# Package 'tripr'

May 29, 2024

Type Package

Title T-cell Receptor/Immunoglobulin Profiler (TRIP)

Version 1.10.0

Description TRIP is a software framework that provides analytics services on antigen receptor (B cell receptor immunoglobulin, BcR IG | T cell receptor, TR) gene sequence data. It is a web application written in R Shiny. It takes as input the output files of the IMGT/HighV-Quest tool. Users can select to analyze the data from each of the input samples separately, or the combined data files from all samples and visualize the results accordingly.

License MIT + file LICENSE

**Encoding UTF-8** 

LazyData false

**biocViews** BatchEffect, MultipleComparison, GeneExpression, ImmunoOncology, TargetedResequencing

**Imports** shinyjs, shinyFiles, plyr, data.table, DT, stringr, stringdist, plot3D, gridExtra, RColorBrewer, plotly, dplyr, config (>= 0.3.1), golem (>= 0.3.1), methods, grDevices, graphics, stats, utils

Enhances parallel

**Suggests** BiocGenerics, shinycssloaders, tidyverse, BiocManager, Biostrings, xtable, rlist, motifStack, knitr, rmarkdown, testthat (>= 3.0.0), fs, BiocStyle, RefManageR, biocthis, pryr

**Depends** shiny (>= 1.6.0), shinyBS

Collate ``tripr-package.R" ``global.R" ``helpers.R" ``run\_TRIP\_without\_ui.R" ``app\_config.R" ``app\_server.R" ``app\_ui.R" ``run\_app.R" ``zzz.R"

URL https://github.com/BiodataAnalysisGroup/tripr

BugReports https://github.com/BiodataAnalysisGroup/tripr/issues

**BiocType** Software **RoxygenNote** 7.2.0

2 run\_app

```
VignetteBuilder knitr
Config/testthat/edition 3
git_url https://git.bioconductor.org/packages/tripr
git_branch RELEASE_3_19
git_last_commit 7bea195
git_last_commit_date 2024-04-30
Repository Bioconductor 3.19
Date/Publication 2024-05-28
Author Maria Th. Kotouza [aut],
    Katerina Gemenetzi [aut],
    Chrysi Galigalidou [aut],
    Elisavet Vlachonikola [aut],
    Nikolaos Pechlivanis [cre],
    Andreas Agathangelidis [aut],
    Raphael Sandaltzopoulos [aut],
    Pericles A. Mitkas [aut],
    Kostas Stamatopoulos [aut],
    Anastasia Chatzidimitriou [aut],
    Fotis E. Psomopoulos [aut],
    Iason Ofeidis [aut],
    Aspasia Orfanou [aut]
Maintainer Nikolaos Pechlivanis <inab.bioinformatics@lists.certh.gr>
Contents
      8
                                                                       9
Index
                     Run the Shiny Application
 run_app
Description
   Run the Shiny Application
```

# Usage

```
run_app(
 onStart = NULL,
  options = list(launch.browser = TRUE),
  enableBookmarking = NULL,
```

```
uiPattern = "/",
    ...
)
```

#### **Arguments**

onStart A function that will be called before the app is actually run. This is only needed

for shinyAppObj, since in the shinyAppDir case, a global.R file can be used

for this purpose.

options Named options that should be passed to the runApp call (these can be any of

the following: "port", "launch.browser", "host", "quiet", "display.mode" and "test.mode"). You can also specify width and height parameters which provide a hint to the embedding environment about the ideal height/width for the

app.

enableBookmarking

Can be one of "url", "server", or "disable". The default value, NULL, will re-

spect the setting from any previous calls to enableBookmarking(). See enableBookmarking()

for more information on bookmarking your app.

uiPattern A regular expression that will be applied to each GET request to determine whether

the ui should be used to handle the request. Note that the entire request path must match the regular expression in order for the match to be considered suc-

cessful.

.. arguments to pass to golem\_opts. See '?golem::get\_golem\_options' for more

details.

#### Value

None

# **Examples**

```
if (interactive()) {
    run_app(options = list(launch.browser = FALSE))
}
```

run\_TRIP

Run tripr analysis via R command line

#### **Description**

run\_TRIP() is a wrapper of {tripr} shiny analysis tool for use via R command line. Output of analysis is saved in *tripr/extdata/output* folder, where R libraries are saved (typically *R/library*).

