

# Package ‘variancePartition’

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**Title** Quantify and interpret drivers of variation in multilevel gene expression experiments

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**Description** Quantify and interpret multiple sources of biological and technical variation in gene expression experiments. Uses a linear mixed model to quantify variation in gene expression attributable to individual, tissue, time point, or technical variables. Includes dream differential expression analysis for repeated measures.

**VignetteBuilder** knitr

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**Encoding** UTF-8

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<https://DiseaseNeuroGenomics.github.io/variancePartition>

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---

.getAllUniContrasts     *Get all univariate contrasts*

---

**Description**

Get all univariate contrasts

**Usage**

.getAllUniContrasts(formula, data)

**Arguments**

formula	specifies variables for the linear (mixed) model. Must only specify covariates, since the rows of exprObj are automatically used as a response. e.g.: ~ a + b + (1 c) Formulas with only fixed effects also work
data	data.frame with columns corresponding to formula

**Value**

Matrix testing each variable one at a time. Contrasts are on rows

---

`.isMixedModelFormula` *Check if model contains a random effect*

---

### Description

Check if model contains a random effect

### Usage

```
.isMixedModelFormula(formula)
```

### Arguments

formula            model formula

---

`.standard_transform` *Compute standard post-processing values*

---

### Description

These values are typically computed by eBayes

### Usage

```
.standard_transform(fit, sigma = fit$sigma)
```

### Arguments

fit                result of dream (MArrayLM2)  
sigma             vector of standard errors used to compute t-statistic. Can be maximum likelihood estimates, or posterior means

### Value

MArrayLM2 object with values computed

---

applyQualityWeights    *Apply pre-specified sample weights*

---

**Description**

Apply pre-specified sample weights by scaling existing precision weights

**Usage**

```
applyQualityWeights(vobj, weights)
```

**Arguments**

vobj	EList from voom or voomWithDreamWeights.
weights	sample level weights

**Details**

Apply pre-specified sample-level weights to the existing precision weights estimated from the data. While the `limma::voomWithQualityWeights` function of Lui et al. (2015) estimates the sample-level weights from voom fit, here the weights are fixed beforehand.

**References**

Liu R, Holik AZ, Su S, Jansz N, Chen K, Leong HS, Blewitt ME, Asselin-Labat M, Smyth GK, Ritchie ME (2015). "Why weight? Modelling sample and observational level variability improves power in RNA-seq analyses." *Nucleic acids research*, **43**(15), e97–e97.

**See Also**

`limma::voomWithQualityWeights`

---

`as.data.frame.varPartResults`  
*Convert to data.frame*

---

**Description**

Convert varPartResults to data.frame

**Usage**

```
## S3 method for class 'varPartResults'
as.data.frame(x, row.names = NULL, optional = FALSE, ...)
```

**Arguments**

x	varPartResults
row.names	pass thru to generic
optional	pass thru to generic
...	other arguments.

**Value**

data.frame

**Examples**

```
# load library
# library(variancePartition)

# load simulated data:
# geneExpr: matrix of gene expression values
# info: information/metadata about each sample
data(varPartData)

# Specify variables to consider
# Age is continuous so we model it as a fixed effect
# Individual and Tissue are both categorical, so we model them as random effects
form <- ~ Age + (1 | Individual) + (1 | Tissue)

# Fit model
varPart <- fitExtractVarPartModel(geneExpr[1:5, ], form, info)

# convert to matrix
as.data.frame(varPart)
```

---

as.matrix, varPartResults-method

*Convert to matrix*

---

**Description**

Convert varPartResults to matrix

**Usage**

```
## S4 method for signature 'varPartResults'
as.matrix(x, ...)
```

**Arguments**

x	varPartResults
...	other arguments.

**Value**

matrix

**Examples**

```

# load library
# library(variancePartition)

# load simulated data:
# geneExpr: matrix of gene expression values
# info: information/metadata about each sample
data(varPartData)

# Specify variables to consider
# Age is continuous so we model it as a fixed effect
# Individual and Tissue are both categorical, so we model them as random effects
form <- ~ Age + (1 | Individual) + (1 | Tissue)

# Fit model
varPart <- fitExtractVarPartModel(geneExpr[1:5, ], form, info)

# convert to matrix
as.matrix(varPart)

```

---

augmentPriorCount	<i>Augment observed read counts with prior count</i>
-------------------	--

---

**Description**

Augment observed read counts with prior count since log of zero counts is undefined. The prior count added to each sample is scaled so that no variance is introduced

**Usage**

```

augmentPriorCount(
  counts,
  lib.size = colSums2(counts),
  prior.count = 0.5,
  scaledByLib = FALSE
)

```

**Arguments**

counts	matrix of read counts with genes as rows and samples as columns
lib.size	library sizes, the sum of all reads for each sample
prior.count	average prior count added to each sample.
scaledByLib	if TRUE, scale pseudocount by lib.size. Else to standard constant pseudocount addition

**Details**

Adding prior counts removes the issue of evaluating the log of zero counts, and stabilizes the log transform when counts are very small. However, adding a constant prior count to all samples can introduce an artifact. Consider two samples each with zero counts for a given gene, but one as a

library size of 1k and the other of 50k. After applying the prior count values become  $pc / 1k$  and  $pc / 50k$ . It appears that there is variance in the expression of this gene, even though no counts are observed. This is driven only by variation in the library size, which does not reflect biology. This issue is most problematic for small counts.

Instead, we make the reasonable assumption that a gene does not have expression variance unless supported sufficiently by counts in the numerator. Consider adding a different prior count to each sample so that genes with zero counts end up with zero variance. This corresponds to adding  $prior.count * lib.size[i] / mean(lib.size)$  to sample  $i$ .

This is done in the backend of `edgeR::cpm()`, but this function allows users to apply it more generally.

### Value

matrix with augmented counts

### See Also

`edgeR::cpm()`

### Examples

```
library(edgeR)

data(varPartDEdata)

# normalize RNA-seq counts
dge <- DGEList(counts = countMatrix)
dge <- calcNormFactors(dge)

countsAugmented <- augmentPriorCount(dge$counts, dge$samples$lib.size, 1)
```

---

BIC.MArrayLM

*BIC from model fit*

---

### Description

BIC from model fit

### Usage

```
## S3 method for class 'MArrayLM'
BIC(object, vobj, ...)
```

### Arguments

<code>object</code>	result of <code>lmFit()</code> or <code>dream()</code>
<code>vobj</code>	<code>EList</code> used to fit model
<code>...</code>	See <code>?stats::BIC</code>



---

BIC.MArrayLM2	<i>BIC from model fit</i>
---------------	---------------------------

---

**Description**

BIC from model fit using edf

**Usage**

```
## S3 method for class 'MArrayLM2'
BIC(object, vobj, ...)
```

**Arguments**

object	result of dream()
vobj	EList used to fit model
...	See ?stats::BIC

---

calcVarPart	<i>Compute variance statistics</i>
-------------	------------------------------------

---

**Description**

Compute fraction of variation attributable to each variable in regression model. Also interpretable as the intra-class correlation after correcting for all other variables in the model.

**Usage**

```
calcVarPart(fit, returnFractions = TRUE, ...)

## S4 method for signature 'lm'
calcVarPart(fit, returnFractions = TRUE, ...)

## S4 method for signature 'lmerMod'
calcVarPart(fit, returnFractions = TRUE, ...)

## S4 method for signature 'glm'
calcVarPart(fit, returnFractions = TRUE, ...)

## S4 method for signature 'negbin'
calcVarPart(fit, returnFractions = TRUE, ...)

## S4 method for signature 'glmerMod'
calcVarPart(fit, returnFractions = TRUE, ...)
```

**Arguments**

fit                    model fit from lm() or lmer()  
 returnFractions  
                       default: TRUE. If TRUE return fractions that sum to 1. Else return unscaled  
                       variance components.  
 ...                    additional arguments (not currently used)

**Details**

For linear model, variance fractions are computed based on the sum of squares explained by each component. For the linear mixed model, the variance fractions are computed by variance component estimates for random effects and sum of squares for fixed effects.

For a generalized linear model, the variance fraction also includes the contribution of the link function so that fractions are reported on the linear (i.e. link) scale rather than the observed (i.e. response) scale. For linear regression with an identity link, fractions are the same on both scales. But for logit or probit links, the fractions are not well defined on the observed scale due to the transformation imposed by the link function.

The variance implied by the link function is the variance of the corresponding distribution:

logit -> logistic distribution -> variance is  $\pi^2/3$

probit -> standard normal distribution -> variance is 1

For the Poisson distribution with rate  $\lambda$ , the variance is  $\log(1 + 1/\lambda)$ .

For the negative binomial distribution with rate  $\lambda$  and shape  $\theta$ , the variance is  $\log(1 + 1/\lambda + 1/\theta)$ .

Variance decomposition is reviewed by Nakagawa and Schielzeth (2012), and expanded to other GLMs by Nakagawa, Johnson and Schielzeth (2017). See McKelvey and Zavoina (1975) for early work on applying to GLMs. Also see DeMaris (2002)

We note that Nagelkerke's pseudo  $R^2$  evaluates the variance explained by the full model. Instead, a variance partitioning approach evaluates the variance explained by each term in the model, so that the sum of each systematic plus random term sums to 1 (Hoffman and Schadt, 2016; Nakagawa and Schielzeth, 2012).

**Value**

fraction of variance explained / ICC for each variable in the regression model

**References**

- Nakagawa S, Johnson PC, Schielzeth H (2017). "The coefficient of determination  $R^2$  and intra-class correlation coefficient from generalized linear mixed-effects models revisited and expanded." *Journal of the Royal Society Interface*, **14**(134), 20170213.
- Nakagawa S, Schielzeth H (2013). "A general and simple method for obtaining  $R^2$  from generalized linear mixed-effects models." *Methods in ecology and evolution*, **4**(2), 133–142.
- McKelvey RD, Zavoina W (1975). "A statistical model for the analysis of ordinal level dependent variables." *Journal of mathematical sociology*, **4**(1), 103–120.
- DeMaris A (2002). "Explained variance in logistic regression: A Monte Carlo study of proposed measures." *Sociological Methods & Research*, **31**(1), 27–74.
- Hoffman GE, Schadt EE (2016). "variancePartition: interpreting drivers of variation in complex gene expression studies." *BMC bioinformatics*, **17**(1), 1–13.

