

Package ‘genomicInstability’

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Title Genomic Instability estimation for scRNA-Seq

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Description This package contain functions to run genomic instability analysis (GIA) from scRNA-Seq data.

GIA estimates the association between gene expression and genomic location of the coding genes.

It uses the aREA algorithm to quantify the enrichment of sets of contiguous genes (loci-blocks) on the gene expression profiles and estimates the Genomic Instability Score (GIS) for each analyzed cell.

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genomicInstability-package

genomicInstability: Genomic Instability estimation for scRNA-Seq

Description

This package contain functions to run genomic instability analysis (GIA) from scRNA-Seq data. GIA estimates the association between gene expression and genomic location of the coding genes. It uses the aREA algorithm to quantify the enrichment of sets of contiguous genes (loci-blocks) on the gene expression profiles and estimates the Genomic Instability Score (GIS) for each analyzed cell.

Details

The basic functionality of this package can be performed by `inferCNV()`, to infer the enrichment of loci-blocks on gene expression; `genomicInstabilityScore()`, to estimate the genomic instability for each of the cells in the scRNASeq dataset; `giLikelihood()`, to estimate the relative likelihood for each cell to be normal (low genomic instability) or tumor (high genomic instability); `plot()` and `giDensityPlot()` to plot the scores per loci-block and the distribution of the genomic instability score, respectively.

Author(s)

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See Also

[`inferCNV()`] for estimating loci-block enrichment, [`genomicInstabilityScore()`] for estimating the genomic instability of each cell in the dataset, [`giLikelihood()`] for estimating the relative likelihood for the cells to be normal or neoplastic, [`plot.inferCNV()`] and [`giDensityPlot()`] to plot the results.

| | |
|------------|--|
| geneLength | <i>Average length of human and mouse known genes</i> |
|------------|--|

Description

A dataset containing the average length for known mouse and human genes

Usage

```
geneLength
```

Format

Vector of integers indicating the average length in bp for each gene, indicated with EntrezIDs as name argument. To access this data use:

```
data(hg38) Human
```

```
data(mm10) Mouse
```

| | |
|--------------|--|
| genePosition | <i>Chromosomal coordinate of human and mouse known genes</i> |
|--------------|--|

Description

A dataset containing the chromosomal coordinate for known human and mouse genes

Usage

```
genePosition
```

Format

data.frame with 2 columns: Chromosome and Coordinate. To access this data use:

```
data(hg38) Human
```

```
data(mm10) Mouse
```

```
generateChromosomeGeneSet
```

Topological gene sets

Description

This function generates a list of sets of k genes encoded by neighbor loci

Usage

```
generateChromosomeGeneSet(species = c("human", "mouse"), k = 100, skip = 25)
```

Arguments

| | |
|---------|---|
| species | Character string indicating the species, either human or mouse |
| k | Integer indicating the number of genes per set |
| skip | Integer indicating the displacement of the window for selecting the k genes |

Value

List of topologically-close gene sets

Examples

```
chrom_set <- generateChromosomeGeneSet('human')
length(chrom_set)
chrom_set[seq_len(2)]
```

```
genomicInstabilityScore
```

Genomic Instability Analysis

Description

This function computes the genomic instability for an object of class inferCNV

Usage

```
genomicInstabilityScore(cnv, likelihood = FALSE)
```

Arguments

| | |
|------------|---|
| cnv | Object of class inferCNV generated by inferCNV() function |
| likelihood | Logical, whether the genomic instability likelihood should be estimated |

Value

Object of class inferCNV with updated slots for gis and gisnull

See Also

[inferCNV()] to infer the enrichment of loci-blocks in the gene expression data.

Examples

```
eh <- ExperimentHub::ExperimentHub()
dset <- eh[["EH5419"]]
tpm_matrix <- SummarizedExperiment::assays(dset)$TPM
set.seed(1)
tpm_matrix <- tpm_matrix[, sample(ncol(tpm_matrix), 500)]
cnv <- inferCNV(tpm_matrix)
cnv <- genomicInstabilityScore(cnv)
plot(density(cnv$gis))
```

giDensityPlot

Genomic instability plot

Description

This function plot the genomic instability distribution, gaussian fits and null distribution if available

Usage

```
giDensityPlot(inferCNV, legend = c("topleft", "top", "topright", "none"), ...)
```

Arguments

| | |
|----------|--|
| inferCNV | Object of class inferCNV |
| legend | Character string indicating the location of the legend. none to not include it |
| ... | Additional parameters for plot() |

Value

None, a figure is created in the default output device

See Also

[giLikelihood()] to estimate the relative likelihood, [genomicInstabilityScore()] to estimate the genomic instability score for each cell in the dataset, and [inferCNV()] to infer the enrichment of loci-blocks in the gene expression data.