#### Usage

```
run_TRIP(
  datapath = fs::path_package("extdata", "dataset", package = "tripr"),
  output_path = fs::path_home("Documents/tripr_output"),
  filelist = c("1_Summary.txt", "2_IMGT-gapped-nt-sequences.txt",
    "4_IMGT-gapped-AA-sequences.txt", "6_Junction.txt"),
  cell = "Bcell",
  throughput = "High Throughput",
 preselection = "1,4C:W",
  selection = "5",
  identity_range = "85:100",
 vgenes = ""
 dgenes = ""
  jgenes = "",
  cdr3_length_range = "".
 aminoacid = ""
 pipeline = "1",
  select_clonotype = "V Gene + CDR3 Amino Acids",
  highly_sim_params = paste0("1-1 2-1 3-1 4-1 5-1 6-1 7-1 8-1 9-1 10-1 11-1 ",
    "12-1 13-1 14-1 15-2 16-2 17-2 18-2 19-2 20-2 21-2 23-2 24-2 25-2 ",
    "26-2 27-2 28-2 29-3 30-3 31-3 32-3 33-3 34-3 35-3 36-3 37-3 38-3 ",
    "39-3 40-3 41-3 42-3 43-3 44-3 45-3 46-3 47-3 48-3 49-3 50-3,1,Yes"),
  shared_clonotypes_params = "reads,1,Yes",
  highly_shared_clonotypes_params = "reads,1,Yes",
  repertoires_params = "1,4,6",
  identity_groups = "85:97,97:99,99:100,100:100",
 multiple_values_params = "2:7,2:3,2:5,2:11",
 alignment_params = "1,both,1,2:20",
 mutations_params = "both, 0.5, 0.5, 2:20"
)
```

#### **Arguments**

filelist

(character) The directory where the folders of the data is located. Note that every sample of the dataset must have **its own individual folder** and every sample folder must be in **one root folder**. Note that **every** file in the root folder will be used in the analysis.

Supposedly the dataset is in user's *Documents/* folder, one could use: fs::path\_home("Documents", "dataset"), with the help of path\_home function. See the package vignette for more.

output\_path

(character) The directory where the output data will be stored. Please provide a valid path, ideally the same way as datapath by using the path\_home function.

The default value points to Documents/twing output directory.

The default value points to *Documents/tripr\_output* directory. (character vector) The character vector of files of the IMGT output that will be

used through the analysis from each sample.

cell (character) 'Bcell' (default) or 'Tcell'.

throughput (character) 'High Throughput' (default) or 'Low Throughput'.

preselection (character) Preselection options: 1 == Only take into account Functional V-Gene, 2 == Only take into account CDR3 with no Special Characters (X, \*, #, .), 3 == Only take into account Productive Sequences, 4 == Only take into account CDR3 with valid start/end landmarks., For Preselection option 4, select start/end landmarks., Use the vertical line 'l' to add more than one start or end landmarks, Use comma',' to seperate the list of options, use semicolon':' to seperate start and end landmarks. selection (character) Selection options: 5 == V-REGION identity 6 == Select Specific V Gene, 7 == Select Specific J Gene, 8 == Select Specific D Gene, 9 == Select CDR3 length range, 10 == Only select CDR3 containing specific amino-acid sequence. Use comma ',' to seperate the list of options. identity\_range (character) V-REGION identity Use colon ':' to seperate identity low and high vgenes (character) Filter in specific V Genes, Separate the different V-Gene names with 'I' e.g. TRBV11-2|TRBV29-1\*03 (F) dgenes (character) Filter in specific D Genes, Separate the different D-Gene names with le.g. TRBD2|TRBD1 jgenes (character) Filter in specific J Genes, Separate the different J-Gene names with le.g. TRBJ2-6|TRBJ2-2 cdr3\_length\_range (character) Filter in rows with CDR3 lengths within a range, Use colon ':' to seperate identity low and high aminoacid (character) Filter in rows with CDR3 containing specific amino-acid sequence pipeline (character) Pipeline options: 1 == Clonotypes Computation, 2 == Highly Similar Clonotypes computation, 3 == Shared Clonotypes Computation, 4 == Highly Similar Shared Clonotypes Computation, 5 == Repertoires Extraction, 6 == Repertoires Comparison, 7 == Highly Similar Repertoires Extraction, 8 == Insert Identity groups, 9 == Somatic hypermutation status, 10 == CDR3 Distribution, 11 == Pi Distribution, 12 == Multiple value comparison, 13 == CDR3 with 1 length difference, 14 == Alignment, 15 == Somatic hypermutations, 16 == Logo,17 == SHM normal,18 == SHM High similarity,

19 == Diagnosis,

Use comma',' to seperate the list of options

#### select\_clonotype

(character) Compute clonotypes.

