

# Package ‘MCbiclust’

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**Type** Package

**Title** Massive correlating biclusters for gene expression data and associated methods

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**Description** Custom made algorithm and associated methods for finding, visualising and analysing biclusters in large gene expression data sets. Algorithm is based on with a supplied gene set of size  $n$ , finding the maximum strength correlation matrix containing  $m$  samples from the data set.

**Depends** R (>= 3.4)

**Imports** BiocParallel, graphics, utils, stats, AnnotationDbi, GO.db, org.Hs.eg.db, GGally, ggplot2, scales, cluster, WGCNA

**Suggests** gplots, knitr, rmarkdown, BiocStyle, gProfileR, MASS, dplyr, pander, devtools, testthat, GSVA

**License** GPL-2

**biocViews** ImmunoOncology, Clustering, Microarray, StatisticalMethod, Software, RNASeq, GeneExpression

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**VignetteBuilder** knitr

**LazyData** true

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CCLE_samples	<i>Clinical information for CCLE data</i>
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### Description

A dataset containing clinical information for the CCLE samples.

### Usage

CCLE\_samples

### Format

A data frame with 967 rows and 14 variables:

- CCLE.name: Sample name identifier.
- Cell.line.primary.name: Cell line name.
- Cell.line.aliases: Any known aliases of cell line.
- Gender: Gender of patient cell line derived from.
- Site.Primary: Primary site cell line derived from.
- Histology: Histology of tumour cell line derived from.
- Hist.Subtype1: Histology subtype of tumour cell line derived from.
- Notes: Additional notes.
- Source: Source of the cell line.
- Expression.arrays: Expression array used.

- SNP.arrays: SNP array used.
- Oncomap: Oncomap mutation array used.
- Hybrid.Capture.Sequencing: Hybrid capture sequencing used.
- Name: Sample name identifier

**Value**

NA

**Source**

<http://www.broadinstitute.org/ccle/data/browseData> Filename: CCLE\_sample\_info\_file\_2012-04-06.txt

---

CCLE\_small

*Subset of expression levels of CCLE data*

---

**Description**

A dataset containing the gene-centric RMA-normalized mRNA expression data for nearly 1000 genes and 500 samples taken as a random subset of the complete CCLE data. 1000 genes were selected randomly such that 500 were mitochondrial and 500 non-mitochondrial.

**Usage**

CCLE\_small

**Format**

A data frame with 1000 rows and 500 variables:

- MKN74\_STOMACH: mRNA expression on sample MKN74\_STOMACH
- OC316\_OVARY: mRNA expression on sample OC316\_OVARY
- ...

@source <http://www.broadinstitute.org/ccle/data/browseData> Filename: CCLE\_Expression\_Entrez\_2012-04-06.gct.gz

**Value**

NA

---

CorScoreCalc

*Calculate correlation score*

---

### Description

The standard method to calculate the correlation score used to judge biclusters in MCBiclust

### Usage

```
CorScoreCalc(gene.expr.matrix, sample.vec)
```

### Arguments

gene.expr.matrix      Gene expression matrix with genes as rows and samples as columns  
sample.vec            Vector of samples

### Value

The correlation score

### Examples

```
data(CCLE_small)
data(Mitochondrial_genes)

mito.loc <- which(row.names(CCLE_small) %in% Mitochondrial_genes)
CCLE.mito <- CCLE_small[mito.loc,]

random.seed <- sample(seq(length = dim(CCLE.mito)[2]),10)
CCLE.seed <- FindSeed(gem = CCLE.mito,
                     seed.size = 10,
                     iterations = 100,
                     messages = 100)

CorScoreCalc(CCLE.mito, random.seed)
CorScoreCalc(CCLE.mito, CCLE.seed)

CCLE.hicor.genes <- as.numeric(HclustGenesHiCor(CCLE.mito,
                                              CCLE.seed,
                                              cuts = 8))

CorScoreCalc(CCLE.mito[CCLE.hicor.genes,], CCLE.seed)
```

---

CVEval

*Method for the calculation of a correlation vector*

---

## Description

Upon identifying a bicluster seed with FindSeed, one of the next steps is to identify which genes not in your chosen gene set are also highly correlated to the bicluster found. This is done by CVEval, and the output is known as the correlation vector.

