

# Package ‘ALDEx2’

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**Type** Package

**Title** Analysis Of Differential Abundance Taking Sample Variation Into Account

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**Author** Greg Gloor, Ruth Grace Wong, Andrew Fernandes, Arianne Albert, Matt Links, Thomas Quinn, Jia Rong Wu

**Maintainer** Greg Gloor <ggloor@uwo.ca>

**biocViews** DifferentialExpression, RNASeq, DNASEq, ChIPSeq, GeneExpression, Bayesian, Sequencing, Software, Microbiome, Metagenomics

**Description** A differential abundance analysis for the comparison of two or more conditions. Useful for analyzing data from standard RNA-seq or meta-RNA-seq assays as well as selected and unselected values from in-vitro sequence selections. Uses a Dirichlet-multinomial model to infer abundance from counts, optimized for three or more experimental replicates. The method infers biological and sampling variation to calculate the expected false discovery rate, given the variation, based on a Wilcox rank test or Welch t-test (via `aldex.ttest`), or a glm and Kruskal-Wallis test (via `aldex.glm`). Reports p-values and Benjamini-Hochberg corrected p-values.

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ALDEx2m-package	<i>Analysis of differential abundance taking sample variation into account</i>
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**Description**

A differential abundance analysis for the comparison of two or more conditions. For example, single-organism and meta-RNA-seq high-throughput sequencing assays, or of selected and unselected values from in-vitro sequence selections. Uses a Dirichlet-multinomial model to infer abundance from counts, that has been optimized for three or more experimental replicates. Infers sampling variation and calculates the expected false discovery rate given the biological and sampling variation using the Wilcoxon rank test or Welch's t-test (`aldex.ttest`) or the `glm` and Kruskal Wallis tests (`aldex.glm`). Reports both P and fdr values calculated by the Benjamini Hochberg correction.

**References**

Please use the citation given by `citation(package="ALDEx")`.

**See Also**

[aldex.clr](#), [aldex.ttest](#), [aldex.glm](#), [aldex.effect](#), [selex](#)

**Examples**

```
# see examples for the aldex.clr, aldex.ttest, aldex.effect, aldex.glm functions
```

---

aldex *Compute an aldex Object*

---

## Description

Welcome to the ALDEx2 package!

The aldex function is a wrapper that performs log-ratio transformation and statistical testing in a single line of code. Specifically, this function: (a) generates Monte Carlo samples of the Dirichlet distribution for each sample, (b) converts each instance using a log-ratio transform, then (c) returns test results for two sample (Welch's t, Wilcoxon) or multi-sample (glm, Kruskal Wallace) tests. This function also estimates effect size for two sample analyses.

## Usage

```
aldex(reads, conditions, mc.samples = 128, test = "t", effect = TRUE,
      include.sample.summary = FALSE, verbose = FALSE, denom = "all")
```

## Arguments

reads	A non-negative, integer-only data.frame or matrix with unique names for all rows and columns. Rows should contain genes and columns should contain sequencing read counts (i.e., sample vectors). Rows with 0 reads in each sample are deleted prior to analysis.
conditions	A character vector. A description of the data structure used for testing. Typically, a vector of group labels.
mc.samples	An integer. The number of Monte Carlo samples to use when estimating the underlying distributions. Since we are estimating central tendencies, 128 is usually sufficient.
test	A character string. Indicates which tests to perform. "t" calls Welch's t and Wilcoxon tests. "glm" calls Kruskal Wallace and glm tests. "iterative" uses the results from an initial "t" routine to seed the denominator (i.e., for the Geometric Mean calculation) of a second "t" routine.
effect	A boolean. Toggles whether to calculate abundances and effect sizes. Applies to test = "t" and test = "iterative".
include.sample.summary	A boolean. Toggles whether to include median clr values for each sample. Applies to effect = TRUE.
verbose	A boolean. Toggles whether to print diagnostic information while running. Useful for debugging errors on large datasets. Applies to effect = TRUE.
denom	A character string. Indicates which features to retain as the denominator for the Geometric Mean calculation. Using "iqlr" accounts for data with systematic variation and centers the features on the set features that have variance that is between the lower and upper quartile of variance. Using "zero" is a more extreme case where there are many non-zero features in one condition but many zeros in another. In this case the geometric mean of each group is calculated using the set of per-group non-zero features.