Select one the following options:

"V Gene + CDR3 Amino Acids",

"V Gene and Allele + CDR3 Amino Acids",

"V Gene + CDR3 Nucleotide",

"V Gene and Allele + CDR3 Nucleotide",

"J Gene + CDR3 Amino Acids".

"J Gene and Allele + CDR3 Amino Acids",

"J Gene + CDR3 Nucleotide",

"J Gene and Allele + CDR3 Nucleotide",

"CDR3 Amino Acids",

"CDR3 Nucleotide",

"Sequence

#### highly\_sim\_params

(character) Select number of missmatches, the threshold of the clonotype frequency and whether you want to take gene into account. Use dashes '-' to show the length of the CDR3 sequences and the number of allowed missmatches and spaces ' to separate. For the CDR3 lengths with not specified number of missmatches the default value is 1. Use comma ',' to separate the three options.

# shared\_clonotypes\_params

(character) Shared clonotypes computation.

Select 'reads' of 'threshold' for clonotypes, the number of reads or the threshold percentage accordingly, and whether you want to take gene into account. Use comma',' to seperate the 3 options

#### highly\_shared\_clonotypes\_params

(character) Highly Similar Shared Clonotypes Computation

Select 'reads' of 'threshold' for clonotypes, the number of reads or the threshold percentage accordingly, and whether you want to take gene into account. Use comma',' to seperate the 3 options

#### repertoires\_params

(character) Repertoires Extraction

Options:

1 == V Gene

2 == V Gene and allele

3 == J Gene

4 == J Gene and allele

5 == D Gene

6 == D Gene and allele

Use comma',' to seperate the selected options

#### identity\_groups

(character) Insert identity groups

Insert low and high values as follows:

low\_values:high\_values

Seperate low\_values and high\_values using comma ','.

#### multiple\_values\_params

(character) Multiple value comparison

Options:

1 == V GENE

2 == V GENE and allele

3 == J GENE

4 == J GENE and allele

5 == D GENE

6 == D GENE and allele

7 == CDR3-IMGT length

8 == D-REGION reading frame

9 == Molecular mass

10 == pI

11 == V-REGION identity Use colon ':' to indicate combinations of 2 values, use comma "," to seperate the selected options

#### alignment\_params

(character) Alignment parameters:

Region for Alignment: 1 == V.D.J.REGION or 2 == V.J.REGION

AA or Nt: Select 'aa' or 'nt' or 'both'

Germline: 1 == Use Allele's germline or 2 == Use Gene's germline

Use: 1 == All clonotypes or 2 == Select top N clonotypes or 3 == Select threshold for clonotypes

Use comma',' to seperate the 4 parameters. If you select option 2 or 3 at the 4th parameter you have to set the N or the threshold as well using colon':'.

#### mutations\_params

(character) Somatic hypermutations parameters:

AA or Nt: Select 'aa' or 'nt' or 'both'

Set threshold for AA

Set threshold for Nt

Use: 1 == All clonotypes or 2 == Select top N clonotypes or 3 == Select threshold for clonotypes

Use comma',' to seperate the 3 parameters. If you select option 2 or 3 at the 3rd parameter you have to set the N or the threshold as well using colon':'.

# Value

None

# **Examples**

```
## Do not run

run_TRIP(
   output_path=tools::R_user_dir("tripr", which="cache"),
   filelist=c("1_Summary.txt", "2_IMGT-gapped-nt-sequences.txt",
        "4_IMGT-gapped-AA-sequences.txt", "6_Junction.txt"),
   cell="Bcell",
   throughput="High Throughput",
   preselection="1,2,3,4C:W",
```

8 tripr

```
selection="5",
identity_range="88:100",
cdr3_length_range="",
pipeline="1",
select_clonotype="V Gene + CDR3 Amino Acids")
```

tripr

tripr

# Description

T-cell Receptor/Immunoglobulin Profiler (TRIP)

# **Details**

The only function you're likely to need from tripr is [run\_app()]. Otherwise refer to the vignettes for using tripr.

# **Index**

```
enableBookmarking(), 3
path_home, 4
run_app, 2
run_TRIP, 3
tripr, 8
```