## Usage

```
CVEval(gem.part, gem.all, seed, splits)
```

## Arguments

gem.part	Part of gene expression matrix only containing gene set of interest with genes as rows and samples as columns
gem.all	All of gene expression matrix
seed	Seed of highly correlating samples
splits	Number of cuts from hierarchical clustering

## Details

CVEval uses hierarchical clustering to select the genes most representative of the bicluster and then uses the average expression of these genes across the sample seed and calculates the correlation of every gene measured across the sample seed to this average expression value.

The correlation vector is the output of this calculation.

## Value

Correlation vector

## Examples

```
data(CCLE_small)
data(Mitochondrial_genes)

mito.loc <- (row.names(CCLE_small) %in% Mitochondrial_genes)
CCLE.mito <- CCLE_small[mito.loc,]

set.seed(102)
CCLE.seed <- FindSeed(gem = CCLE.mito,
                     seed.size = 10,
                     iterations = 100,
                     messages = 1000)

CCLE.sort <- SampleSort(gem = CCLE.mito, seed = CCLE.seed, sort.length = 11)
```

```

# Full ordering are in Vignette_sort in sysdata.rda
CCLE.samp.sort <- MCbiclust:::Vignette_sort[[1]]

CCLE.pc1 <- PC1VecFun(top.gem = CCLE.mito,
                    seed.sort = CCLE.samp.sort,
                    n = 10)

CCLE.cor.vec <- CVEval(gem.part = CCLE.mito,
                     gem.all = CCLE_small,
                     seed = CCLE.seed,
                     splits = 10)

CCLE.bic <- ThresholdBic(cor.vec = CCLE.cor.vec, sort.order = CCLE.samp.sort,
                       pc1 = as.numeric(CCLE.pc1))

CCLE.pc1 <- PC1Align(gem = CCLE_small, pc1 = CCLE.pc1,
                    cor.vec = CCLE.cor.vec,
                    sort.order = CCLE.samp.sort,
                    bic = CCLE.bic)

CCLE.fork <- ForkClassifier(CCLE.pc1, samp.num = length(CCLE.bic[[2]]))

```

---

CVPlot

---

*Make correlation vector plot*


---

## Description

A function to visualise the differences between different found biclusters. Output is a matrix of plots. Each correlation vector is plotted against each other across the entire measured gene set in the lower diagonal plots, and a chosen gene set (e.g. mitochondrial) in the upper diagonal plots. The diagonal plots themselves show the density plots of the entire measured and chosen gene set. There are addition options to set the transparency of the data points and names of the correlation vectors.

## Usage

```
CVPlot(cv.df, geneset.loc, geneset.name, alpha1 = 0.005, alpha2 = 0.1,
       cnames = NULL)
```

## Arguments

cv.df	A dataframe containing the correlation vectors of one or more patterns.
geneset.loc	A gene set of interest (e.g. mitochondrial) to be plotted separately from rest of genes.
geneset.name	Name of geneset (e.g. mitochondrial genes)
alpha1	Transparency level of non-gene set genes



**Arguments**

gem	Gene expression matrix with genes as rows and samples as columns
seed.size	Size of sample seed
iterations	Number of iterations
initial.seed	Initial seed used, if NULL randomly chosen
messages	frequency of progress messages

**Details**

Additional options allow for the search to start at a chosen seed, for instance if a improvement to a known seed is desired. The result of FindSeed() is dependent on the number of iterations, with above 1000 usually providing a good seed, and above 10000 an optimum seed.

**Value**

Highly correlated seed

**Examples**

```
data(CCLE_small)
data(Mitochondrial_genes)

mito.loc <- which(row.names(CCLE_small) %in% Mitochondrial_genes)
CCLE.mito <- CCLE_small[mito.loc,]

random.seed <- sample(seq(length = dim(CCLE.mito)[2]),10)
CCLE.seed <- FindSeed(gem = CCLE.mito,
                     seed.size = 10,
                     iterations = 100,
                     messages = 100)

CorScoreCalc(CCLE.mito, random.seed)
CorScoreCalc(CCLE.mito, CCLE.seed)

CCLE.hicor.genes <- as.numeric(HclustGenesHiCor(CCLE.mito,
                                              CCLE.seed,
                                              cuts = 8))

CorScoreCalc(CCLE.mito[CCLE.hicor.genes,], CCLE.seed)
```



## Description

The Mann-Whitney test is typically used due to the values of the correlation vector, not being normally distributed. GOEnrichmentAnalysis provides an interface with the GO database annotation to find the most significant GO terms.