## Details

See "Examples" below for a description of the sample input.

**Value**

Returns a number of values that depends on the set of options. See the return values of `aldex.ttest`, `aldex.glm`, and `aldex.effect` for explanations and example.

**Author(s)**

Greg Gloor, Andrew Fernandes, and Matt Links contributed to the original package. Thom Quinn added the "iterative" test method.

**References**

Please use the citation given by `citation(package="ALDEx")`.

**See Also**

[aldex.ttest](#), [aldex.glm](#), [aldex.effect](#), [aldex.corr](#), [selex](#)

**Examples**

```
# The 'reads' data.frame should have row
# and column names that are unique, and
# looks like the following:
#
#           T1a T1b T2 T3 N1 N2 Nx
# Gene_00001  0  0  2  0  0  1  0
# Gene_00002 20  8 12  5 19 26 14
# Gene_00003  3  0  2  0  0  0  1
# Gene_00004 75 84 241 149 271 257 188
# Gene_00005 10 16  4  0  4  10 10
# Gene_00006 129 126 451 223 243 149 209
#           ... many more rows ...

data(selex)
selex <- selex[1201:1600,] # subset for efficiency
conds <- c(rep("NS", 7), rep("S", 7))
x <- aldex(selex, conds, mc.samples=2, denom="all",
           test="t", effect=FALSE)
```

---

aldex.clr

---

*Compute an aldex.clr Object*


---

**Description**

Generate Monte Carlo samples of the Dirichlet distribution for each sample. Convert each instance using the centred log-ratio transform This is the input for all further analyses.

**Usage**

```
aldex.clr(reads, conds, mc.samples = 128, denom="all", verbose=FALSE, useMC=FALSE)
```

**Arguments**

reads	A <code>data.frame</code> or <code>RangedSummarizedExperiment</code> object containing non-negative integers only and with unique names for all rows and columns, where each row is a different gene and each column represents a sequencing read-count. Rows with 0 reads in each sample are deleted prior to analysis.
conds	A vector containing a descriptor for the samples, allowing them to be grouped and compared.
mc.samples	The number of Monte Carlo samples to use to estimate the underlying distributions; since we are estimating central tendencies, 128 is usually sufficient.
denom	A character variable (all, iqlr, zero, lvha) indicating which features to use as the denominator for the Geometric Mean calculation. The default "all" uses the geometric mean abundance of all features. Using "iqlr" uses the features that are between the first and third quartile of the variance of the clr values across all samples. Using "zero" uses the non-zero features in each group as the denominator. This approach is an extreme case where there are many nonzero features in one condition but many zeros in another. Using "lvha" uses features that have low variance (bottom quartile) and high relative abundance (top quartile in every sample). It is also possible to supply a vector of row indices to use as the denominator. Here, the experimentalist is determining a-priori which rows are thought to be invariant. In the case of RNA-seq, this could include ribosomal protein genes and other house-keeping genes.
verbose	Print diagnostic information while running. Useful only for debugging if fails on large datasets.
useMC	Use multicore by default (FALSE). Multi core processing will be attempted with the BiocParallel package. Serial processing will be used if this is not possible.

**Details**

An explicit description of the input format for the reads object is shown under ‘Examples’, below.

**Value**

The object produced by the `clr` function contains the clr transformed values for each Monte-Carlo Dirichlet instance, which can be accessed through `getMonteCarloInstances(x)`, where `x` is the `clr` function output. Each list element is named by the sample ID. `getFeatures(x)` returns the features, `getSampleIDs(x)` returns sample IDs, and `getFeatureNames(x)` returns the feature names.

**Author(s)**

Greg Gloor, Ruth Grace Wong, Andrew Fernandes, Matt Links and Jia Rong Wu contributed to this code.

**References**

Please use the citation given by `citation(package="ALDEX")`.