Examples

```
eh <- ExperimentHub::ExperimentHub()
dset <- eh[["EH5419"]]
tpm_matrix <- SummarizedExperiment::assays(dset)$TPM
set.seed(1)
tpm_matrix <- tpm_matrix[, sample(ncol(tpm_matrix), 500)]
cnv <- inferCNV(tpm_matrix)
cnv <- genomicInstabilityScore(cnv)
cnv <- giLikelihood(cnv, distros=c(3, 3), tumor=2:3)
giDensityPlot(cnv)
```

| | |
|--------------|---------------------------------------|
| giLikelihood | <i>Genomic instability likelihood</i> |
|--------------|---------------------------------------|

Description

This function computes the genomic instability likelihood

Usage

```
giLikelihood(  
  inferCNV,  
  recompute = TRUE,  
  distros = c(1, 3),  
  tumor = NULL,  
  normal = NULL  
)
```

Arguments

| | |
|-----------|--|
| inferCNV | InferCNV-class object |
| recompute | Logical, whether the model fits should be re-computed |
| distros | Vector of 2 integers indicating the minimum and maximum number of Gaussian models to fit |
| tumor | Optional vector of integers indicating the Gaussians considered as tumors |
| normal | Optional vector of integers indicating the Gaussians considered as normal. This is only useful when no null model has been provided for the analysis |

Value

Updated inferCNV-class object with `gi_likelihoood` slot

See Also

[`genomicInstabilityScore()`] to estimate the genomic instability score for each cell in the dataset, and [`inferCNV()`] to infer the enrichment of loci-blocks in the gene expression data.

Examples

```
eh <- ExperimentHub::ExperimentHub()  
dset <- eh[["EH5419"]]  
tpm_matrix <- SummarizedExperiment::assays(dset)$TPM  
set.seed(1)  
tpm_matrix <- tpm_matrix[, sample(ncol(tpm_matrix), 500)]  
cnv <- inferCNV(tpm_matrix)  
cnv <- genomicInstabilityScore(cnv)  
cnv <- giLikelihood(cnv, distros=c(3, 3), tumor=2:3)  
print(cnv$gi_fit)  
plot(density(cnv$gi_likelihoood, from=0, to=1))
```

`inferCNV`*Inference of CNV from expression data*

Description

This function estimates the CNV score based on expression data

Usage

```
inferCNV(  
  expmat,  
  nullmat = NULL,  
  species = c("human", "mouse"),  
  k = 100,  
  skip = 25,  
  min_geneset = 10,  
  verbose = TRUE  
)
```

Arguments

| | |
|--------------------------|--|
| <code>expmat</code> | Matrix of gene expression profiles or signatures with genes '(entrezID) in rows and samples in columns |
| <code>nullmat</code> | Optional matrix with same number of rows as <code>expmat</code> to be used as null model |
| <code>species</code> | Character string indicating the species, either human or mouse |
| <code>k</code> | Integer indicating the number of genes per set |
| <code>skip</code> | Integer indicating the displacement of the window for selecting the k genes |
| <code>min_geneset</code> | Integer indicating the minimum size for the genesets |
| <code>verbose</code> | Logical, whether progress should be reported |

Value

Object of class `inferCNV`, which is a list containing matrix of nes, and parameters (`param`), including species, window (`k`) and `skip`

Examples

```
eh <- ExperimentHub::ExperimentHub()  
dset <- eh[["EH5419"]]  
tpm_matrix <- SummarizedExperiment::assays(dset)$TPM  
set.seed(1)  
tpm_matrix <- tpm_matrix[, sample(ncol(tpm_matrix), 500)]  
cnv <- inferCNV(tpm_matrix)  
class(cnv)  
names(cnv)  
cnv$nes[1:5, 1:3]
```

plot.inferCNV *Plot chromosome map*

Description

This function generates a chromosomes map plot for the inferred CNVs

Usage

```
## S3 method for class 'inferCNV'  
plot(x, output = NULL, threshold = 0.2, gamma = 1.5, resolution = 150, ...)
```

Arguments

| | |
|------------|---|
| x | Object of class inferCNV |
| output | Optional output PDF file name (with extension) |
| threshold | Likelihood threshold for identifying genomically instable cells/samples, 0 disables this filter |
| gamma | Number indicating the gamma transformation for the colors |
| resolution | Integer indicating the ppi for the png and jpg output files |
| ... | Additional parameters for plot |

Value

Nothing, a plot is generated in the default output devise

See Also

[giLikelihood()] to estimate the relative likelihood, [genomicInstabilityScore()] to estimate the genomic instability score for each cell in the dataset, and [inferCNV()] to infer the enrichment of loci-blocks in the gene expression data.

Examples

```
eh <- ExperimentHub::ExperimentHub()  
dset <- eh[["EH5419"]]  
tpm_matrix <- SummarizedExperiment::assays(dset)$TPM  
set.seed(1)  
tpm_matrix <- tpm_matrix[, sample(ncol(tpm_matrix), 500)]  
cnv <- inferCNV(tpm_matrix)  
cnv <- genomicInstabilityScore(cnv)  
cnv <- giLikelihood(cnv, distros=c(3, 3), tumor=2:3)  
plot(cnv, output='test.png')
```


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