## Usage

```
GOEnrichmentAnalysis(gene.names, gene.values, sig.rate)
```

## Arguments

gene.names	Names of the genes in standard gene name format.
gene.values	Values associated with the genes, e.g the correlation vector output of CVEval.
sig.rate	Level of significance required after multiple hypothesis adjustment.

## Value

Data frame of the significant gene sets, with GOID, GO Term, number of genes, number of genes in GO Term, number of genes in GO Term also in gene set, adjusted p-value, average value of correlation vector in gene set and phenotype describing whether average value of correlation vector is above or below the total average.

## Examples

```
data(CCLE_small)
data(Mitochondrial_genes)

mito.loc <- (row.names(CCLE_small) %in% Mitochondrial_genes)
CCLE.mito <- CCLE_small[mito.loc,]

set.seed(101)
CCLE.seed <- FindSeed(gem = CCLE.mito,
                     seed.size = 10,
                     iterations = 100,
                     messages = 100)

CCLE.cor.vec <- CVEval(gem.part = CCLE.mito,
                     gem.all = CCLE_small,
                     seed = CCLE.seed, splits = 10)

# Significant GO terms can be calculated as follows:
# GEA <- GOEnrichmentAnalysis(gene.names = row.names(CCLE_small),
#                             #
#                             gene.values = CCLE.cor.vec,
#                             #
#                             sig.rate = 0.05)
```

---

HclustGenesHiCor      *Find the most highly correlated genes using hierarchical clustering*

---

### Description

Upon finding an initial bicluster with FindSeed() not all the genes in the chosen geneset will be highly correlated to the bicluster. HclustGenesHiCor() uses the output of FindSeed() and hierarchical clustering to only select the genes that are most highly correlated to the bicluster. This is achieved by cutting the dendrogram produced from the clustering into a set number of groups and then only selecting the groups that are most highly correlated to the bicluster

### Usage

```
HclustGenesHiCor(gem, seed, cuts)
```

### Arguments

gem	Gene expression matrix with genes as rows and samples as columns
seed	Seed of highly correlating samples
cuts	Number of groups to cut dendrogram into

### Value

Numeric vector of most highly correlated genes

### Examples

```
data(CCLE_small)
data(Mitochondrial_genes)

mito.loc <- which(row.names(CCLE_small) %in% Mitochondrial_genes)
CCLE.mito <- CCLE_small[mito.loc,]

random.seed <- sample(seq(length = dim(CCLE.mito)[2]),10)
CCLE.seed <- FindSeed(gem = CCLE.mito,
                     seed.size = 10,
                     iterations = 100,
                     messages = 100)

CorScoreCalc(CCLE.mito, random.seed)
CorScoreCalc(CCLE.mito, CCLE.seed)

CCLE.hicor.genes <- as.numeric(HclustGenesHiCor(CCLE.mito,
                                              CCLE.seed,
                                              cuts = 8))

CorScoreCalc(CCLE.mito[CCLE.hicor.genes,], CCLE.seed)
```

---

MCbiclust

*MCbiclust: Massively Correlated biclustering*

---

### Description

MCbiclust is a R package for running massively correlating biclustering analysis. MCbiclust aims to find large scale biclusters with selected features being highly correlated with each other over a subset of samples.

### Details

The package was originally designed in order to solve a problem in bioinformatics: to find biclusters representing different modes of regulation of mitochondria gene expression in disease states such as breast cancer. The same methods however, can be used on any gene expression data set to find biclusters of interest.

To learn more about MCbiclust, start with the vignette: `browseVignettes(package = "MCbiclust")`

---

Mitochondrial\_genes

*List of known mitochondrial genes*

---

### Description

A dataset from MitoCarta1.0 containing the 1023 mitochondrial genes Available from the broad institute: <http://www.broadinstitute.org/scientific-community/science/programs/metabolic-disease-program/publications/mitocarta/mitocarta-in-0>

### Usage

Mitochondrial\_genes

### Format

A Character vector of the HGNC approved gene names

### Value

NA

### Source

<https://www.broadinstitute.org/publications/broad807s>

---

PC1VecFun

*Calculate PC1 vector of found pattern*


---

**Description**

The correlations found between the chosen geneset in a subset of samples can be summarised by looking at the first principal component (PC1) using principal component analysis (PCA).