**Examples**

```

library(lme4)
data(varPartData)

# Linear mixed model
fit <- lmer(geneExpr[1, ] ~ (1 | Tissue) + Age, info)
calcVarPart(fit)

# Linear model
# Note that the two models produce slightly different results
# This is expected: they are different statistical estimates
# of the same underlying value
fit <- lm(geneExpr[1, ] ~ Tissue + Age, info)
calcVarPart(fit)

```

---

canCorPairs

*canCorPairs*


---

**Description**

Assess correlation between all pairs of variables in a formula

**Usage**

```
canCorPairs(formula, data, showWarnings = TRUE)
```

**Arguments**

formula	standard additive linear model formula (doesn't support random effects currently, so just change the syntax)
data	data.frame with the data for the variables in the formula
showWarnings	default to true

**Details**

Canonical Correlation Analysis (CCA) is similar to correlation between two vectors, except that CCA can accommodate matrices as well. For a pair of variables, `canCorPairs` assesses the degree to which they co-vary and contain the same information. Variables in the formula can be a continuous variable or a discrete variable expanded to a matrix (which is done in the backend of a regression model). For a pair of variables, `canCorPairs` uses CCA to compute the correlation between these variables and returns the pairwise correlation matrix.

Statistically, let  $\rho$  be the array of correlation values returned by the standard R function `cancor` to compute CCA. `canCorPairs()` returns  $\sqrt{\text{mean}(\rho^2)}$ , which is the fraction of the maximum possible correlation. When comparing a two vectors, or a vector and a matrix, this gives the same value as the absolute correlation. When comparing two sets of categorical variables (i.e. expanded to two matrices), this is equivalent to Cramer's V statistic.

Note that CCA returns correlation values between 0 and 1.

**Value**

Matrix of correlation values between all pairs of variables.

**Examples**

```
# load library
# library(variancePartition)

# load simulated data:
data(varPartData)

# specify formula
form <- ~ Individual + Tissue + Batch + Age + Height

# Compute Canonical Correlation Analysis (CCA)
# between all pairs of variables
# returns absolute correlation value
C <- canCorPairs(form, info)

# Plot correlation matrix
plotCorrMatrix(C)
```

---

`classifyTestsF`*Multiple Testing Genewise Across Contrasts*

---

**Description**

For each gene, classify a series of related t-statistics as up, down or not significant.

**Usage**

```
classifyTestsF(object, ...)
```

**Arguments**

<code>object</code>	numeric matrix of t-statistics or an 'MArrayLM2' object from which the t-statistics may be extracted.
<code>...</code>	additional arguments

**Details**

Works like `limma::classifyTestsF`, except `object` can have a list of covariance matrices `object$cov.coefficients.list`, instead of just one in `object$cov.coefficients`

**See Also**

```
limma::classifyTestsF
```

---

 classifyTestsF,MArrayLM2-method

*Multiple Testing Genewise Across Contrasts*


---

### Description

For each gene, classify a series of related t-statistics as up, down or not significant.

### Usage

```
## S4 method for signature 'MArrayLM2'
classifyTestsF(
  object,
  cor.matrix = NULL,
  df = Inf,
  p.value = 0.01,
  fstat.only = FALSE
)
```

### Arguments

object	numeric matrix of t-statistics or an 'MArrayLM2' object from which the t-statistics may be extracted.
cor.matrix	covariance matrix of each row of t-statistics. Defaults to the identity matrix.
df	numeric vector giving the degrees of freedom for the t-statistics. May have length 1 or length equal to the number of rows of tstat.
p.value	numeric value between 0 and 1 giving the desired size of the test
fstat.only	logical, if 'TRUE' then return the overall F-statistic as for 'FStat' instead of classifying the test results

### Details

Works like `limma::classifyTestsF`, except `object` can have a list of covariance matrices `object$cov.coefficients.list`, instead of just one in `object$cov.coefficients`

### See Also

`limma::classifyTestsF`

---

 colinearityScore

*Collinearity score*


---

### Description

Collinearity score for a regression model indicating if variables are too highly correlated to give meaningful results

**Usage**

```
colinearityScore(fit)
```

**Arguments**

```
fit          regression model fit from lm() or lmer()
```

**Value**

Returns the collinearity score between 0 and 1, where a score > 0.999 means the degree of collinearity is too high. This function reports the correlation matrix between coefficient estimates for fixed effects. The collinearity score is the maximum absolute correlation value of this matrix. Note that the values are the correlation between the parameter estimates, and not between the variables themselves.

**Examples**

```
# load library
# library(variancePartition)

# load simulated data:
data(varPartData)
#
form <- ~ Age + (1 | Individual) + (1 | Tissue)

res <- fitVarPartModel(geneExpr[1:10, ], form, info)

# evaluate the collinearity score on the first model fit
# this reports the correlation matrix between coefficients estimates
# for fixed effects
# the collinearity score is the maximum absolute correlation value
# If the collinearity score > .999 then the variance partition
# estimates may be problematic
# In that case, a least one variable should be omitted
colinearityScore(res[[1]])
```

---

deviation

*Deviation from expectation for each observation*


---

**Description**

Given a model fit for each features, residuals are computed and transformed based on an absolute value or squaring transform.

**Usage**

```
deviation(fit, method = c("AD", "SQ"), scale = c("leverage", "none"))

## S4 method for signature 'MArrayLM'
deviation(fit, method = c("AD", "SQ"), scale = c("leverage", "none"))
```

**Arguments**

fit                    model fit from dream()  
 method                transform the residuals using absolute deviation ("AD") or squared deviation ("SQ").  
 scale                  scale each observation by "leverage", or no scaling ("none")

**Value**

matrix of deviations from expectation for each observation

**See Also**

diffVar()

**Examples**

```
# library(variancePartition)
library(edgeR)
data(varPartDEdata)

# filter genes by number of counts
isexpr <- rowSums(cpm(countMatrix) > 0.1) >= 5

# Standard usage of limma/voom
dge <- DGEList(countMatrix[isexpr, ])
dge <- calcNormFactors(dge)

# make this vignette faster by analyzing a subset of genes
dge <- dge[1:1000, ]

# regression formula
form <- ~Disease

# estimate precision weights
vobj <- voomWithDreamWeights(dge, form, metadata)

# fit dream model
fit <- dream(vobj, form, metadata)
fit <- eBayes(fit)

# Compute deviation from expectation for each observation
# using model residuals
z <- deviation(fit)
z[1:4, 1:4]
```

---

diffVar

*Test differential variance*

---

**Description**

Test the association between a covariate of interest and the response's deviation from expectation.

**Usage**

```
diffVar(
  fit,
  method = c("AD", "SQ"),
  scale = c("leverage", "none"),
  BPPARAM = SerialParam(),
  ...
)

## S4 method for signature 'MArrayLM'
diffVar(
  fit,
  method = c("AD", "SQ"),
  scale = c("leverage", "none"),
  BPPARAM = SerialParam(),
  ...
)
```

**Arguments**

fit	model fit from <code>dream()</code>
method	transform the residuals using absolute deviation ("AD") or squared deviation ("SQ").
scale	scale each observation by "leverage", or no scaling ("none")
BPPARAM	parameters for parallel evaluation
...	other parameters passed to <code>dream()</code>

**Details**

This method performs a test of differential variance between two subsets of the data, in a way that generalizes to multiple categories, continuous variables and metrics of spread beyond variance. For the two category test, this method is similar to Levene's test. This model was adapted from Phipson, et al (2014), extended to linear mixed models, and adapted to be compatible with `dream()`.

This method is composed of multiple steps where 1) a typical linear (mixed) model is fit with `dream()`, 2) residuals are computed and transformed based on an absolute value or squaring transform, 3) a second regression is performed with `dream()` to test if a variable is associated with increased deviation from expectation. Both regression take advantage of the `dream()` linear (mixed) modelling framework followed by empirical Bayes shrinkage that extends the `limma::voom()` framework.

Note that `diffVar()` takes the results of the first regression as a parameter to use as a starting point.

**Value**

MArrayLM object storing differential results to be passed to `topTable()`

**References**

Phipson B, Oshlack A (2014). "DiffVar: a new method for detecting differential variability with application to methylation in cancer and aging." *Genome biology*, **15**(9), 1–16.



**See Also**

```
missMethyl::diffVar(), car::leveneTest()
```

**Examples**

```
# library(variancePartition)
library(edgeR)
data(varPartDEdata)

# filter genes by number of counts
isexpr <- rowSums(cpm(countMatrix) > 0.1) >= 5

# Standard usage of limma/voom
dge <- DGEList(countMatrix[isexpr, ])
dge <- calcNormFactors(dge)

# make this vignette faster by analyzing a subset of genes
dge <- dge[1:1000, ]

# regression formula
form <- ~Disease

# estimate precision weights
vobj <- voomWithDreamWeights(dge, form, metadata)

# fit dream model
fit <- dream(vobj, form, metadata)
fit <- eBayes(fit)

# fit differential variance model
res <- diffVar(fit)

# extract results for differential variance based on Disease
topTable(res, coef = "Disease1", number = 3)

# Box plot of top hit
# Since ASCL3 has a negative logFC,
# the deviation from expectation is *smaller* in
# Disease==1 compared to baseline.
gene <- "ENST00000325884.1 gene=ASCL3"
boxplot(vobj$E[gene, ] ~ metadata$Disease, main = gene)
```

---

dream

*Differential expression with linear mixed model*

---

**Description**

Fit linear mixed model for differential expression and perform hypothesis test on fixed effects as specified in the contrast matrix L

**Usage**

```

dream(
  exprObj,
  formula,
  data,
  L,
  ddf = c("adaptive", "Satterthwaite", "Kenward-Roger"),
  useWeights = TRUE,
  control = vpcontrol,
  hideErrorsInBackend = FALSE,
  REML = TRUE,
  BPPARAM = SerialParam(),
  ...
)

```

**Arguments**

exprObj	matrix of expression data (g genes x n samples), or ExpressionSet, or EList returned by voom() from the limma package
formula	specifies variables for the linear (mixed) model. Must only specify covariates, since the rows of exprObj are automatically used as a response. e.g.: ~ a + b + (1 c) Formulas with only fixed effects also work, and lmFit() followed by contrasts.fit() are run.
data	data.frame with columns corresponding to formula
L	contrast matrix specifying a linear combination of fixed effects to test
ddf	Specify "Satterthwaite" or "Kenward-Roger" method to estimate effective degrees of freedom for hypothesis testing in the linear mixed model. Note that Kenward-Roger is more accurate, but is <i>*much*</i> slower. Satterthwaite is a good enough approximation for most datasets. "adaptive" (Default) uses KR for <= 20 samples.
useWeights	if TRUE, analysis uses heteroskedastic error estimates from voom(). Value is ignored unless exprObj is an EList() from voom() or weightsMatrix is specified
control	control settings for lmer()
hideErrorsInBackend	default FALSE. If TRUE, hide errors in attr(., "errors") and attr(., "error.initial")
REML	use restricted maximum likelihood to fit linear mixed model. default is TRUE. See Details.
BPPARAM	parameters for parallel evaluation
...	Additional arguments for lmer() or lm()

**Details**

A linear (mixed) model is fit for each gene in exprObj, using formula to specify variables in the regression (Hoffman and Roussos, 2021). If categorical variables are modeled as random effects (as is recommended), then a linear mixed model is used. For example if formula is ~ a + b + (1|c), then the model is

```
fit <- lmer( exprObj[j,] ~ a + b + (1|c), data=data)
```

useWeights=TRUE causes weightsMatrix[j,] to be included as weights in the regression model.