**See Also**

[aldex.ttest](#), [aldex.glm](#), [aldex.effect](#), [selex](#)

## Examples

```
# The 'reads' data.frame or
# RangedSummarizedExperiment object should
# have row and column names that are unique,
# and looks like the following:
#
#           T1a T1b T2 T3 N1 N2 Nx
# Gene_00001  0  0  2  0  0  1  0
# Gene_00002 20  8 12  5 19 26 14
# Gene_00003  3  0  2  0  0  0  1
# Gene_00004 75 84 241 149 271 257 188
# Gene_00005 10 16  4  0  4 10 10
# Gene_00006 129 126 451 223 243 149 209
#           ... many more rows ...

data(selex)
#subset for efficiency
selex <- selex[1201:1600,]
conds <- c(rep("NS", 7), rep("S", 7))
x <- aldex.clr(selex, conds, mc.samples=2, denom="all", verbose=FALSE)
```

---

aldex.clr-class

*The aldex.clr class*


---

## Description

The aldex.clr S4 class is a class which stores the data generated by the aldex.clr method.

## Details

An aldex.clr object contains the Monte Carlo Dirichlet instances derived from estimating the technical variance of the raw read count data. It is created by the aldex.clr.function, which is invoked by the aldex.clr method. It consists of four attributes: the sample names, the feature names, the conditions vector (assigns each sample to a condition), and the Monte Carlo Dirichlet instances themselves. These can be accessed, along with information about the length of some attributes. A single Monte Carlo instance can also be retrieved.

## Value

The aldex.clr object contains the clr transformed values for each Monte-Carlo Dirichlet instance, which can be accessed through getMonteCarloInstances(x), where x is the clr function output. Each list element is named by the sample ID. getFeatures(x) returns the features, getSampleIDs(x) returns sample IDs, and getFeatureNames(x) returns the feature names.

## Methods

In the code below, x is an aldex.clr object, and i is a numeric whole number.

getMonteCarloInstances(x): Returns x's Monte Carlo Dirichlet instances.

getSampleIDs(x): Returns the names of the samples. These can be used to access the original reads, as in reads\$sampleID (if the reads are a data frame).

`getFeatures(x)`: Returns the names of the features as a vector.  
`numFeatures(x)`: Returns the number of features associated with the data.  
`numMCInstances(x)`: Returns the names of the keys that can be used to subset the data rows. The keys values are the rsid's.  
`getFeatureNames(x)`: Returns the names of the keys that can be used to subset the data rows. The keys values are the rsid's.  
`getReads(x)`: Returns the names of the keys that can be used to subset the data rows. The keys values are the rsid's.  
`numConditions(x)`: Returns the names of the keys that can be used to subset the data rows. The keys values are the rsid's.  
`getMonteCarloReplicate(x, i)`: Returns the names of the keys that can be used to subset the data rows. The keys values are the rsid's.

### Author(s)

Greg Gloor, Ruth Grace Wong, Andrew Fernandes, Jia Rong Wu and Matt Links contributed to this code

### References

Please use the citation given by `citation(package="ALDEX")`.

### See Also

[aldex.clr.function](#)

### Examples

```

# The 'reads' data.frame or
# SummarizedExperiment object should have
# row and column names that are unique,
# and looks like the following:
#
#           T1a T1b T2 T3 N1 N2 Nx
# Gene_00001  0  0  2  0  0  1  0
# Gene_00002 20  8 12  5 19 26 14
# Gene_00003  3  0  2  0  0  0  1
# Gene_00004 75 84 241 149 271 257 188
# Gene_00005 10 16  4  0  4 10 10
# Gene_00006 129 126 451 223 243 149 209
#           ... many more rows ...

data(selex)
#subset for efficiency
selex <- selex[1201:1600,]
conds <- c(rep("NS", 7), rep("S", 7))

# x is an object of type aldex.clr
x <- aldex.clr(selex, conds, mc.samples = 2, denom="all", verbose = FALSE)

# get all of the Monte Carlo Dirichlet instances
monteCarloInstances <- getMonteCarloInstances(x)
  
```

```
# get sample names
sampleIDs <- getSampleIDs(x)

# get features
features <- getFeatures(x)

# get number of features
numFeatures <- numFeatures(x)

# get number of Monte Carlo Dirochlet instances
numInstances <- numMCInstances(x)

# get names of features
featureNames <- getFeatureNames(x)

# get number of conditions
conditions <- numConditions(x)

# get number of conditions
reads <- getReads(x)

# retrieve the first Monte Carlo Dirochlet instance.
monteCarloInstance <- getMonteCarloReplicate(x,1)
```

---

aldex.corr

*calculate Pearson's Product moment and Spearman's rank correlations*

---

## Description

calculates expected values of Pearson's Product moment and Spearman's rank correlations on the data returned by aldex.clr. NOTE: this function will be replaced by a compositionally correct method in the next release cycle.

