TRUE (default) will remove the features that have 1 or 2 measurements across runs.
removeOxidationMpeptides	
	TRUE will remove the peptides including 'oxidation (M)' in modification. FALSE is default.
removeProtein_with1Peptide	
	TRUE will remove the proteins which have only 1 peptide and charge. FALSE is default.
use_log_file	logical. If TRUE, information about data processing will be saved to a file.
append	logical. If TRUE, information about data processing will be added to an existing log file.
verbose	logical. If TRUE, information about data processing wil be printed to the console.
log_file_path	character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If 'append = TRUE', has to be a valid path to a file.
...	additional parameters to 'data.table::fread'.

Value

data.frame in the MSstats required format.

Author(s)

Meena Choi, Olga Vitek, Ulrich Omasits

Examples

```
progenesis_raw = system.file("tinytest/raw_data/Progenesis/progenesis_input.csv",
                             package = "MSstatsConvert")
annot = system.file("tinytest/raw_data/Progenesis/progenesis_annot.csv",
                    package = "MSstatsConvert")
progenesis_raw = data.table::fread(progenesis_raw)
annot = data.table::fread(annot)

progenesis_imported = ProgenesistoMSstatsFormat(progenesis_raw, annot,
                                                use_log_file = FALSE)
head(progenesis_imported)
```

---

quantification *Protein sample quantification or group quantification*

Description

Model-based quantification for each condition or for each biological sample per protein in a targeted Selected Reaction Monitoring (SRM), Data-Dependent Acquisition (DDA or shotgun), and Data-Independent Acquisition (DIA or SWATH-MS) experiment. Quantification takes the processed data set by dataProcess as input and automatically generate the quantification results (data.frame) in a long or matrix format.

Usage

```
quantification(
  data,
  type = "Sample",
  format = "matrix",
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL
)
```

Arguments

data	name of the (processed) data set.
type	choice of quantification. "Sample" or "Group" for protein sample quantification or group quantification.
format	choice of returned format. "long" for long format which has the columns named Protein, Condition, LogIntensities (and BioReplicate if it is subject quantification), NumFeature for number of transitions for a protein, and NumPeaks for number of observed peak intensities for a protein. "matrix" for data matrix format which has the rows for Protein and the columns, which are Groups(or Conditions) for group quantification or the combinations of BioReplicate and Condition (labeled by "BioReplicate"_"Condition") for sample quantification. Default is "matrix"
use_log_file	logical. If TRUE, information about data processing will be saved to a file.
append	logical. If TRUE, information about data processing will be added to an existing log file.
verbose	logical. If TRUE, information about data processing wil be printed to the console.
log_file_path	character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If 'append = TRUE', has to be a valid path to a file.

Details

- Sample quantification : individual biological sample quantification for each protein. The label of each biological sample is a combination of the corresponding group and the sample ID. If there are no technical replicates or experimental replicates per sample, sample quantification is the same as run summarization from dataProcess. If there are technical replicates or experimental replicates, sample quantification is median among run quantification corresponding MS runs.

- Group quantification : quantification for individual group or individual condition per protein. It is median among sample quantification.

- The quantification for endogenous samples is based on run summarization from subplot model, with TMP robust estimation.

Value

data.frame as described in details.

Examples

```
# Consider quantitative data (i.e. QuantData) from a yeast study with ten time points of
# interests, three biological replicates, and no technical replicates which is
# a time-course experiment.
# Sample quantification shows model-based estimation of protein abundance in each biological
# replicate within each time point.
# Group quantification shows model-based estimation of protein abundance in each time point.
QuantData<-dataProcess(SRMRawData, use_log_file = FALSE)
```

```
head(QuantData$FeatureLevelData)
# Sample quantification
sampleQuant<-quantification(QuantData, use_log_file = FALSE)
head(sampleQuant)
# Group quantification
groupQuant<-quantification(QuantData, type="Group", use_log_file = FALSE)
head(groupQuant)
```

savePlot   *Save a plot to pdf file*

Description

Save a plot to pdf file

Usage

```
savePlot(name_base, file_name, width, height)
```

Arguments

name_base	path to a folder (or "" for working directory)
file_name	name of a file to save. If this file already exists, an integer will be appended to this name
width	width of a plot
height	height of a plot