Note: Fitting the model for 20,000 genes can be computationally intensive. To accelerate computation, models can be fit in parallel using `BiocParallel` to run code in parallel. Parallel processing must be enabled before calling this function. See below.

The regression model is fit for each gene separately. Samples with missing values in either gene expression or metadata are omitted by the underlying call to `lmer`.

Hypothesis tests and degrees of freedom are produced by `lmerTest` and `pbkrtest` packages

While `REML=TRUE` is required by `lmerTest` when `ddf='Kenward-Roger'`, `ddf='Satterthwaite'` can be used with `REML` as `TRUE` or `FALSE`. Since the Kenward-Roger method gave the best power with an accurate control of false positive rate in our simulations, and since the Satterthwaite method with `REML=TRUE` gives p-values that are slightly closer to the Kenward-Roger p-values, `REML=TRUE` is the default. See Vignette "3) Theory and practice of random effects and REML"

## Value

`MArrayLM2` object (just like `MArrayLM` from `limma`), and the directly estimated p-value (without `eBayes`)

## References

Hoffman GE, Roussos P (2021). "dream: Powerful differential expression analysis for repeated measures designs." *Bioinformatics*, **37**(2), 192–201.

## Examples

```
# library(variancePartition)

# load simulated data:
# geneExpr: matrix of *normalized* gene expression values
# info: information/metadata about each sample
data(varPartData)

form <- ~ Batch + (1 | Individual) + (1 | Tissue)

# Fit linear mixed model for each gene
# run on just 10 genes for time
# NOTE: dream() runs on *normalized* data
fit <- dream(geneExpr[1:10, ], form, info)
fit <- eBayes(fit)

# view top genes
topTable(fit, coef = "Batch2", number = 3)

# get contrast matrix testing if the coefficient for Batch3 is
# different from coefficient for Batch2
# Name this comparison as 'compare_3_2'
# The variable of interest must be a fixed effect
L <- makeContrastsDream(form, info, contrasts = c(compare_3_2 = "Batch3 - Batch2"))

# plot contrasts
plotContrasts(L)

# Fit linear mixed model for each gene
# run on just 10 genes for time
fit2 <- dream(geneExpr[1:10, ], form, info, L)
fit2 <- eBayes(fit2)
```

```

# view top genes for this contrast
topTable(fit2, coef = "compare_3_2", number = 3)

# Parallel processing using multiple cores with reduced memory usage
param <- SnowParam(4, "SOCK", progressbar = TRUE)
fit3 <- dream(geneExpr[1:10, ], form, info, L, BPPARAM = param)
fit3 <- eBayes(fit3)

# Fit fixed effect model for each gene
# Use lmFit in the backend
form <- ~Batch
fit4 <- dream(geneExpr[1:10, ], form, info, L)
fit4 <- eBayes(fit4)

# view top genes
topTable(fit4, coef = "compare_3_2", number = 3)

# Compute residuals using dream
residuals(fit4)[1:4, 1:4]

```

---

dscchisq

*Scaled chi-square*


---

### Description

Scaled chi-square density using a gamma distribution

### Usage

```
dscchisq(x, a, b)
```

### Arguments

x	vector of quantiles.
a	scale
b	degrees of freedom

---

eBayes,MArrayLM2-method

*eBayes for MArrayLM2*


---

### Description

eBayes for result of linear mixed model for with dream() using residual degrees of freedom approximated with rdf.merMod()

**Usage**

```
## S4 method for signature 'MArrayLM2'
eBayes(
  fit,
  proportion = 0.01,
  stdev.coef.lim = c(0.1, 4),
  trend = FALSE,
  robust = FALSE,
  winsor.tail.p = c(0.05, 0.1),
  legacy = NULL
)
```

**Arguments**

fit	fit
proportion	proportion
stdev.coef.lim	stdev.coef.lim
trend	trend
robust	robust
winsor.tail.p	winsor.tail.p
legacy	legacy

**Value**

results of eBayes using approximated residual degrees of freedom

**See Also**

dream(), rdf.merMod(), limma::eBayes()

---

ESS

*Effective sample size*

---

**Description**

Compute effective sample size based on correlation structure in linear mixed model

**Usage**

```
ESS(fit, method = "full")
```

```
## S4 method for signature 'lmerMod'
ESS(fit, method = "full")
```

**Arguments**

fit	model fit from lmer()
method	"full" uses the full correlation structure of the model. The "approximate" method makes the simplifying assumption that the study has a mean of m samples in each of k groups, and computes m based on the study design. When the study design is evenly balanced (i.e. the assumption is met), this gives the same results as the "full" method.

**Details**

Effective sample size calculations are based on:

Liu, G., and Liang, K. Y. (1997). Sample size calculations for studies with correlated observations. *Biometrics*, 53(3), 937-47.

"full" method: if

$$V_x = \text{var}(Y; x)$$

is the variance-covariance matrix of Y, the response, based on the covariate x, then the effective sample size corresponding to this covariate is

$$\sum_{i,j} (V_x^{-1})_{i,j}$$

. In R notation, this is: `sum(solve(V_x))`. In practice, this can be evaluated as `sum(w)`, where R

"approximate" method: Letting m be the mean number of samples per group,

$$k$$

be the number of groups, and

$$\rho$$

be the intraclass correlation, the effective sample size is

$$mk / (1 + \rho(m - 1))$$

Note that these values are equal when there are exactly m samples in each group. If m is only an average then this is an approximation.

**Value**

effective sample size for each random effect in the model

**Examples**

```
library(lme4)
data(varPartData)

# Linear mixed model
fit <- lmer(geneExpr[1, ] ~ (1 | Individual) + (1 | Tissue) + Age, info)

# Effective sample size
ESS(fit)
```

---

extractVarPart

*Extract variance statistics*

---

**Description**

Extract variance statistics from list of models fit with `lm()` or `lmer()`

**Usage**

```
extractVarPart(modelList, ...)
```

**Arguments**

```

modellist      list of lmer() model fits
...           other arguments

```

**Value**

data.frame of fraction of variance explained by each variable, after correcting for all others.

**Examples**

```

# library(variancePartition)

library(BiocParallel)

# load simulated data:
# geneExpr: matrix of gene expression values
# info: information/metadata about each sample
data(varPartData)

# Specify variables to consider
# Age is continuous so we model it as a fixed effect
# Individual and Tissue are both categorical, so we model them as random effects
form <- ~ Age + (1 | Individual) + (1 | Tissue)

# Step 1: fit linear mixed model on gene expression
# If categorical variables are specified, a linear mixed model is used
# If all variables are modeled as continuous, a linear model is used
# each entry in results is a regression model fit on a single gene
# Step 2: extract variance fractions from each model fit
# for each gene, returns fraction of variation attributable to each variable
# Interpretation: the variance explained by each variable
# after correction for all other variables
varPart <- fitExtractVarPartModel(geneExpr, form, info)

# violin plot of contribution of each variable to total variance
plotVarPart(sortCols(varPart))

# Advanced:
# Fit model and extract variance in two separate steps
# Step 1: fit model for each gene, store model fit for each gene in a list
results <- fitVarPartModel(geneExpr, form, info)

# Step 2: extract variance fractions
varPart <- extractVarPart(results)

```

---

```
fitExtractVarPartModel
```

*Fit linear (mixed) model, report variance fractions*

---

**Description**

Fit linear (mixed) model to estimate contribution of multiple sources of variation while simultaneously correcting for all other variables. Report fraction of variance attributable to each variable

**Usage**

```
fitExtractVarPartModel(  
  exprObj,  
  formula,  
  data,  
  REML = FALSE,  
  useWeights = TRUE,  
  control = vpcontrol,  
  hideErrorsInBackend = FALSE,  
  showWarnings = TRUE,  
  BPPARAM = SerialParam(),  
  ...  
)  
  
## S4 method for signature 'matrix'  
fitExtractVarPartModel(  
  exprObj,  
  formula,  
  data,  
  REML = FALSE,  
  useWeights = TRUE,  
  control = vpcontrol,  
  hideErrorsInBackend = FALSE,  
  showWarnings = TRUE,  
  BPPARAM = SerialParam(),  
  ...  
)  
  
## S4 method for signature 'data.frame'  
fitExtractVarPartModel(  
  exprObj,  
  formula,  
  data,  
  REML = FALSE,  
  useWeights = TRUE,  
  control = vpcontrol,  
  hideErrorsInBackend = FALSE,  
  showWarnings = TRUE,  
  BPPARAM = SerialParam(),  
  ...  
)  
  
## S4 method for signature 'EList'  
fitExtractVarPartModel(  
  exprObj,  
  formula,  
  data,  
  REML = FALSE,  
  useWeights = TRUE,  
  control = vpcontrol,  
  hideErrorsInBackend = FALSE,  
  showWarnings = TRUE,
```



```

    BPPARAM = SerialParam(),
    ...
)

## S4 method for signature 'ExpressionSet'
fitExtractVarPartModel(
  exprObj,
  formula,
  data,
  REML = FALSE,
  useWeights = TRUE,
  control = vpcontrol,
  hideErrorsInBackend = FALSE,
  showWarnings = TRUE,
  BPPARAM = SerialParam(),
  ...
)

## S4 method for signature 'sparseMatrix'
fitExtractVarPartModel(
  exprObj,
  formula,
  data,
  REML = FALSE,
  useWeights = TRUE,
  control = vpcontrol,
  hideErrorsInBackend = FALSE,
  showWarnings = TRUE,
  BPPARAM = SerialParam(),
  ...
)

```

### Arguments

exprObj	matrix of expression data (g genes x n samples), or ExpressionSet, or EList returned by voom() from the limma package
formula	specifies variables for the linear (mixed) model. Must only specify covariates, since the rows of exprObj are automatically used as a response. e.g.: ~ a + b + (1 c)
data	data.frame with columns corresponding to formula
REML	use restricted maximum likelihood to fit linear mixed model. default is FALSE. See Details.
useWeights	if TRUE, analysis uses heteroskedastic error estimates from voom(). Value is ignored unless exprObj is an EList() from voom() or weightsMatrix is specified
control	control settings for lmer()
hideErrorsInBackend	default FALSE. If TRUE, hide errors in attr(., "errors") and attr(., "error.initial")
showWarnings	default TRUE. Indicate model failures
BPPARAM	parameters for parallel evaluation
...	Additional arguments for lmer() or lm()

## Details

A linear (mixed) model is fit for each gene in `exprObj`, using formula to specify variables in the regression. If categorical variables are modeled as random effects (as is recommended), then a linear mixed model is used. For example if formula is  $\sim a + b + (1|c)$ , then the model is

```
fit <- lmer( exprObj[j,] ~ a + b + (1|c), data=data)
```

If there are no random effects, so formula is  $\sim a + b + c$ , a 'standard' linear model is used:

```
fit <- lm( exprObj[j,] ~ a + b + c, data=data)
```

In both cases, `useWeights=TRUE` causes `weightsMatrix[j,]` to be included as weights in the regression model.