## Usage

```
aldex.corr(clr, covar)
```

## Arguments

clr	clr is the data output of the aldex.clr function
covar	a per-sample continuous variable to be correlated with the clr values

## Details

An explicit example for two conditions is shown in the 'Examples' below.

## Value

Outputs a dataframe with the following information:

pearson.ecor	a vector containing the expected Pearson's Product moment value for each feature
--------------	--



pearson.ep	a vector containing the expected P value of the Pearson Product moment value for each feature
pearson.eBH	a vector containing the expected Benjamini-Hochberg corrected P value of the Pearson Product moment value for each feature
spearman.erho	a vector containing the expected Spearman's rank correlation value for each feature
spearman.ep	a vector containing the expected P value of Spearman's rank correlation value for each feature
spearman.eBH	a vector containing the expected Benjamini-Hochberg corrected P value of Spearman's rank correlation value for each feature

**Author(s)**

Arianne Albert

**References**

Please use the citation given by `citation(package="ALDEX")`.

**See Also**

[aldex.clr](#), [aldex.glm](#), [aldex.effect](#), [selex](#)

**Examples**

```
# x is the output of the \code{x <- aldex.clr(data, mc.samples)} function
# conditions is a description of the data
# aldex.ttest(clr, covar)
```

---

aldex.effect	<i>calculate effect sizes and differences between conditions</i>
--------------	--

---

**Description**

determines the median clr abundance of the feature in all samples and in groups determines the median difference between the two groups determines the median variation within each two group determines the effect size, which is the median of the ratio of the between group difference and the larger of the variance within groups

**Usage**

```
aldex.effect(clr, conditions, verbose = TRUE, include.sample.summary = FALSE,
             useMC=FALSE)
```

**Arguments**

clr	clr is the data output of <code>aldex.clr</code>
conditions	a description of the data structure to be used for testing
verbose	Print diagnostic information while running. Useful only for debugging if fails on large datasets
include.sample.summary	include median clr values for each sample, defaults to FALSE
useMC	use multicore by default (FALSE)

## Details

An explicit example for two conditions is shown in the ‘Examples’ below.

## Value

returns a dataframe with the following information:

<code>rab.all</code>	a vector containing the median clr value for each feature
<code>rab.win.conditionA</code>	a vector containing the median clr value for each feature in condition A
<code>rab.win.conditionB</code>	a vector containing the median clr value for each feature in condition B
<code>diff.btw</code>	a vector containing the per-feature median difference between condition A and B
<code>diff.win</code>	a vector containing the per-feature maximum median difference between Dirichlet instances within conditions
<code>effect</code>	a vector containing the per-feature effect size
<code>overlap</code>	a vector containing the per-feature proportion of effect size that is 0 or less

## Author(s)

Greg Gloor, Andrew Fernandes, Matt Links

## References

Please use the citation given by `citation(package="ALDEx")`.

## See Also

[aldex.clr](#), [aldex.ttest](#), [aldex.glm](#), [selex](#)

## Examples

```
# x is the output of the \code{x <- clr(data, mc.samples)} function
# conditions is a description of the data
# for the selex dataset, conditions <- c(rep("N", 7), rep("S", 7))
data(selex)
#subset for efficiency
selex <- selex[1201:1600,]
conds <- c(rep("NS", 7), rep("S", 7))
x <- aldex.clr(selex, conds, mc.samples=2, denom="all")
effect.test <- aldex.effect(x, conds)
```

---

aldex.glm                      *calculate glm and Kruskal Wallis test statistics*

---

### Description

calculates expected values of the glm and Kruskal Wallis functions on the data returned by `clr_function.r`

### Usage

```
aldex.glm(clr, conditions, useMC=FALSE)
```

### Arguments

<code>clr</code>	<code>clr</code> is the data output of <code>aldex.clr</code>
<code>conditions</code>	a description of the data structure to be used for testing
<code>useMC</code>	use multicore by default (FALSE)

### Details

An explicit example for two conditions is shown in the ‘Examples’ below.