**Usage**

```
PC1VecFun(top.gem, seed.sort, n)
```

**Arguments**

top.gem	Gene expression matrix containing only highly correlating genes
seed.sort	Ordering of samples according to strength of correlation
n	Number of samples to use in calculation of PC1

**Details**

PC1VecFun() takes a gene expression matrix and the sample ordering and fits a PC1 value to all the samples based on a PCA analysis done on the first n samples.

**Value**

PC1 value for each sample

**Examples**

```
data(CCLE_small)
data(Mitochondrial_genes)

mito.loc <- (row.names(CCLE_small) %in% Mitochondrial_genes)
CCLE.mito <- CCLE_small[mito.loc,]

set.seed(102)
CCLE.seed <- FindSeed(gem = CCLE.mito,
                     seed.size = 10,
                     iterations = 100,
                     messages = 1000)

CCLE.sort <- SampleSort(gem = CCLE.mito, seed = CCLE.seed, sort.length = 11)

# Full ordering are in Vignette_sort in sysdata.rda
CCLE.samp.sort <- MCbiclust:::Vignette_sort[[1]]

CCLE.pc1 <- PC1VecFun(top.gem = CCLE.mito,
                    seed.sort = CCLE.samp.sort,
                    n = 10)
```

```

CCLE.cor.vec <- CVEval(gem.part = CCLE.mito,
                      gem.all = CCLE_small,
                      seed = CCLE.seed,
                      splits = 10)

CCLE.bic <- ThresholdBic(cor.vec = CCLE.cor.vec, sort.order = CCLE.samp.sort,
                       pc1 = as.numeric(CCLE.pc1))

CCLE.pc1 <- PC1Align(gem = CCLE_small, pc1 = CCLE.pc1,
                    cor.vec = CCLE.cor.vec ,
                    sort.order = CCLE.samp.sort,
                    bic = CCLE.bic)

CCLE.fork <- ForkClassifier(CCLE.pc1, samp.num = length(CCLE.bic[[2]]))

```

---

PointScoreCalc

*Calculate PointScore*


---

### Description

Using two gene sets that are represented of a known bicluster (one gene set being up regulated while other gene set is down regulated), samples are scored based on how well they match the known regulation of the bicluster.

### Usage

```
PointScoreCalc(gene.expr.matrix, gene.loc1, gene.loc2)
```

### Arguments

gene.expr.matrix	Gene expression matrix with genes as rows and samples as columns
gene.loc1	Location of the rows containing the genes in gene set 1 within the gene expression matrix
gene.loc2	Location of the rows containing the genes in gene set 2 within the gene expression matrix

### Details

The PointScore of a sample can be directly compared to the PC1 value. The PointScore is typically used to identify samples related to the upper/lower fork of a bicluster without running the complete main MCBiclust pipeline on a dataset.

### Value

Vector of point scores for each sample in the gene expression matrix

**Examples**

```

data(CCLE_small)
data(Mitochondrial_genes)

mito.loc <- (row.names(CCLE_small) %in% Mitochondrial_genes)
CCLE.mito <- CCLE_small[mito.loc,]

set.seed(102)
CCLE.seed <- FindSeed(gem = CCLE.mito,
                      seed.size = 10,
                      iterations = 100,
                      messages = 1000)

CCLE.sort <- SampleSort(gem = CCLE.mito, seed = CCLE.seed, sort.length = 11)

# Full ordering are in Vignette_sort in sysdata.rda
CCLE.samp.sort <- MCBiclust:::Vignette_sort[[1]]

CCLE.pc1 <- PC1VecFun(top.gem = CCLE.mito,
                     seed.sort = CCLE.samp.sort,
                     n = 10)

CCLE.hicor.genes <- as.numeric(HclustGenesHiCor(CCLE.mito,
                                               CCLE.seed,
                                               cuts = 8))

CCLE.cor.mat <- cor(t(CCLE.mito[CCLE.hicor.genes, CCLE.seed]))

gene.set1 <- labels(as.dendrogram(hclust(dist(CCLE.cor.mat))))[[1]]
gene.set2 <- labels(as.dendrogram(hclust(dist(CCLE.cor.mat))))[[2]]

gene.set1.loc <- which(row.names(CCLE.mito) %in% gene.set1)
gene.set2.loc <- which(row.names(CCLE.mito) %in% gene.set2)

ps.vec <- PointScoreCalc(CCLE.mito, gene.set1.loc, gene.set2.loc)

cor(ps.vec[CCLE.samp.sort], CCLE.pc1)
plot(ps.vec[CCLE.samp.sort])
plot(CCLE.pc1)

```

**Description**

After finding an initial bicluster with `FindSeed()` the next step is to extend the bicluster by ordering the remaining samples by how they preserve the correlation found.