Note: Fitting the model for 20,000 genes can be computationally intensive. To accelerate computation, models can be fit in parallel using `BiocParallel` to run in parallel. Parallel processing must be enabled before calling this function. See below.

The regression model is fit for each gene separately. Samples with missing values in either gene expression or metadata are omitted by the underlying call to `lm/lmer`.

`REML=FALSE` uses maximum likelihood to estimate variance fractions. This approach produced unbiased estimates, while `REML=TRUE` can show substantial bias. See Vignette "3) Theory and practice of random effects and REML"

## Value

`list()` of where each entry is a model fit produced by `lmer()` or `lm()`

## Examples

```
# load library
# library(variancePartition)

library(BiocParallel)

# load simulated data:
# geneExpr: matrix of gene expression values
# info: information/metadata about each sample
data(varPartData)

# Specify variables to consider
# Age is continuous so we model it as a fixed effect
# Individual and Tissue are both categorical, so we model them as random effects
form <- ~ Age + (1 | Individual) + (1 | Tissue)

# Step 1: fit linear mixed model on gene expression
# If categorical variables are specified, a linear mixed model is used
# If all variables are modeled as continuous, a linear model is used
# each entry in results is a regression model fit on a single gene
# Step 2: extract variance fractions from each model fit
# for each gene, returns fraction of variation attributable to each variable
# Interpretation: the variance explained by each variable
# after correction for all other variables
varPart <- fitExtractVarPartModel(geneExpr, form, info)

# violin plot of contribution of each variable to total variance
plotVarPart(sortCols(varPart))

# Note: fitExtractVarPartModel also accepts ExpressionSet
```

```

data(sample.ExpressionSet, package = "Biobase")

# ExpressionSet example
form <- ~ (1 | sex) + (1 | type) + score
info2 <- Biobase::pData(sample.ExpressionSet)
varPart2 <- fitExtractVarPartModel(sample.ExpressionSet, form, info2)

```

---

fitVarPartModel	<i>Fit linear (mixed) model</i>
-----------------	---------------------------------

---

## Description

Fit linear (mixed) model to estimate contribution of multiple sources of variation while simultaneously correcting for all other variables.

## Usage

```

fitVarPartModel(
  exprObj,
  formula,
  data,
  REML = FALSE,
  useWeights = TRUE,
  fxn = identity,
  control = vpcontrol,
  hideErrorsInBackend = FALSE,
  showWarnings = TRUE,
  BPPARAM = SerialParam(),
  ...
)

## S4 method for signature 'matrix'
fitVarPartModel(
  exprObj,
  formula,
  data,
  REML = FALSE,
  useWeights = TRUE,
  fxn = identity,
  control = vpcontrol,
  hideErrorsInBackend = FALSE,
  showWarnings = TRUE,
  BPPARAM = SerialParam(),
  ...
)

## S4 method for signature 'data.frame'
fitVarPartModel(
  exprObj,
  formula,

```

```
data,
REML = FALSE,
useWeights = TRUE,
fxn = identity,
control = vpcontrol,
hideErrorsInBackend = FALSE,
showWarnings = TRUE,
BPPARAM = SerialParam(),
...
)

## S4 method for signature 'EList'
fitVarPartModel(
  exprObj,
  formula,
  data,
  REML = FALSE,
  useWeights = TRUE,
  fxn = identity,
  control = vpcontrol,
  hideErrorsInBackend = FALSE,
  showWarnings = TRUE,
  BPPARAM = SerialParam(),
  ...
)

## S4 method for signature 'ExpressionSet'
fitVarPartModel(
  exprObj,
  formula,
  data,
  REML = FALSE,
  useWeights = TRUE,
  fxn = identity,
  control = vpcontrol,
  hideErrorsInBackend = FALSE,
  showWarnings = TRUE,
  BPPARAM = SerialParam(),
  ...
)

## S4 method for signature 'sparseMatrix'
fitVarPartModel(
  exprObj,
  formula,
  data,
  REML = FALSE,
  useWeights = TRUE,
  fxn = identity,
  control = vpcontrol,
  hideErrorsInBackend = FALSE,
  showWarnings = TRUE,
```

```

    BPPARAM = SerialParam(),
    ...
  )

```

### Arguments

exprObj	matrix of expression data (g genes x n samples), or ExpressionSet, or EList returned by voom() from the limma package
formula	specifies variables for the linear (mixed) model. Must only specify covariates, since the rows of exprObj are automatically used as a response. e.g.: ~ a + b + (1 c)
data	data.frame with columns corresponding to formula
REML	use restricted maximum likelihood to fit linear mixed model. default is FALSE. See Details.
useWeights	if TRUE, analysis uses heteroskedastic error estimates from voom(). Value is ignored unless exprObj is an EList() from voom() or weightsMatrix is specified
fxn	apply function to model fit for each gene. Defaults to identify function so it returns the model fit itself
control	control settings for lmer()
hideErrorsInBackend	default FALSE. If TRUE, hide errors in attr(., "errors") and attr(., "error.initial")
showWarnings	default TRUE. Indicate model failures
BPPARAM	parameters for parallel evaluation
...	Additional arguments for lmer() or lm()

### Details

A linear (mixed) model is fit for each gene in exprObj, using formula to specify variables in the regression. If categorical variables are modeled as random effects (as is recommended), then a linear mixed model is used. For example if formula is ~ a + b + (1|c), then the model is

```
fit <- lmer( exprObj[j,] ~ a + b + (1|c), data=data)
```

If there are no random effects, so formula is ~ a + b + c, a 'standard' linear model is used:

```
fit <- lm( exprObj[j,] ~ a + b + c, data=data)
```

In both cases, useWeights=TRUE causes weightsMatrix[j,] to be included as weights in the regression model.

Note: Fitting the model for 20,000 genes can be computationally intensive. To accelerate computation, models can be fit in parallel using BiocParallel to run in parallel. Parallel processing must be enabled before calling this function. See below.

The regression model is fit for each gene separately. Samples with missing values in either gene expression or metadata are omitted by the underlying call to lm/lmer.

Since this function returns a list of each model fit, using this function is slower and uses more memory than fitExtractVarPartModel().

REML=FALSE uses maximum likelihood to estimate variance fractions. This approach produced unbiased estimates, while REML=TRUE can show substantial bias. See Vignette "3) Theory and practice of random effects and REML"

**Value**

list() of where each entry is a model fit produced by lmer() or lm()

**Examples**

```
# load library
# library(variancePartition)

library(BiocParallel)

# load simulated data:
# geneExpr: matrix of gene expression values
# info: information/metadata about each sample
data(varPartData)

# Specify variables to consider
# Age is continuous so we model it as a fixed effect
# Individual and Tissue are both categorical, so we model them as random effects
form <- ~ Age + (1 | Individual) + (1 | Tissue)

# Step 1: fit linear mixed model on gene expression
# If categorical variables are specified, a linear mixed model is used
# If all variables are modeled as continuous, a linear model is used
# each entry in results is a regression model fit on a single gene
# Step 2: extract variance fractions from each model fit
# for each gene, returns fraction of variation attributable to each variable
# Interpretation: the variance explained by each variable
# after correction for all other variables
varPart <- fitExtractVarPartModel(geneExpr, form, info)

# violin plot of contribution of each variable to total variance
# also sort columns
plotVarPart(sortCols(varPart))

# Advanced:
# Fit model and extract variance in two separate steps
# Step 1: fit model for each gene, store model fit for each gene in a list
results <- fitVarPartModel(geneExpr, form, info)

# Step 2: extract variance fractions
varPart <- extractVarPart(results)

# Note: fitVarPartModel also accepts ExpressionSet
data(sample.ExpressionSet, package = "Biobase")

# ExpressionSet example
form <- ~ (1 | sex) + (1 | type) + score
info2 <- Biobase::pData(sample.ExpressionSet)
results2 <- fitVarPartModel(sample.ExpressionSet, form, info2)
```

**Description**

Extract contrast matrix, L, testing a single variable. Contrasts involving more than one variable can be constructed by modifying L directly

**Usage**

```
getContrast(exprObj, formula, data, coefficient)
```

**Arguments**

exprObj	matrix of expression data (g genes x n samples), or ExpressionSet, or EList returned by voom() from the limma package
formula	specifies variables for the linear (mixed) model. Must only specify covariates, since the rows of exprObj are automatically used as a response. e.g.: ~ a + b + (1 c) Formulas with only fixed effects also work
data	data.frame with columns corresponding to formula
coefficient	the coefficient to use in the hypothesis test

**Value**

Contrast matrix testing one variable

**Examples**

```
# load simulated data:
# geneExpr: matrix of gene expression values
# info: information/metadata about each sample
data(varPartData)

# get contrast matrix testing if the coefficient for Batch2 is zero
# The variable of interest must be a fixed effect
form <- ~ Batch + (1 | Individual) + (1 | Tissue)
L <- getContrast(geneExpr, form, info, "Batch3")

# get contrast matrix testing if Batch3 - Batch2 = 0
form <- ~ Batch + (1 | Individual) + (1 | Tissue)
L <- getContrast(geneExpr, form, info, c("Batch3", "Batch2"))

# To test against Batch1 use the formula:
# ~ 0 + Batch + (1|Individual) + (1|Tissue)
# to estimate Batch1 directly instead of using it as the baseline
```

---

getTreat

*Test if coefficient is different from a specified value*

---

**Description**

Test if coefficient is different from a specified value

**Usage**

```

getTreat(fit, lfc = log2(1.2), coef = 1, number = 10, sort.by = "p")

## S4 method for signature 'MArrayLM'
getTreat(fit, lfc = log2(1.2), coef = 1, number = 10, sort.by = "p")

## S4 method for signature 'MArrayLM2'
getTreat(fit, lfc = log2(1.2), coef = 1, number = 10, sort.by = "p")