### Value

Outputs a dataframe with the following information:

<code>kw.ep</code>	a vector containing the expected P value of the Kruskal Wallis test for each feature
<code>kw.eBH</code>	a vector containing the expected value of the Benjamini Hochberg corrected P value for each feature
<code>glm.ep</code>	a vector containing the expected P value of the glm test for each feature
<code>glm.eBH</code>	a vector containing the expected value of the Benjamini Hochberg corrected P value for each feature

### Author(s)

Arianne Albert

### References

Please use the citation given by `citation(package="ALDEX")`.

### See Also

[aldex.clr](#), [aldex.ttest](#), [aldex.effect](#), [selex](#)

**Examples**

```
# x is the output of the \code{x <- aldex.clr(data, mc.samples)} function
# conditions is a description of the data
# for the selex dataset, conditions <- c(rep("N", 7), rep("S", 7))
data(selex)
#subset for efficiency
selex <- selex[1201:1600,]
conds <- c(rep("NS", 7), rep("S", 7))
x <- aldex.clr(selex, conds, mc.samples=1, denom="all")
glm.test <- aldex.glm(x, conds)
```

aldex.plot

*Plot an aldex Object***Description**

Create 'MW'- or 'MA'-type plots from the given aldex object.

**Usage**

```
## S3 method for class 'plot'
aldex( x, ..., type=c("MW","MA"),
       xlab=NULL, ylab=NULL, xlim=NULL, ylim=NULL,
       all.col=rgb(0,0,0,0.2), all.pch=19, all.cex=0.4,
       called.col=red, called.pch=20, called.cex=0.6,
       thres.line.col=darkgrey, thres.lwd=1.5,
       test=welch, cutoff=0.1, rare.col=black, rare=0,
       rare.pch=20, rare.cex=0.2 )
```

**Arguments**

x	an object of class aldex produced by the aldex function
...	optional, unused arguments included for compatibility with the S3 method signature
type	which type of plot is to be produced. MA is a Bland-Altman style plot; MW is a difference between to a variance within plot as described in the paper
test	the method of calculating significance, one of: welch = welch's t test; wilcox = wilcox rank test; glm = glm; kruskal = Kruskal-Wallace test
cutoff	the Benjamini-Hochberg fdr cutoff, default 0.1
xlab	the x-label for the plot, as per the parent plot function
ylab	the y-label for the plot, as per the parent plot function
xlim	the x-limits for the plot, as per the parent plot function
ylim	the y-limits for the plot, as per the parent plot function
all.col	the default colour of the plotted points
all.pch	the default plotting symbol
all.cex	the default symbol size
called.col	the colour of points with false discovery rate, $q \leq 0.1$

called.pch	the symbol of points with false discovery rate, $q \leq 0.1$
called.cex	the character expansion of points with false discovery rate, $q \leq 0.1$
thres.line.col	the colour of the threshold line where within and between group variation is equivalent
thres.lwd	the width of the threshold line where within and between group variation is equivalent
rare	relative abundance cutoff for rare features, default 0 or the mean abundance
rare.col	color for rare features, default black
rare.pch	the default symbol of rare features
rare.cex	the default symbol size of rare points

### Details

This particular specialization of the `plot` function is relatively simple and provided for convenience. For more advanced control of the plot is best to use the values returned by `summary(x)`.

### Value

None.

### References

Please use the citation given by `citation(package="ALDEX")`.

### See Also

[aldex](#), [aldex.effect](#), [aldex.ttest](#), [aldex.glm](#)

### Examples

```
# See the examples for 'aldex'.
```

---

aldex.set.mode	<i>identify set of denominator features for log-ratio calculation</i>
----------------	---

---

### Description

calculate the features that are to be used as the denominator for the Geometric Mean calculation in `clr_function.R`

### Usage

```
aldex.set.mode(reads, conds, denom="all")
```

**Arguments**

reads	A data frame containing the samples and features per sample.
conds	A vector describing which samples belong to what condition.
denom	Character argument specifying which indices to return. 'all' returns all features in both conditions. 'zero' returns the nonzero count features per condition. 'iqlr' returns the features whose variance falls within the inter-quantile range of the CLR-transformed data. In cases of malformed or null queries, input defaults to 'all'. Additionally, the input can be a numeric vector, which contains a set of row indices to center the data against. Only for advanced users who can pre-determine the invariant set of features within their data.