**Usage**

```
SampleSort(gem, seed, num.cores = 1, sort.length = NULL)
```

```
MultiSampleSortPrep(gem, av.corvec, top.genes.num, groups, initial.seeds)
```

**Arguments**

<code>gem</code>	Gene expression matrix with genes as rows and samples as columns
<code>seed</code>	Sample seed of highly correlating genes
<code>num.cores</code>	Number of cores used in parallel evaluation
<code>sort.length</code>	Number of samples to be sorted
<code>av.corvec</code>	List of average correlation vector
<code>top.genes.num</code>	Number of the top genes in correlation vector to use for sorting samples
<code>groups</code>	List showing what runs belong to which correlation vector group
<code>initial.seeds</code>	List of sample seeds from all runs

**Details**

`SampleSort()` is the basic function that achieves this, it takes the gene expression matrix, seed of samples, and also has options for the number of cores to run the method on and the number of samples to sort.

`MultiSampleSortPrep()` is a preparation function for `SampleSort()` when `MCbiclust` has been run multiple times and returns a list of gene expression matrices and seeds for each ‘distinct’ bi-cluster found.

**Value**

Order of samples by strength to correlation pattern

**Examples**

```
data(CCLE_small)
data(Mitochondrial_genes)

mito.loc <- (row.names(CCLE_small) %in% Mitochondrial_genes)
CCLE.mito <- CCLE_small[mito.loc,]

set.seed(102)
CCLE.seed <- FindSeed(gem = CCLE.mito,
                     seed.size = 10,
                     iterations = 100,
                     messages = 1000)

CCLE.sort <- SampleSort(gem = CCLE.mito, seed = CCLE.seed, sort.length = 11)

# Full ordering are in Vignette_sort in sysdata.rda
CCLE.samp.sort <- MCbiclust:::Vignette_sort[[1]]
```

```

CCLE.pc1 <- PC1VecFun(top.gem = CCLE.mito,
                    seed.sort = CCLE.samp.sort,
                    n = 10)

CCLE.cor.vec <- CVEval(gem.part = CCLE.mito,
                    gem.all = CCLE_small,
                    seed = CCLE.seed,
                    splits = 10)

CCLE.bic <- ThresholdBic(cor.vec = CCLE.cor.vec, sort.order = CCLE.samp.sort,
                    pc1 = as.numeric(CCLE.pc1))

CCLE.pc1 <- PC1Align(gem = CCLE_small, pc1 = CCLE.pc1,
                    cor.vec = CCLE.cor.vec ,
                    sort.order = CCLE.samp.sort,
                    bic = CCLE.bic)

CCLE.fork <- ForkClassifier(CCLE.pc1, samp.num = length(CCLE.bic[[2]]))

```

---

SilhouetteClustGroups *Silhouette validation of correlation vector clusters*

---

### Description

MCbiclust is a stochastic method and needs to be run multiple times to identify different biclusters. SilhouetteClustGroups() examines the correlation vectors calculated from different runs and uses the technique of examining silhouette widths to identify the number of distinct clusters (and hence biclusters) found.

### Usage

```
SilhouetteClustGroups(cor.vec.mat, max.clusters, plots = FALSE, seed1 = 100,
                    rand.vec = TRUE)
```

### Arguments

cor.vec.mat	Correlation matrix of the correlation vectors (CVs)
max.clusters	Maximum number of clusters to divide CVs into
plots	True or False for whether to show silhouette plots
seed1	Value used to set random seed
rand.vec	True or False for whether to add random correlation vector used for comparison