```

**Arguments**

fit	fit
lfc	a minimum log <sub>2</sub> -fold-change below which changes not considered scientifically meaningful
coef	which coefficient to test
number	number of genes to return
sort.by	column to sort by

**Value**

results of getTreat

**Examples**

```

data(varPartData)

form <- ~ Age + Batch + (1 | Individual) + (1 | Tissue)

fit <- dream(geneExpr, form, info)
fit <- eBayes(fit)

coef <- "Age"

# Evaluate treat()/topTreat() in a way that works seamlessly for dream()
getTreat(fit, lfc = log2(1.03), coef, sort.by = "none", number = 3)

```

---

get\_prediction

*Compute predicted value of formula for linear (mixed) model*

---

**Description**

Compute predicted value of formula for linear (mixed) model for with lm or lmer

**Usage**

```

get_prediction(fit, formula)

## S4 method for signature 'lmerMod'
get_prediction(fit, formula)

## S4 method for signature 'lm'
get_prediction(fit, formula)

```



**Arguments**

fit	model fit with lm or lmer
formula	formula of fixed and random effects to predict

**Details**

Similar motivation as `lme4::predict.merMod()`, but that function cannot use just a subset of the fixed effects: it either uses none or all. Note that the intercept is included in the formula by default. To exclude it from the prediction use `~ 0 + ...` syntax

**Value**

Predicted values from formula using parameter estimates from fit linear (mixed) model

**Examples**

```
library(lme4)

# Linear model
fit <- lm(Reaction ~ Days, sleepstudy)

# prediction of intercept
get_prediction(fit, ~1)

# prediction of Days without intercept
get_prediction(fit, ~ 0 + Days)

# Linear mixed model

# fit model
fm1 <- lmer(Reaction ~ Days + (Days | Subject), sleepstudy)

# predict Days, but exclude intercept
get_prediction(fm1, ~ 0 + Days)

# predict Days and (Days | Subject) random effect, but exclude intercept
get_prediction(fm1, ~ 0 + Days + (Days | Subject))
```

---

ggColorHue

*Default colors for ggplot*

---

**Description**

Return an array of n colors the same as the default used by ggplot2

**Usage**

```
ggColorHue(n)
```

**Arguments**

n	number of colors
---	------------------

**Value**

array of colors of length n

**Examples**

```
ggColorHue(4)
```

---

```
hatvalues, MArrayLM-method
      Compute hatvalues
```

---

**Description**

Compute hatvalues from dream fit

**Usage**

```
## S4 method for signature 'MArrayLM'
hatvalues(model, vobj, ...)

## S4 method for signature 'MArrayLM2'
hatvalues(model, ...)
```

**Arguments**

model	model fit from dream()
vobj	EList returned by voom() or voomWithDreamWeights().
...	other arguments, currently ignored

---

```
isRunnableFormula      Test if formula is full rank on this dataset
```

---

**Description**

Test if formula is full rank on this dataset

**Usage**

```
isRunnableFormula(exprObj, formula, data)
```

**Arguments**

exprObj	expression object
formula	formula
data	data

---

logLik.MArrayLM	<i>Log-likelihood from model fit</i>
-----------------	--------------------------------------

---

**Description**

Log-likelihood from model fit

**Usage**

```
## S3 method for class 'MArrayLM'  
logLik(object, vobj, ...)
```

**Arguments**

object	result of lmFit() or dream()
vobj	EList used to fit model
...	See ?stats::logLik

---

logLik.MArrayLM2	<i>Log-likelihood from model fit</i>
------------------	--------------------------------------

---

**Description**

Log-likelihood from model fit

**Usage**

```
## S3 method for class 'MArrayLM2'  
logLik(object, ...)
```

**Arguments**

object	result of lmFit() or dream()
...	See ?stats::logLik

---

makeContrastsDream      *Construct Matrix of Custom Contrasts*

---

### Description

Construct the contrast matrix corresponding to specified contrasts of a set of parameters. Each specified set of contrast weights must sum to 1.

### Usage

```
makeContrastsDream(
  formula,
  data,
  ...,
  contrasts = NULL,
  suppressWarnings = FALSE,
  nullOnError = FALSE
)
```

### Arguments

formula	specifies variables for the linear (mixed) model. Must only specify covariates, since the rows of exprObj are automatically used as a response. e.g.: $\sim a + b + (1 c)$ Formulas with only fixed effects also work
data	data.frame with columns corresponding to formula
...	expressions, or character strings which can be parsed to expressions, specifying contrasts
contrasts	character vector specifying contrasts
suppressWarnings	(default FALSE). suppress warnings for univariate contrasts
nullOnError	(default FALSE). When a contrast entry is invalid, throw warning and return NULL for that contrast entry

### Details

This function expresses contrasts between a set of parameters as a numeric matrix. The parameters are usually the coefficients from a linear (mixed) model fit, so the matrix specifies which comparisons between the coefficients are to be extracted from the fit. The output from this function is usually used as input to dream().

This function creates a matrix storing the contrasts weights that are applied to each coefficient.

Consider a variable  $v$  with levels  $c('A', 'B', 'C')$ . A contrast comparing A and B is ' $vA - vB$ ' and tests whether the difference between these levels is different than zero. Coded for the 3 levels this has weights  $c(1, -1, 0)$ . In order to compare A to the other levels, the contrast is ' $vA - (vB + vC)/2$ ' so that A is compared to the average of the other two levels. This is encoded as  $c(1, -0.5, -0.5)$ . This type of proper matching in testing multiple levels is enforced by ensuring that the contrast weights sum to 1. Based on standard regression theory only weighted sums of the estimated coefficients are supported.

This function is inspired by `limma::makeContrasts()` but is designed to be compatible with linear mixed models for dream()

Names in ... and contrasts will be used as column names in the returned value.

**Value**

matrix of linear contrasts between regression coefficients

**See Also**

plotContrasts()

**Examples**

```
# load library
# library(variancePartition)

library(BiocParallel)

# load simulated data:
# geneExpr: matrix of gene expression values
# info: information/metadata about each sample
data(varPartData)

form <- ~ 0 + Batch + (1 | Individual) + (1 | Tissue)

# Define contrasts
# Note that for each contrast, the weights sum to 1
L <- makeContrastsDream(form, info, contrasts = c(Batch1_vs_2 = "Batch1 - Batch2", Batch3_vs_4 = "Batch3 - Batch4"))

# show contrasts matrix
L

# Plot to visualize contrasts matrix
plotContrasts(L)

# Fit linear mixed model for each gene
# run on just 10 genes for time
fit <- dream(geneExpr[1:10, ], form, info, L = L)

# examine contrasts after fitting
head(coef(fit))

# show results from first contrast
topTable(fit, coef = "Batch1_vs_2")

# show results from second contrast
topTable(fit, coef = "Batch3_vs_4")

# show results from third contrast
topTable(fit, coef = "Batch1_vs_34")
```

---

MArrayLM2-class

*Class MArrayLM2*


---

**Description**

Class MArrayLM2

---

mvTest

*Multivariate tests on results from dream()*


---

### Description

Evaluate multivariate tests on results from `dream()` using `vcov()` to compute the covariance between estimated regression coefficients across multiple responses. A joint test to see if the coefficients are jointly different from zero is performed using meta-analysis methods that account for the covariance.

### Usage

```
mvTest(
  fit,
  vobj,
  features,
  coef,
  method = c("FE.empirical", "FE", "RE2C", "tstat", "hotelling", "sidak", "fisher"),
  shrink.cov = TRUE,
  BPPARAM = SerialParam(),
  ...
)

## S4 method for signature 'MArrayLM,EList,vector'
mvTest(
  fit,
  vobj,
  features,
  coef,
  method = c("FE.empirical", "FE", "RE2C", "tstat", "hotelling", "sidak", "fisher"),
  shrink.cov = TRUE,
  BPPARAM = SerialParam(),
  ...
)

## S4 method for signature 'MArrayLM,EList,missing'
mvTest(
  fit,
  vobj,
  features,
  coef,
  method = c("FE.empirical", "FE", "RE2C", "tstat", "hotelling", "sidak", "fisher"),
  shrink.cov = TRUE,
  BPPARAM = SerialParam(),
  ...
)

## S4 method for signature 'MArrayLM,EList,list'
mvTest(
  fit,
  vobj,
```

```

    features,
    coef,
    method = c("FE.empirical", "FE", "RE2C", "tstat", "hotelling", "sidak", "fisher"),
    shrink.cov = TRUE,
    BPPARAM = SerialParam(),
    ...
)

## S4 method for signature 'mvTest_input,ANY,ANY'
mvTest(
  fit,
  vobj,
  features,
  coef,
  method = c("FE.empirical", "FE", "RE2C", "tstat", "hotelling", "sidak", "fisher"),
  shrink.cov = TRUE,
  BPPARAM = SerialParam(),
  ...
)

## S4 method for signature 'MArrayLM,matrix,ANY'
mvTest(
  fit,
  vobj,
  features,
  coef,
  method = c("FE.empirical", "FE", "RE2C", "tstat", "hotelling", "sidak", "fisher"),
  shrink.cov = TRUE,
  BPPARAM = SerialParam(),
  ...
)

```

### Arguments

fit	MArrayLM or MArrayLM2 returned by dream()
vobj	matrix or EList object returned by voom()
features	a) indeces or names of features to perform multivariate test on, b) list of indeces or names. If missing, perform joint test on all features.
coef	name of coefficient or contrast to be tested
method	statistical method used to perform multivariate test. See details. 'FE' is a fixed effect test that models the covariance between coefficients. 'FE.empirical' use compute empirical p-values by sampling from the null distribution and fitting with a gamma. 'RE2C' is a random effect test of heterogeneity of the estimated coefficients that models the covariance between coefficients, and also incorporates a fixed effects test too. 'tstat' combines the t-statistics and models the covariance between coefficients. 'hotelling' performs the Hotelling T2 test. 'sidak' returns the smallest p-value and accounting for the number of tests. 'fisher' combines the p-value using Fisher's method assuming independent tests.
shrink.cov	shrink the covariance matrix between coefficients using the Schafer-Strimmer method

BPPARAM            parameters for parallel evaluation  
 ...                other arguments

### Details

See package `remaCor` for details about the `remaCor::RE2C()` test, and see `remaCor::LS()` for details about the fixed effect test. When only 1 feature is selected, the original p-value is returned and the test statistic is set to NA.