**Details**

An explicit example for two conditions is shown in the 'Examples' below.

**Value**

Outputs a vector containing indices per condition.

**Author(s)**

Jia Rong Wu

**References**

Please use the citation given by `citation(package="ALDEX")`.

**See Also**

[aldex.clr](#), [aldex.ttest](#), [aldex.effect](#), [selex](#)

**Examples**

```
# x is the output of the \code{x <- clr(data, mc.samples)} function
# conditions is a description of the data
# for the selex dataset, conditions <- c(rep("N", 7), rep("S", 7))
# input can be "all", "iqlr", "zero" or numeric for advanced users
data(selex)
#subset for efficiency
selex <- selex[1201:1600,]
conds <- c(rep("NS", 7), rep("S", 7))
x <- aldex.clr(selex, conds, mc.samples=2, denom="all")
```

---

aldex.ttest

*calculate Welch's t-test and Wilcoxon test statistics*

---

**Description**

calculates expected values of the Welch's t-test and Wilcoxon rank test on the data returned by `clr_function.r`

**Usage**

```
aldex.ttest(clr, conditions, paired.test = FALSE, hist.plot=FALSE)
```

**Arguments**

<code>clr</code>	<code>clr</code> is the data output of the <code>aldex.clr</code> function
<code>conditions</code>	a description of the data structure to be used for testing
<code>paired.test</code>	whether the Welch's test should be paired or not
<code>hist.plot</code>	whether to plot a histogram of P values for an individual Dirichlet Monte-Carlo instance. Plot is output to the standard R plotting device.

**Details**

An explicit example for two conditions is shown in the 'Examples' below.

**Value**

Outputs a dataframe with the following information:

<code>we.ep</code>	a vector containing the expected P value of the Welch's t-test for each feature
<code>we.eBH</code>	a vector containing the expected value of the Benjamini Hochberg corrected P value for each feature
<code>wi.ep</code>	a vector containing the expected P value of the Wilcoxon test for each feature
<code>wi.eBH</code>	a vector containing the expected value of the Benjamini Hochberg corrected P value for each feature

**Author(s)**

Greg Gloor

**References**

Please use the citation given by `citation(package="ALDEX")`.

**See Also**

[aldex.clr](#), [aldex.glm](#), [aldex.effect](#), [selex](#)

**Examples**

```
# x is the output of the \code{x <- aldex.clr(data, mc.samples)} function
# conditions is a description of the data
# for the selex dataset, conditions <- c(rep("N", 7), rep("S", 7))
data(selex)
#subset for efficiency
selex <- selex[1201:1600,]
conds <- c(rep("NS", 7), rep("S", 7))
x <- aldex.clr(selex, conds, mc.samples=2, denom="all")
ttest.test <- aldex.ttest(x, conds)
```

---

getDenom	<i>getDenom</i>
----------	-----------------

---

## Description

Returns the denominator used as the basis for the log-ratio, for an `aldex.clr` object.

## Usage

```
getDenom(.object)
```

## Arguments

<code>.object</code>	A <code>aldex.clr</code> object containing the Monte Carlo Dirichlet instances derived from estimating the technical variance of the raw read count data, along with sample and feature information.
----------------------	--

## Details

Returns the denominator used to calculate the log-ratios. "all" is the centred log-ratio. "iqlr" is the interquartile log-ratio. A vector of numbers is the offset of the variables used in the denominator

## Value

A vector of values.

## See Also

`aldex.clr`

## Examples

```
data(selex)
  #subset for efficiency
  selex <- selex[1201:1600,]
conds <- c(rep("NS", 7), rep("S", 7))
x <- aldex.clr(selex, conds, mc.samples = 2, denom = "iqlr", verbose = FALSE)
Denom <- getDenom(x)

# to find the names of housekeeping genes used
getFeatureNames(x)[getDenom(x)]
```



---

getFeatureNames	<i>getFeatureNames</i>
-----------------	------------------------

---

**Description**

Returns the names of the features as a vector, for an `aldex.clr` object.

**Usage**

```
getFeatureNames(.object)
```

**Arguments**

<code>.object</code>	A <code>aldex.clr</code> object containing the Monte Carlo Dirichlet instances derived from estimating the technical variance of the raw read count data, along with sample and feature information.
----------------------	--

**Details**

Returns the names of the keys that can be used to subset the data rows. The keys values are the `rsid`'s.

