Package 'msdata'

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Title Various Mass Spectrometry raw data example files

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Depends R (>= 2.10)

Suggests xcms, mzR, MSnbase

ZipData no

Description Ion Trap positive ionization mode data in mzData file
format. Subset from 500-850 m/z and 1190-1310 seconds, incl. MS2
and MS3, intensity threshold 100.000. Extracts from FTICR Apex III,
m/z 400-450. Subset of UPLC - Bruker micrOTOFq data, both mzData,
mzML and mz5. LC-MSMS and MRM files from proteomics experiments. PSI
mzIdentML example files for various search engines.

biocViews ExperimentData, MassSpectrometryData

License GPL (>= 2)

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R topics documented:

msdata	*Sample FTICR, LC/MS and MS^n data*

Description

x object containing a subset of LC/MS raw data from a Thermo Finnigan LCQ Deca XP The data is a subset from 500-850 m/z and 1190-1310 seconds, incl. MS2 and MS3, intensity threshhold 100.000. It was collected in positive ionization mode.

xs object containing a subset of FTICR data from a Bruker APex III FTICR. The data is a subset from 400-450 m/z, collected in positive ionization mode.

Usage

```
data(xs)
```

Format

The format is:

```
xs
```

Details

The corresponding raw mzdata files are located in the `fticr` and `iontrap` subdirectory of this package.

See Also

`xcmsSet`, `xcmsRaw`

Examples

```
## The directory with the mzData LC/MS files
data(xs)
mzdatapath <- file.path(find.package("msdata"), "iontrap")
mzdatapath
files <- list.files(mzdatapath, recursive = TRUE, full.names = TRUE)
files

if (require(xcms)) {

## xcmsSet Summary
show(xs)

## Access raw data file
x <- xcmsRaw(files[1])
x

}
```

proteomics	*Proteomics data in* msdata

Description

This function returns proteomics mass spectrometry files. These files are all stored in the proteomics directory in the msdata package. Each file/data is described in more details below.

Usage

```
proteomics(...)
```

Arguments

...	Additional arguments passed to [list.files]().

Details

- TMT_Erwinia_1uLSike_Top10HCD_isol2_45stepped_60min_01.mzML.gz: A LC-MSMS data file containing iTRAQ 4-plex data. The data is described in more details in Gatto L. and Christoforou A. *Using R and Bioconductor for proteomics data analysis* (PMID 23692960). This file only contains a subset of the fill data (spectra 1002 to 1510) and was generated from the full data using msconvert (ProteoWizard release: 3.0.9283 (2016-1-11)) using following command

  msconvert TMT_Erwinia_1uLSike_Top10HCD_isol2_45stepped_60min_01-20141210.mzML --filter "index [1002,1510]" -o subset

  The complete file is TMT_Erwinia_1uLSike_Top10HCD_isol2_45stepped_60min_01-20141210.mzML.gz, also available here, and can also be downloaded from the ProteomeXchange PXD000001 project (see the rpx package).

  An MS2 identification file, ident/TMT_Erwinia_1uLSike_Top10HCD_isol2_45stepped_60min_01-20141210.m generated searching the raw data against the *Erwinia carotovora* database (see reference above) is also available through the ident function.

- MS3TMT10_01022016_32917-33481.mzML.gz:A subset of 565 spectra from a currently unpublished TMT 10-plex experiment run on an Thermo Orbitrap Lumos with synchronous precursor selection (SPS) MS3. Only the MS2 spectra were centroided during convertion using msconvert (ProteoWizard release: 3.0.9283 (2016-1-11)) using vendor libraries.

- MS3TMT11.mzML:A subset of 994 spectra from a currently unpublished MS3 SPS TMT 11-plex experiment converted to mzML using msconvert. The file contains 30, 482 and 482 MS1, MS2 and MS3 spectra, respectively. The MS1 spectra are in profile mode; other MS levels are centroided. See *Sensitive and Accurate Quantitation of Phosphopeptides Using TMT Isobaric Labeling Technique* for details about the acquisition method.

  An feature data containing identification data is available with data(fdms3tmt11), which can be used to directly update the feature data, as shown in the example below.

- MRM-standmix-5.mzML.gz:Sample from mouse brain acquired by HILIC ESI-QqQ/MS in Dynamic multiple reaction monitoring mode (MRM). HPLC system was a 1290 Infinity (Agilent Technologies) coupled to ion-Funnel Triple quadrupole 6490 mass spectrometer (Agilent Technologies). This file was contributed by Xavi Domingo-Almenara from theThe Scripps Research Institute, San Diego, CA.

Value

A `character` with file names.

Author(s)

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See Also

For more access to mass spectrometry-based proteomics data, see the `rpx` and `ProteomicsAnnotationHubData` packages.

Examples

```
(f <- proteomics(full.names = TRUE))

library("mzR")
openMSfile(f[2])


library("MSnbase")
## The MS3 TMT11 raw data
(fms3 <- proteomics(full.names = TRUE, pattern = "MS3TMT11.mzML"))
ms3 <- readMSData(fms3, mode = "onDisk")
ms3

## Additional feature metadata
data(fdms3tmt11)
names(fdms3tmt11)

fData(ms3) <- fdms3tmt11
validObject(ms3)
```

---

  sciexdata   *AB Sciex LC-MS data files*

Description

The mzML files in the `sciex` directory in the `msdata` package represent profile-mode LC-MS data of pooled human serum samples (the same pool being measured). The samples were analyzed by ultra high-performance liquid chromatography (UHPLC; Agilent 1290) coupled to a Q-TOF mass spectrometer (TripleTOF 5600+ AB Sciex). The chromatographic separation was based in hydrophilic interaction liquid chromatography (HILIC) and performed using an Waters Acquity BEH Amide, 100 x 2.1 mm column.

The mass specctrometer was operated in full scan mode in the mass range from 50 to 1000 m/z and with an accumulation time of 250 ms. The files represent a subset of spectra/scans from m/z 105 to 134 and from retention time 0 to 260 seconds. The files were generated in the same LC-MS run, but from different injections. Details on the individual files are provided below.

Details

- 20171016_POOL_POS_1_105-134.mzML profile-mode LC-MS data of pooled human serum samples. Injection index: 1.

- 20171016_POOL_POS_3_105-134.mzML profile-mode LC-MS data of pooled human serum samples. Injection index: 19.

Author(s)

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Examples

```
## List the files in the sciex folder
dir(system.file("sciex", package = "msdata"))
```

Index