For the "RE2C" test, the final test statistic is the sum of a test statistic for the mean effect (`stat.FE`) and heterogeneity across effects (`stat.het`). `mvTest()` returns 0 if `stat.het` is negative in extremely rare cases.

### Value

Returns a data.frame with the statistics from each test, the pvalue from the test, `n_features`, `method`, and `lambda` from the Schafer-Strimmer method to shrink the estimated covariance. When `shrink.cov=FALSE`, `lambda = 0`.

### Examples

```
# library(variancePartition)
library(edgeR)
library(BiocParallel)

data(varPartDEdata)

# normalize RNA-seq counts
dge <- DGEList(counts = countMatrix)
dge <- calcNormFactors(dge)

# specify formula with random effect for Individual
form <- ~ Disease + (1 | Individual)

# compute observation weights
vobj <- voomWithDreamWeights(dge[1:20, ], form, metadata)

# fit dream model
fit <- dream(vobj, form, metadata)
fit <- eBayes(fit)

# Multivariate test of features 1 and 2
mvTest(fit, vobj, 1:2, coef = "Disease1")

# Test multiple sets of features
lst <- list(a = 1:2, b = 3:4)
mvTest(fit, vobj, lst, coef = "Disease1", BPPARAM = SnowParam(2))
```

---

mvTest\_input-class      *Class mvTest\_input*

---

### Description

Class `mvTest_input` work is with `iterRowsSplit()`



---

plotCompareP

*Compare p-values from two analyses*


---

### Description

Plot  $-\log_{10}$  p-values from two analyses and color based on donor component from variancePartition analysis

### Usage

```
plotCompareP(
  p1,
  p2,
  vpDonor,
  dupcorvalue,
  fraction = 0.2,
  xlabel = bquote(duplicateCorrelation ~ (-log[10] ~ p)),
  ylabel = bquote(dream ~ (-log[10] ~ p))
)
```

### Arguments

p1	p-value from first analysis
p2	p-value from second analysis
vpDonor	donor component for each gene from variancePartition analysis
dupcorvalue	scalar donor component from duplicateCorrelation
fraction	fraction of highest/lowest values to use for best fit lines
xlabel	for x-axis
ylabel	label for y-axis

### Value

ggplot2 plot

### Examples

```
# load library
# library(variancePartition)

library(BiocParallel)

# load simulated data:
# geneExpr: matrix of gene expression values
# info: information/metadata about each sample
data(varPartData)

# Perform very simple analysis for demonstration

# Analysis 1
form <- ~Batch
fit <- dream(geneExpr, form, info)
```

```
fit <- eBayes(fit)
res <- topTable(fit, number = Inf, coef = "Batch3")

# Analysis 2
form <- ~ Batch + (1 | Tissue)
fit2 <- dream(geneExpr, form, info)
res2 <- topTable(fit2, number = Inf, coef = "Batch3")

# Compare p-values
plotCompareP(res$P.Value, res2$P.Value, runif(nrow(res)), .3)
```

---

plotContrasts

*Plot representation of contrast matrix*

---

## Description

Plot contrast matrix to clarify interpretation of hypothesis tests with linear contrasts

## Usage

```
plotContrasts(L)
```

## Arguments

L contrast matrix

## Details

This plot shows the contrasts weights that are applied to each coefficient.

Consider a variable  $v$  with levels  $c('A', 'B', 'C')$ . A contrast comparing A and B is ' $vA - vB$ ' and tests whether the difference between these levels is different than zero. Coded for the 3 levels this has weights  $c(1, -1, 0)$ . In order to compare A to the other levels, the contrast is ' $vA - (vB + vC)/2$ ' so that A is compared to the average of the other two levels. This is encoded as  $c(1, -0.5, -0.5)$ . This type of proper matching in testing multiple levels is enforced by ensuring that the contrast weights sum to 1. Based on standard regression theory only weighted sums of the estimated coefficients are supported.

## Value

ggplot2 object

## See Also

makeContrastsDream()

**Examples**

```

# load library
# library(variancePartition)

# load simulated data:
# geneExpr: matrix of gene expression values
# info: information/metadata about each sample
data(varPartData)

# 1) get contrast matrix testing if the coefficient for Batch2 is different from Batch3
form <- ~ Batch + (1 | Individual) + (1 | Tissue)
L <- makeContrastsDream(form, info, contrasts = c(Batch_3_vs_2 = "Batch3 - Batch2"))

# plot contrasts
plotContrasts(L)

```

---

plotCorrMatrix

*plotCorrMatrix*


---

**Description**

Plot correlation matrix

**Usage**

```

plotCorrMatrix(
  C,
  dendrogram = "both",
  sort = TRUE,
  margins = c(13, 13),
  key.xlab = "correlation",
  ...
)

```

**Arguments**

C	correlation matrix: R or R <sup>2</sup> matrix
dendrogram	character string indicating whether to draw 'both' or none'
sort	sort rows and columns based on clustering
margins	spacing of plot
key.xlab	label of color gradient
...	additional arguments to heatmap.2

**Details**

Plots image of correlation matrix using customized call to heatmap.2

**Value**

Image of correlation matrix

**Examples**

```
# simulate simple matrix of 10 variables
mat <- matrix(rnorm(1000), ncol = 10)

# compute correlation matrix
C <- cor(mat)

# plot correlations
plotCorrMatrix(C)

# plot squared correlations
plotCorrMatrix(C^2, dendrogram = "none")
```

---

plotCorrStructure      *plotCorrStructure*

---

**Description**

Plot correlation structure of a gene based on random effects

**Usage**

```
plotCorrStructure(
  fit,
  varNames = names(coef(fit)),
  reorder = TRUE,
  pal = colorRampPalette(c("white", "red", "darkred")),
  hclust.method = "complete"
)
```

**Arguments**

fit	linear mixed model fit of a gene produced by lmer() or fitVarPartModel()
varNames	variables in the metadata for which the correlation structure should be shown. Variables must be random effects
reorder	how to reorder the rows/columns of the correlation matrix. reorder=FALSE gives no reorder. reorder=TRUE reorders based on hclust. reorder can also be an array of indices to reorder the samples manually
pal	color palette
hclust.method	clustering methods for hclust

**Value**

Image of correlation structure between each pair of experiments for a single gene

**Examples**

```

# load library
# library(variancePartition)

library(BiocParallel)

# load simulated data:
data(varPartData)

# specify formula
form <- ~ Age + (1 | Individual) + (1 | Tissue)

# fit and return linear mixed models for each gene
fitList <- fitVarPartModel(geneExpr[1:10, ], form, info)

# Focus on the first gene
fit <- fitList[[1]]

# plot correlation structure based on Individual, reordering samples with hclust
plotCorrStructure(fit, "Individual")

# don't reorder
plotCorrStructure(fit, "Individual", reorder = FALSE)

# plot correlation structure based on Tissue, reordering samples with hclust
plotCorrStructure(fit, "Tissue")

# don't reorder
plotCorrStructure(fit, "Tissue", FALSE)

# plot correlation structure based on all random effects
# reorder manually by Tissue and Individual
idx <- order(info$Tissue, info$Individual)
plotCorrStructure(fit, reorder = idx)

# plot correlation structure based on all random effects
# reorder manually by Individual, then Tissue
idx <- order(info$Individual, info$Tissue)
plotCorrStructure(fit, reorder = idx)

```

---

plotPercentBars      *Bar plot of gene fractions*

---

**Description**

Bar plot of fractions for a subset of genes

**Usage**

```

plotPercentBars(
  x,
  col = c(ggColorHue(ncol(x) - 1), "grey85"),
  genes = rownames(x),

```

```

    width = NULL,
    ...
)

## S4 method for signature 'matrix'
plotPercentBars(
  x,
  col = c(ggColorHue(ncol(x) - 1), "grey85"),
  genes = rownames(x),
  width = NULL,
  ...
)

## S4 method for signature 'data.frame'
plotPercentBars(
  x,
  col = c(ggColorHue(ncol(x) - 1), "grey85"),
  genes = rownames(x),
  width = NULL,
  ...
)

## S4 method for signature 'varPartResults'
plotPercentBars(
  x,
  col = c(ggColorHue(ncol(x) - 1), "grey85"),
  genes = rownames(x),
  width = NULL,
  ...
)

```

### Arguments

x	object storing fractions
col	color of bars for each variable
genes	name of genes to plot
width	specify width of bars
...	other arguments

### Value

Returns ggplot2 barplot

### Examples

```

# library(variancePartition)

library(BiocParallel)

# load simulated data:
# geneExpr: matrix of gene expression values
# info: information/metadata about each sample
data(varPartData)

```

```

# Specify variables to consider
form <- ~ Age + (1 | Individual) + (1 | Tissue)

# Fit model
varPart <- fitExtractVarPartModel(geneExpr, form, info)

# Bar plot for a subset of genes showing variance fractions
plotPercentBars(varPart[1:5, ])

# Move the legend to the top
plotPercentBars(varPart[1:5, ]) + theme(legend.position = "top")

```

---

plotStratify

*plotStratify*


---

## Description

Plot gene expression stratified by another variable

## Usage

```

plotStratify(
  formula,
  data,
  xlab,
  ylab,
  main,
  sortBy,
  colorBy,
  sort = TRUE,
  text = NULL,
  text.y = 1,
  text.size = 5,
  pts.cex = 1,
  ylim = NULL,
  legend = TRUE,
  x.labels = FALSE
)

```

## Arguments

formula	specify variables shown in the x- and y-axes. Y-axis should be continuous variable, x-axis should be discrete.
data	data.frame storing continuous and discrete variables specified in formula
xlab	label x-axis. Defaults to value of xval
ylab	label y-axis. Defaults to value of yval
main	main label
sortBy	name of column in geneExpr to sort samples by. Defaults to xval

<code>colorBy</code>	name of column in <code>geneExpr</code> to color box plots. Defaults to <code>xval</code>
<code>sort</code>	if TRUE, sort boxplots by median value, else use default ordering
<code>text</code>	plot text on the top left of the plot
<code>text.y</code>	indicate position of the text on the y-axis as a fraction of the y-axis range
<code>text.size</code>	size of text
<code>pts.cex</code>	size of points
<code>ylim</code>	specify range of y-axis
<code>legend</code>	show legend
<code>x.labels</code>	show x axis labels

**Value**

ggplot2 object

**Examples**

```
# Note: This is a newer, more convient interface to plotStratifyBy()

# load library
# library(variancePartition)

# load simulated data:
data(varPartData)

# Create data.frame with expression and Tissue information for each sample
GE <- data.frame(Expression = geneExpr[1, ], Tissue = info$Tissue)

# Plot expression stratified by Tissue
plotStratify(Expression ~ Tissue, GE)

# Omit legend and color boxes grey
plotStratify(Expression ~ Tissue, GE, colorBy = NULL)

# Specify colors
col <- c(B = "green", A = "red", C = "yellow")
plotStratify(Expression ~ Tissue, GE, colorBy = col, sort = FALSE)
```