**Value**

A vector of feature names.

**See Also**

`aldex.clr`

**Examples**

```
data(selex)
#subset for efficiency
selex <- selex[1201:1600,]
conds <- c(rep("NS", 7), rep("S", 7))
x <- aldex.clr(selex, conds, mc.samples = 2, denom="all", verbose = FALSE)
featureNames <- getFeatureNames(x)
```

---

getFeatures	<i>getFeatures</i>
-------------	--------------------

---

**Description**

Returns the features as a vector, for an `aldex.clr` object.

**Usage**

```
getFeatures(.object)
```

**Arguments**

`.object` A `aldex.clr` object containing the Monte Carlo Dirichlet instances derived from estimating the technical variance of the raw read count data, along with sample and feature information.

**Details**

Returns the features as a vector, for an `aldex.clr` object.

**Value**

A vector of features.

**See Also**

`aldex.clr`

**Examples**

```
data(selex)
  #subset for efficiency
  selex <- selex[1201:1600,]
conds <- c(rep("NS", 7), rep("S", 7))
x <- aldex.clr(selex, conds, mc.samples = 2, denom="all", verbose = FALSE)
features <- getFeatures(x)
```

---

`getMonteCarloInstances`

*getMonteCarloInstances*

---

**Description**

Returns the Monte Carlo Dirichlet instances used to create an `aldex.clr` object.

**Usage**

```
getMonteCarloInstances(.object)
```

**Arguments**

`.object` A `aldex.clr` object containing the Monte Carlo Dirichlet instances derived from estimating the technical variance of the raw read count data, along with sample and feature information.

**Details**

Returns the Monte Carlo Dirichlet instances used to create an `aldex.clr` object.

**Value**

A list of data frames of Monte Carlo Dirichlet instances derived from estimating the technical variance of the raw read count data.

**See Also**`aldex.clr`**Examples**

```
data(selex)
  #subset for efficiency
  selex <- selex[1201:1600,]
conds <- c(rep("NS", 7), rep("S", 7))
x <- aldex.clr(selex, conds, mc.samples = 2, denom = "all", verbose = FALSE)
monteCarloInstances <- getMonteCarloInstances(x)
```

---

`getMonteCarloReplicate`*getMonteCarloReplicate*

---

**Description**

Returns the designated Monte Carlo Dirichlet replicate generated from analysis, for an `aldex.clr` object.

**Usage**

```
getMonteCarloReplicate(.object, i)
```

**Arguments**

<code>.object</code>	A <code>aldex.clr</code> object containing the Monte Carlo Dirichlet instances derived from estimating the technical variance of the raw read count data, along with sample and feature information.
<code>i</code>	The numeric index of the desired replicate.

**Details**

Returns the designated Monte Carlo Dirichlet replicate generated from analysis.

**Value**

A data frame representing the designated Monte Carlo Dirichlet replicate generated from analysis.

**See Also**`aldex.clr`**Examples**

```
data(selex)
  #subset for efficiency
  selex <- selex[1201:1600,]
conds <- c(rep("NS", 7), rep("S", 7))
x <- aldex.clr(selex, conds, mc.samples = 2, denom = "all", verbose = FALSE)
monteCarloInstance <- getMonteCarloReplicate(x,1)
```

getReads

*getReads*

---

**Description**

Returns the count table used as input for analysis, for an `aldex.clr` object.

**Usage**

```
getReads(.object)
```

**Arguments**

`.object` A `aldex.clr` object containing the Monte Carlo Dirichlet instances derived from estimating the technical variance of the raw read count data, along with sample and feature information.

**Details**

Returns the count table.

**Value**

A data frame representing the count table used as input for analysis.

**See Also**

`aldex.clr`

**Examples**

```
data(selex)
#subset for efficiency
selex <- selex[1201:1600,]
conds <- c(rep("NS", 7), rep("S", 7))
x <- aldex.clr(selex, conds, mc.samples = 2, denom = "all", verbose = FALSE)
reads <- getReads(x)
```

---

getSampleIDs*getSampleIDs*

---

**Description**

Returns the names of the samples for an `aldex.clr` object. These can be used to access the original reads, as in `reads$sampleID` (if the reads are a data frame).