### Value

The distinct clusters of correlation vectors



**Examples**

```

data(CCLE_small)
data(Mitochondrial_genes)

mito.loc <- (row.names(CCLE_small) %in% Mitochondrial_genes)
CCLE.mito <- CCLE_small[mito.loc,]

CCLE.seed <- list()
CCLE.cor.vec <- list()

for(i in 1:5){
  set.seed(i)
  CCLE.seed[[i]] <- FindSeed(gem = CCLE.mito,
                            seed.size = 10,
                            iterations = 100,
                            messages = 100)}

for(i in 1:5){
  CCLE.cor.vec[[i]] <- CVEval(gem.part = CCLE.mito,
                             gem.all = CCLE_small,
                             seed = CCLE.seed[[i]],
                             splits = 10)}

CCLE.cor.mat <- as.matrix(as.data.frame(CCLE.cor.vec))

CCLE.clust.groups <- SilhouetteClustGroups(cor.vec.mat = CCLE.cor.mat,
                                           plots = TRUE,
                                           max.clusters = 10)

av.corvec.fun <- function(x) rowMeans(CCLE.cor.mat[,x])
CCLE.average.corvec <- lapply(X = CCLE.clust.groups,
                              FUN = av.corvec.fun)

multi.sort.prep <- MultiSampleSortPrep(gem = CCLE_small,
                                       av.corvec = CCLE.average.corvec,
                                       top.genes.num = 750,
                                       groups = CCLE.clust.groups,
                                       initial.seeds = CCLE.seed)

multi.sort <- list()
for(i in seq_len(length(CCLE.clust.groups))){
  multi.sort[[i]] <- SampleSort(multi.sort.prep[[1]][[i]],
                               seed = multi.sort.prep[[2]][[i]],
                               sort.length = 11)
}

```

**Description**

A bicluster is the fundamental result found using `MCbiclust`. These three functions are essential for the precise definition of these biclusters.

**Usage**

```
ThresholdBic(cor.vec, sort.order, pc1, samp.sig = 0)
```

```
PC1Align(gem, pc1, cor.vec, sort.order, bic)
```

```
ForkClassifier(pc1, samp.num)
```

**Arguments**

<code>cor.vec</code>	Correlation vector (output of <code>CVEval()</code> ).
<code>sort.order</code>	Order of samples (output of <code>SampleSort()</code> ).
<code>pc1</code>	PC1 values for samples (output of <code>PC1VecFun</code> ).
<code>samp.sig</code>	Value between 0 and 1 determining number of samples in bicluster
<code>gem</code>	Gene expression matrix containing genes as rows and samples as columns.
<code>bic</code>	bicluster (output of <code>ThresholdBic()</code> )
<code>samp.num</code>	Number of samples in the bicluster

**Details**

`ThresholdBic()` takes as its main inputs the correlation vector (output of `CVEval()`), sample ordering (output of `SampleSort()`), PC1 vector (output of `PC1VecFun`) and returns a list of the genes and samples which belong to the bicluster according to a certain level of significance.

`PC1Align()` is a function used once the bicluster has been found to ensure that the upper fork samples (those with higher PC1 values) correspond to those samples that have genes with positive correlation vector values up-regulated.

`ForkClassifier()` is a function used to classify which samples are in the upper or lower fork.

**Value**

Defined bicluster

**Examples**

```
data(CCLE_small)
data(Mitochondrial_genes)

mito.loc <- (row.names(CCLE_small) %in% Mitochondrial_genes)
CCLE.mito <- CCLE_small[mito.loc,]

set.seed(102)
CCLE.seed <- FindSeed(gem = CCLE.mito,
                     seed.size = 10,
```

```
        iterations = 100,
        messages = 1000)

CCLE.sort <- SampleSort(gem = CCLE.mito, seed = CCLE.seed, sort.length = 11)

# Full ordering are in Vignette_sort in sysdata.rda
CCLE.samp.sort <- MCbiclust:::Vignette_sort[[1]]

CCLE.pc1 <- PC1VecFun(top.gem = CCLE.mito,
                    seed.sort = CCLE.samp.sort,
                    n = 10)

CCLE.cor.vec <- CVEval(gem.part = CCLE.mito,
                    gem.all = CCLE_small,
                    seed = CCLE.seed,
                    splits = 10)

CCLE.bic <- ThresholdBic(cor.vec = CCLE.cor.vec, sort.order = CCLE.samp.sort,
                    pc1 = as.numeric(CCLE.pc1))

CCLE.pc1 <- PC1Align(gem = CCLE_small, pc1 = CCLE.pc1,
                    cor.vec = CCLE.cor.vec ,
                    sort.order = CCLE.samp.sort,
                    bic = CCLE.bic)

CCLE.fork <- ForkClassifier(CCLE.pc1, samp.num = length(CCLE.bic[[2]]))
```

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