---

`plotStratifyBy`

*plotStratifyBy*

---

**Description**

Plot gene expression stratified by another variable



**Usage**

```
plotStratifyBy(  
  geneExpr,  
  xval,  
  yval,  
  xlab = xval,  
  ylab = yval,  
  main = NULL,  
  sortBy = xval,  
  colorBy = xval,  
  sort = TRUE,  
  text = NULL,  
  text.y = 1,  
  text.size = 5,  
  pts.cex = 1,  
  ylim = NULL,  
  legend = TRUE,  
  x.labels = FALSE  
)
```

**Arguments**

geneExpr	data.frame of gene expression values and another variable for each sample. If there are multiple columns, the user can specify which one to use
xval	name of column in geneExpr to be used along x-axis to stratify gene expression
yval	name of column in geneExpr indicating gene expression
xlab	label x-axis. Defaults to value of xval
ylab	label y-axis. Defaults to value of yval
main	main label
sortBy	name of column in geneExpr to sort samples by. Defaults to xval
colorBy	name of column in geneExpr to color box plots. Defaults to xval
sort	if TRUE, sort boxplots by median value, else use default ordering
text	plot text on the top left of the plot
text.y	indicate position of the text on the y-axis as a fraction of the y-axis range
text.size	size of text
pts.cex	size of points
ylim	specify range of y-axis
legend	show legend
x.labels	show x axis labels

**Value**

ggplot2 object

**Examples**

```

# load library
# library(variancePartition)

# load simulated data:
data(varPartData)

# Create data.frame with expression and Tissue information for each sample
GE <- data.frame(Expression = geneExpr[1, ], Tissue = info$Tissue)

# Plot expression stratified by Tissue
plotStratifyBy(GE, "Tissue", "Expression")

# Omit legend and color boxes grey
plotStratifyBy(GE, "Tissue", "Expression", colorBy = NULL)

# Specify colors
col <- c(B = "green", A = "red", C = "yellow")
plotStratifyBy(GE, "Tissue", "Expression", colorBy = col, sort = FALSE)

```

---

plotVarianceEstimates *Plot Variance Estimates*

---

**Description**

Plot Variance Estimates

**Usage**

```

plotVarianceEstimates(
  fit,
  fitEB,
  var_true = NULL,
  xmax = quantile(fit$sigma^2, 0.999)
)

```

**Arguments**

fit	model fit from dream()
fitEB	model fit from eBayes()
var_true	array of true variance values from simulation (optional)
xmax	maximum value on the x-axis

---

plotVarPart	<i>Violin plot of variance fractions</i>
-------------	--

---

**Description**

Violin plot of variance fraction for each gene and each variable

**Usage**

```
plotVarPart(  
  obj,  
  col = c(ggColorHue(ncol(obj) - 1), "grey85"),  
  label.angle = 20,  
  main = "",  
  ylab = "",  
  convertToPercent = TRUE,  
  ...  
)  
  
## S4 method for signature 'matrix'  
plotVarPart(  
  obj,  
  col = c(ggColorHue(ncol(obj) - 1), "grey85"),  
  label.angle = 20,  
  main = "",  
  ylab = "",  
  convertToPercent = TRUE,  
  ...  
)  
  
## S4 method for signature 'data.frame'  
plotVarPart(  
  obj,  
  col = c(ggColorHue(ncol(obj) - 1), "grey85"),  
  label.angle = 20,  
  main = "",  
  ylab = "",  
  convertToPercent = TRUE,  
  ...  
)  
  
## S4 method for signature 'varPartResults'  
plotVarPart(  
  obj,  
  col = c(ggColorHue(ncol(obj) - 1), "grey85"),  
  label.angle = 20,  
  main = "",  
  ylab = "",  
  convertToPercent = TRUE,  
  ...  
)
```

**Arguments**

obj	varParFrac object returned by fitExtractVarPart or extractVarPart
col	vector of colors
label.angle	angle of labels on x-axis
main	title of plot
ylab	text on y-axis
convertToPercent	multiply fractions by 100 to convert to percent values
...	additional arguments

**Value**

Makes violin plots of variance components model. This function uses the graphics interface from ggplot2. Warnings produced by this function usually ggplot2 warning that the window is too small.

**Examples**

```
# load library
# library(variancePartition)

library(BiocParallel)

# load simulated data:
# geneExpr: matrix of gene expression values
# info: information/metadata about each sample
data(varPartData)

# Specify variables to consider
# Age is continuous so we model it as a fixed effect
# Individual and Tissue are both categorical, so we model them as random effects
form <- ~ Age + (1 | Individual) + (1 | Tissue)

varPart <- fitExtractVarPartModel(geneExpr, form, info)

# violin plot of contribution of each variable to total variance
plotVarPart(sortCols(varPart))
```

---

rdf

*Residual degrees of freedom*


---

**Description**

Residual degrees of freedom

**Usage**

```
rdf(fit)
```

**Arguments**

fit	model fit from lm(), glm(), lmer()
-----	------------------------------------

**See Also**

rdf.merMod

**Examples**

```
library(lme4)

fit <- lm(Reaction ~ Days, sleepstudy)
rdf(fit)
```

---

rdf.merMod

*Approximate residual degrees of freedom*

---

**Description**

For a linear model with  $n$  samples and  $p$  covariates,  $RSS/\sigma^2 \sim \chi_\nu^2$  where  $\nu = n - p$  is the residual degrees of freedom. In the case of a linear mixed model, the distribution is no longer exactly a chi-square distribution, but can be approximated with a chi-square distribution.

Given the hat matrix,  $H$ , that maps between observed and fitted responses, the approximate residual degrees of freedom is  $\nu = tr((I - H)^T(I - H))$ . For a linear model, this simplifies to the well known form  $\nu = n - p$ . In the more general case, such as a linear mixed model, the original form simplifies only to  $n - 2tr(H) + tr(HH)$  and is an approximation rather than being exact. The third term here is quadratic time in the number of samples,  $n$ , and can be computationally expensive to evaluate for larger datasets. Here we develop a linear time algorithm that takes advantage of the fact that  $H$  is low rank.

$H$  is computed as  $A^T A + B^T B$  for  $A=CL$  and  $B=CR$  defined in the code. Since  $A$  and  $B$  are low rank, there is no need to compute  $H$  directly. Instead, the terms  $tr(H)$  and  $tr(HH)$  can be computed using the eigen decompositions of  $AA^T$  and  $BB^T$  which is linear time in the number of samples.

**Usage**

```
rdf.merMod(model, method = c("linear", "quadratic"))
```

**Arguments**

model	An object of class merMod
method	Use algorithm that is "linear" (default) or quadratic time in the number of samples

**Details**

Compute the approximate residual degrees of freedom from a linear mixed model.

**Value**

residual degrees of freedom

**See Also**

rdf\_from\_matrices

**Examples**

```
library(lme4)

# Fit linear mixed model
fit <- lmer(Reaction ~ Days + (Days | Subject), sleepstudy)

# Evaluate the approximate residual degrees of freedom
rdf.merMod(fit)
```

---

rdf\_from\_matrices      *Fast approximate residual degrees of freedom*

---

**Description**

Defining  $H = A^T A + B^T B$  where  $A$  and  $B$  are low rank, compute  $n - 2tr(H) + tr(HH)$  in  $O(np^2)$  instead of  $O(n^2p^2)$ .

**Usage**

```
rdf_from_matrices(A, B)
```

**Arguments**

$A$                     a matrix or sparseMatrix  
 $B$                     a matrix or sparseMatrix

**See Also**

```
rdf.merMod
```

---

reOnly                    *Adapted from lme4:::reOnly*

---

**Description**

Adapted from lme4:::reOnly

**Usage**

```
reOnly(f, response = FALSE)
```

**Arguments**

$f$                     formula  
response              (FALSE) is there a response in the formula

---

```
residuals,MArrayLM-method
      residuals for MArrayLM
```

---

**Description**

residuals for MArrayLM

**Usage**

```
## S4 method for signature 'MArrayLM'
residuals(object, y, ..., type = c("response", "pearson"))
```

**Arguments**

object	MArrayLM object from dream
y	EList object used in dream()
...	other arguments, currently ignored
type	compute either response or pearson residuals

**Value**

results of residuals

---

```
residuals,MArrayLM2-method
      residuals for MArrayLM2
```

---

**Description**

residuals for MArrayLM2

**Usage**

```
## S4 method for signature 'MArrayLM2'
residuals(object, y, type = c("response", "pearson"), ...)
```

**Arguments**

object	MArrayLM2 object from dream
y	EList object used in dream()
type	compute either response or pearson residuals
...	other arguments, currently ignored

**Value**

results of residuals

---

residuals, VarParFitList-method  
*Residuals from model fit*

---

### Description

Extract residuals for each gene from model fit with fitVarPartModel()

### Usage

```
## S4 method for signature 'VarParFitList'  
residuals(object, ...)
```

### Arguments

object	object produced by fitVarPartModel()
...	other arguments.

### Details

If model is fit with missing data, residuals returns NA for entries that were missing in the original data

### Value

Residuals extracted from model fits stored in object

### Examples

```
# load library  
# library(variancePartition)  
  
library(BiocParallel)  
  
# load simulated data:  
# geneExpr: matrix of gene expression values  
# info: information/metadata about each sample  
data(varPartData)  
  
# Specify variables to consider  
# Age is continuous so we model it as a fixed effect  
# Individual and Tissue are both categorical, so we model them as random effects  
form <- ~ Age + (1 | Individual) + (1 | Tissue)  
  
# Fit model  
modelFit <- fitVarPartModel(geneExpr, form, info)  
  
# Extract residuals of model fit  
res <- residuals(modelFit)
```



---

residuals.MArrayLM2     *Residuals for result of dream*

---

### Description

Residuals for result of dream

### Usage

```
residuals.MArrayLM2(object, y, ..., type = c("response", "pearson"))
```

### Arguments

object	See ?stats::residuals
y	EList object used in dream()
...	See ?stats::residuals
type	compute either response or pearson residuals

---

shrinkageMetric     *Shrinkage metric for eBayes*

---

### Description

Evaluates the coefficient from the linear regression of  $s2.post \sim \sigma Sq$ . When there is no shrinkage, this value is 1. Values less than 1 indicate the amount of shrinkage.