SkylinetoMSstatsFormat

Import Skyline files

Description

Import Skyline files

Usage

```
SkylinetoMSstatsFormat(
  input,
  annotation = NULL,
  removeiRT = TRUE,
  filter_with_Qvalue = TRUE,
  qvalue_cutoff = 0.01,
  useUniquePeptide = TRUE,
```

```
    removeFewMeasurements = TRUE,
    removeOxidationMpeptides = FALSE,
    removeProtein_with1Feature = FALSE,
    use_log_file = TRUE,
    append = FALSE,
    verbose = TRUE,
    log_file_path = NULL,
    ...
)
```

Arguments

input	name of MSstats input report from Skyline, which includes feature-level data.
annotation	name of 'annotation.txt' data which includes Condition, BioReplicate, Run. If annotation is already complete in Skyline, use annotation=NULL (default). It will use the annotation information from input.
removeiRT	TRUE (default) will remove the proteins or peptides which are labeld 'iRT' in 'StandardType' column. FALSE will keep them.
filter_with_Qvalue	
	TRUE(default) will filter out the intensities that have greater than qvalue_cutoff in DetectionQValue column. Those intensities will be replaced with zero and will be considered as censored missing values for imputation purpose.
qvalue_cutoff	Cutoff for DetectionQValue. default is 0.01.
useUniquePeptide	
	TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.
removeFewMeasurements	
	TRUE (default) will remove the features that have 1 or 2 measurements across runs.
removeOxidationMpeptides	
	TRUE will remove the peptides including 'oxidation (M)' in modification. FALSE is default.
removeProtein_with1Feature	
	TRUE will remove the proteins which have only 1 feature, which is the combination of peptide, precursor charge, fragment and charge. FALSE is default.
use_log_file	logical. If TRUE, information about data processing will be saved to a file.
append	logical. If TRUE, information about data processing will be added to an existing log file.
verbose	logical. If TRUE, information about data processing wil be printed to the console.
log_file_path	character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If 'append = TRUE', has to be a valid path to a file.
...	additional parameters to 'data.table::fread'.

Value

data.frame in the MSstats required format.

Author(s)

Meena Choi, Olga Vitek

Examples

```
skyline_raw = system.file("tinytest/raw_data/Skyline/skyline_input.csv",
                          package = "MSstatsConvert")
skyline_raw = data.table::fread(skyline_raw)
skyline_imported = SkylinetoMSstatsFormat(skyline_raw)
head(skyline_imported)
```

SpectronauttoMSstatsFormat
                            Import Spectronaut files

Description

Import Spectronaut files

Usage

```
SpectronauttoMSstatsFormat(
  input,
  annotation = NULL,
  intensity = "PeakArea",
  filter_with_Qvalue = TRUE,
  qvalue_cutoff = 0.01,
  useUniquePeptide = TRUE,
  removeFewMeasurements = TRUE,
  removeProtein_with1Feature = FALSE,
  summaryforMultipleRows = max,
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL,
  ...
)
```

Arguments

input	name of Spectronaut output, which is long-format. ProteinName, PeptideSequence, PrecursorCharge, FragmentIon, ProductCharge, IsotopeLabelType, Condition, BioReplicate, Run, Intensity, F.ExcludedFromQuantification are required. Rows with F.ExcludedFromQuantification=True will be removed.
annotation	name of 'annotation.txt' data which includes Condition, BioReplicate, Run. If annotation is already complete in Spectronaut, use annotation=NULL (default). It will use the annotation information from input.
intensity	'PeakArea'(default) uses not normalized peak area. 'NormalizedPeakArea' uses peak area normalized by Spectronaut.
filter_with_Qvalue	
	TRUE(default) will filter out the intensities that have greater than qvalue_cutoff in EG.Qvalue column. Those intensities will be replaced with zero and will be considered as censored missing values for imputation purpose.
qvalue_cutoff	Cutoff for EG.Qvalue. default is 0.01.
useUniquePeptide	
	TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.
removeFewMeasurements	
	TRUE (default) will remove the features that have 1 or 2 measurements across runs.
removeProtein_with1Feature	
	TRUE will remove the proteins which have only 1 feature, which is the combination of peptide, precursor charge, fragment and charge. FALSE is default.
summaryforMultipleRows	
	max(default) or sum - when there are multiple measurements for certain feature and certain run, use highest or sum of multiple intensities.
use_log_file	logical. If TRUE, information about data processing will be saved to a file.
append	logical. If TRUE, information about data processing will be added to an existing log file.
verbose	logical. If TRUE, information about data processing wil be printed to the console.
log_file_path	character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If 'append = TRUE', has to be a valid path to a file.
...	additional parameters to 'data.table::fread'.