**Usage**

```
getSampleIDs(.object)
```

**Arguments**

`.object` A `aldex.clr` object containing the Monte Carlo Dirichlet instances derived from estimating the technical variance of the raw read count data, along with sample and feature information.

**Details**

Returns the names of the samples. These can be used to access the original reads, as in `reads$sampleID` (if the reads are a data frame).

**Value**

A vector of sample names.

**See Also**

`aldex.clr`

**Examples**

```
data(selex)
#subset for efficiency
selex <- selex[1201:1600,]
conds <- c(rep("NS", 7), rep("S", 7))
x <- aldex.clr(selex, conds, mc.samples = 2, denom = "all", verbose = FALSE)
sampleIDs <- getSampleIDs(x)
```

---

numConditions

*numConditions*

---

**Description**

Returns the number of conditions compared for analysis, for an `aldex.clr` object.

**Usage**

```
numConditions(.object)
```

**Arguments**

`.object` A `aldex.clr` object containing the Monte Carlo Dirichlet instances derived from estimating the technical variance of the raw read count data, along with sample and feature information.

**Details**

Returns the number of conditions compared.

**Value**

A numeric representing the number of conditions compared.

**See Also**

aldex.clr

**Examples**

```
data(selex)
  #subset for efficiency
  selex <- selex[1201:1600,]
conds <- c(rep("NS", 7), rep("S", 7))
x <- aldex.clr(selex, conds, mc.samples = 2, denom = "all", verbose = FALSE)
conditions <- numConditions(x)
```

---

numFeatures

*numFeatures*


---

**Description**

Returns the number of features associated with the data, for an aldex.clr object.

**Usage**

```
numFeatures(.object)
```

**Arguments**

<code>.object</code>	A aldex.clr object containing the Monte Carlo Dirichlet instances derived from estimating the technical variance of the raw read count data, along with sample and feature information.
----------------------	---

**Details**

Returns the number of features associated with the data.

**Value**

A numeric representing the number of features associated with the data.

**See Also**

aldex.clr

**Examples**

```
data(selex)
  #subset for efficiency
  selex <- selex[1201:1600,]
conds <- c(rep("NS", 7), rep("S", 7))
x <- aldex.clr(selex, conds, mc.samples = 2, denom = "all", verbose = FALSE)
numFeatures <- numFeatures(x)
```

---

numMCInstances	<i>numMCInstances</i>
----------------	-----------------------

---

**Description**

Returns the number of Monte Carlo Dirichlet instances generated for analysis, for an `aldex.clr` object.

**Usage**

```
numMCInstances(.object)
```

**Arguments**

<code>.object</code>	A <code>aldex.clr</code> object containing the Monte Carlo Dirichlet instances derived from estimating the technical variance of the raw read count data, along with sample and feature information.
----------------------	--

**Details**

Returns the number of Monte Carlo Dirichlet instances generated for analysis.

**Value**

A numeric representing the number of Monte Carlo Dirichlet instances generated for analysis.

**See Also**

`aldex.clr`

**Examples**

```
data(selex)
#subset for efficiency
selex <- selex[1201:1600,]
conds <- c(rep("NS", 7), rep("S", 7))
x <- aldex.clr(selex, conds, mc.samples = 2, denom = "all", verbose = FALSE)
numInstances <- numMCInstances(x)
```

---

selex	<i>Selection-based differential sequence variant abundance dataset</i>
-------	--

---

**Description**

This data set gives the differential abundance of 1600 enzyme variants grown under selective (NS) and selective (S) conditions

**Usage**

```
selex
```

**Format**

A dataframe of 1600 features and 14 samples. The first 7 samples are non-selected, the last 7 are selected.

**Source**

McMurrough et al (2014) PNAS doi:10.1073/pnas.1322352111

**References**

McMurrough et al (2014) PNAS doi:10.1073/pnas.1322352111

---

synth2

*Synthetic asymmetric dataset*

---

**Description**

This synthetic dataset contains 2 percent sparsity as 0 values asymmetrically distributed. It is used as a test dataset.

**Usage**

selex

**Format**

A dataframe of 1000 features and 16 samples. The first 8 samples contain 20 features set to 0, the last 8 samples contain counts.

**Source**

Gloor et al (2017) notes



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