Value

data.frame in the MSstats required format.

Author(s)

Meena Choi, Olga Vitek

Examples

```
spectronaut_raw = system.file("tinytest/raw_data/Spectronaut/spectronaut_input.csv",
                              package = "MSstatsConvert")
spectronaut_raw = data.table::fread(spectronaut_raw)
spectronaut_imported = SpectronauttoMSstatsFormat(spectronaut_raw, use_log_file = FALSE)
head(spectronaut_imported)
```

SRMRawData   *Example dataset from a SRM experiment with stable isotope labeled*
                      reference of a time course yeast study

Description

This is a partial data set obtained from a published study (Picotti, et. al, 2009). The experiment
targeted 45 proteins in the glycolysis/gluconeogenesis/TCA cycle/glyoxylate cycle network, which
spans the range of protein abundance from less than 128 to 10E6 copies per cell. Three biological
replicates were analyzed at ten time points (T1-T10), while yeasts transited through exponential
growth in a glucose-rich medium (T1-T4), diauxic shift (T5-T6), post-diauxic phase (T7-T9), and
stationary phase (T10). Prior to trypsinization, the samples were mixed with an equal amount of
proteins from the same N15-labeled yeast sample, which was used as a reference. Each sample
was profiled in a single mass spectrometry run, where each protein was represented by up to two
peptides and each peptide by up to three transitions. The goal of this study is to detect significantly
change in protein abundance across time points. Transcriptional activity under the same experimen-
tal conditions has been previously investigated by (DeRisi et. al., 1997). Genes coding for 29 of
the proteins are differentially expressed between conditions similar to those represented by T7 and
T1 and could be treated as external sources to validate the proteomics analysis. In this exampled
data set, two of the targeted proteins are selected and validated with gene expression study: Pro-
tein IDHC (gene name IDP2) is differentially expressed in time point 1 and time point 7, whereas,
Protein PMG2 (gene name GPM2) is not. The protein names are based on Swiss Prot Name.

Usage

```
SRMRawData
```

Format

data.frame

Details

The raw data (input data for MSstats) is required to contain variable of ProteinName, PeptideSe-
quence, PrecursorCharge, FragmentIon, ProductCharge, IsotopeLabelType, Condition, BioRepli-
cate, Run, Intensity. The variable names should be fixed.

If the information of one or more columns is not available for the original raw data, please retain
the column variables and type in fixed value. For example, the original raw data does not contain

the information of ProductCharge, we retain the column ProductCharge and type in NA for all transitions in RawData.

The column BioReplicate should label with unique patient ID (i.e., same patients should label with the same ID).

Variable Intensity is required to be original signal without any log transformation and can be specified as the peak of height or the peak of area under curve.

Value

data.frame with the required format of MSstats.

Author(s)

Meena Choi, Olga Vitek.

Maintainer: Meena Choi (<mnchoi67@gmail.com>)

References

Ching-Yun Chang, Paola Picotti, Ruth Huttenhain, Viola Heinzelmann-Schwarz, Marko Jovanovic, Ruedi Aebersold, Olga Vitek. Protein significance analysis in selected reaction monitoring (SRM) measurements. *Molecular & Cellular Proteomics*, 11:M111.014662, 2012.

Examples

```
head(SRMRawData)
```

---

theme_msstats   *Theme for MSstats plots*

Description

Theme for MSstats plots

Usage

```
theme_msstats(
  type,
  x.axis.size = 10,
  y.axis.size = 10,
  legend_size = 13,
  strip_background = element_rect(fill = "gray95"),
  strip_text_x = element_text(colour = c("black"), size = 14),
  legend_position = "top",
  legend_box = "vertical",
  text_angle = 0,
  text_hjust = NULL,
  text_vjust = NULL,
  ...
)
```

Arguments

`type`	type of a plot
`x.axis.size`	size of text on the x axis
`y.axis.size`	size of text on the y axis
`legend_size`	size of the legend
`strip_background`	
	background of facet
`strip_text_x`	size of text on facets
`legend_position`	
	position of the legend
`legend_box`	legend.box
`text_angle`	angle of text on the x axis (for condition and comparison plots)
`text_hjust`	hjust parameter for x axis text (for condition and comparison plots)
`text_vjust`	vjust parameter for x axis text (for condition and comparison plots)
`...`	additional parameters passed on to ggplot2::theme()

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