### Usage

```
shrinkageMetric(sigmaSq, s2.post)
```

### Arguments

sigmaSq	maximum likelihood residual variance for every gene
s2.post	empirical Bayes posterior estimate of residual variance for every gene

### Details

Shrinkage metric for eBayes quantifying the amount of shrinkage that is applied to shrink the maximum likelihood residual variance to the empirical Bayes posterior estimate

---

sortCols	<i>Sort variance partition statistics</i>
----------	---

---

### Description

Sort columns returned by `extractVarPart()` or `fitExtractVarPartModel()`

### Usage

```
sortCols(
  x,
  FUN = median,
  decreasing = TRUE,
  last = c("Residuals", "Measurement.error"),
  ...
)

## S4 method for signature 'matrix'
sortCols(
  x,
  FUN = median,
  decreasing = TRUE,
  last = c("Residuals", "Measurement.error"),
  ...
)

## S4 method for signature 'data.frame'
sortCols(
  x,
  FUN = median,
  decreasing = TRUE,
  last = c("Residuals", "Measurement.error"),
  ...
)

## S4 method for signature 'varPartResults'
sortCols(
  x,
  FUN = median,
  decreasing = TRUE,
  last = c("Residuals", "Measurement.error"),
  ...
)
```

### Arguments

<code>x</code>	object returned by <code>extractVarPart()</code> or <code>fitExtractVarPartModel()</code>
<code>FUN</code>	function giving summary statistic to sort by. Defaults to <code>median</code>
<code>decreasing</code>	logical. Should the sorting be increasing or decreasing?
<code>last</code>	columns to be placed on the right, regardless of values in these columns
<code>...</code>	other arguments to sort

**Value**

data.frame with columns sorted by mean value, with Residuals in last column

**Examples**

```
# library(variancePartition)

library(BiocParallel)

# load simulated data:
# geneExpr: matrix of gene expression values
# info: information/metadata about each sample
data(varPartData)

# Specify variables to consider
# Age is continuous so we model it as a fixed effect
# Individual and Tissue are both categorical, so we model them as random effects
form <- ~ Age + (1 | Individual) + (1 | Tissue)

# Step 1: fit linear mixed model on gene expression
# If categorical variables are specified, a linear mixed model is used
# If all variables are modeled as continuous, a linear model is used
# each entry in results is a regression model fit on a single gene
# Step 2: extract variance fractions from each model fit
# for each gene, returns fraction of variation attributable to each variable
# Interpretation: the variance explained by each variable
# after correction for all other variables
varPart <- fitExtractVarPartModel(geneExpr, form, info)

# violin plot of contribution of each variable to total variance
# sort columns by median value
plotVarPart(sortCols(varPart))
```

---

topTable

*Table of Top Genes from Linear Model Fit*


---

**Description**

topTable generic

topTable generic MArrayLM

topTable generic MArrayLM2

**Usage**

```
topTable(
  fit,
  coef = NULL,
  number = 10,
  genelist = fit$genes,
  adjust.method = "BH",
  sort.by = "B",
```

```

    resort.by = NULL,
    p.value = 1,
    lfc = 0,
    confint = FALSE
)

## S4 method for signature 'MArrayLM'
topTable(
  fit,
  coef = NULL,
  number = 10,
  genelist = fit$genes,
  adjust.method = "BH",
  sort.by = "p",
  resort.by = NULL,
  p.value = 1,
  lfc = 0,
  confint = FALSE
)

## S4 method for signature 'MArrayLM2'
topTable(
  fit,
  coef = NULL,
  number = 10,
  genelist = fit$genes,
  adjust.method = "BH",
  sort.by = "p",
  resort.by = NULL,
  p.value = 1,
  lfc = 0,
  confint = FALSE
)

```

### Arguments

fit	fit
coef	coef
number	number
genelist	genelist
adjust.method	adjust.method
sort.by	sort.by
resort.by	resort.by
p.value	p.value
lfc	lfc
confint	confint

### Value

results of toptable

results of toptable  
 results of toptable

---

VarParCList-class      *Class VarParCList*

---

### Description

Class VarParCList

---

VarParFitList-class      *Class VarParFitList*

---

### Description

Class VarParFitList

---

varParFrac-class      *Class varParFrac*

---

### Description

Class varParFrac

---

varPartConfInf      *Linear mixed model confidence intervals*

---

### Description

Fit linear mixed model to estimate contribution of multiple sources of variation while simultaneously correcting for all other variables. Then perform parametric bootstrap sampling to get a 95% confidence intervals for each variable for each gene.

### Usage

```
varPartConfInf(
  exprObj,
  formula,
  data,
  REML = FALSE,
  useWeights = TRUE,
  control = vpcontrol,
  nsim = 1000,
  ...
)
```

**Arguments**

exprObj	matrix of expression data (g genes x n samples), or ExpressionSet, or EList returned by voom() from the limma package
formula	specifies variables for the linear (mixed) model. Must only specify covariates, since the rows of exprObj are automatically used as a response. e.g.: ~ a + b + (1 c)
data	data.frame with columns corresponding to formula
REML	use restricted maximum likelihood to fit linear mixed model. default is FALSE. Strongly discourage against changing this option, but here for compatibility.
useWeights	if TRUE, analysis uses heteroskedastic error estimates from voom(). Value is ignored unless exprObj is an EList from voom() or weightsMatrix is specified
control	control settings for lmer()
nsim	number of bootstrap datasets
...	Additional arguments for lmer() or lm()

**Details**

A linear mixed model is fit for each gene, and bootMer() is used to generate parametric bootstrap confidence intervals. use.u=TRUE is used so that the  $\hat{u}$  values from the random effects are used as estimated and are not re-sampled. This gives confidence intervals as if additional data were generated from these same current samples. Conversely, use.u=FALSE assumes that this dataset is a sample from a larger population. Thus it simulates  $\hat{u}$  based on the estimated variance parameter. This approach gives confidence intervals as if additional data were collected from the larger population from which this dataset is sampled. Overall, use.u=TRUE gives smaller confidence intervals that are appropriate in this case.

**Value**

list() of where each entry is the result for a gene. Each entry is a matrix of the 95% confidence interval of the variance fraction for each variable

**Examples**

```
# load library
# library(variancePartition)

library(BiocParallel)

# load simulated data:
# geneExpr: matrix of gene expression values
# info: information/metadata about each sample
data(varPartData)

# Specify variables to consider
# Age is continuous so we model it as a fixed effect
# Individual and Tissue are both categorical, so we model them as random effects
form <- ~ Age + (1 | Individual) + (1 | Tissue)

# Compute bootstrap confidence intervals for each variable for each gene
resCI <- varPartConfInf(geneExpr[1:5, ], form, info, nsim = 100)
```

---

varPartData	<i>Simulation dataset for examples</i>
-------------	--

---

**Description**

A simulated dataset of gene expression and metadata

A simulated dataset of gene counts

A simulated dataset of gene counts

A simulated dataset of gene counts

**Usage**

```
data(varPartData)
```

```
data(varPartData)
```

```
data(varPartData)
```

```
data(varPartData)
```

**Format**

A dataset of 100 samples and 200 genes

A dataset of 100 samples and 200 genes

A dataset of 100 samples and 200 genes

A dataset of 100 samples and 200 genes

**Details**

- geneCounts gene expression in the form of RNA-seq counts
- geneExpr gene expression on a continuous scale
- info metadata about the study design
  
- geneCounts gene expression in the form of RNA-seq counts
- geneExpr gene expression on a continuous scale
- info metadata about the study design
  
- geneCounts gene expression in the form of RNA-seq counts
- geneExpr gene expression on a continuous scale
- info metadata about the study design
  
- geneCounts gene expression in the form of RNA-seq counts
- geneExpr gene expression on a continuous scale
- info metadata about the study design

---

varPartDEdata	<i>A simulated dataset of gene counts</i>
---------------	---

---

### Description

- geneCounts gene expression in the form of RNA-seq counts
- geneExpr gene expression on a continuous scale
- info metadata about the study design
- geneCounts gene expression in the form of RNA-seq counts
- geneExpr gene expression on a continuous scale
- info metadata about the study design

### Usage

```
data(varPartData)
```

```
data(varPartData)
```

### Format

A dataset of 24 samples and 19,364 genes

A dataset of 24 samples and 19,364 genes

---

varPartResults-class	<i>Class varPartResults</i>
----------------------	-----------------------------

---

### Description

Class varPartResults

---

vcov,MArrayLM-method	<i>Co-variance matrix for dream() fit</i>
----------------------	---

---

### Description

Define generic vcov() for result of lmFit() and dream()

### Usage

```
## S4 method for signature 'MArrayLM'
vcov(object, vobj, coef)
```

### Arguments

object	MArrayLM object return by lmFit() or dream()
vobj	EList object returned by voom()
coef	name of coefficient to be extracted



**Value**

variance-covariance matrix

---

vcov,MArrayLM2-method *Co-variance matrix for dream() fit*

---

**Description**

Define generic vcov() for result of lmFit() and dream()

**Usage**

```
## S4 method for signature 'MArrayLM2'
vcov(object, vobj, coef)
```

**Arguments**

object	MArrayLM object return by lmFit() or dream()
vobj	EList object returned by voom()
coef	name of coefficient to be extracted

**Value**

variance-covariance matrix

---

vcovSqrt *Sqrt of co-variance matrix for dream() fit*

---

**Description**

Define generic vcovSqrt() for result of lmFit() and dream()

**Usage**

```
vcovSqrt(object, vobj, coef, approx = TRUE)

## S4 method for signature 'MArrayLM'
vcovSqrt(object, vobj, coef, approx = TRUE)

## S4 method for signature 'MArrayLM2'
vcovSqrt(object, vobj, coef, approx = TRUE)
```

**Arguments**

object	MArrayLM object return by lmFit() or dream()
vobj	EList object returned by voom()
coef	name of coefficient to be extracted
approx	use fast approximation

**Value**

Computes factor of covariance matrix so that `vcov(object)` is the same as `crossprod(vcovSqrt(object))`

**Examples**

```
# load simulated data:
# geneExpr: matrix of *normalized* gene expression values
# info: information/metadata about each sample
data(varPartData)

form <- ~Batch

fit <- dream(geneExpr[1:2, ], form, info)
fit <- eBayes(fit)

# Compute covariance directly
Sigma <- vcov(fit, geneExpr[1:2, ])

# Compute factor of covariance
S <- crossprod(vcovSqrt(fit, geneExpr[1:2, ]))
```

---

`voomWithDreamWeights` *Transform RNA-Seq Data Ready for Linear Mixed Modelling with dream()*

---

**Description**

Transform count data to log<sub>2</sub>-counts per million (logCPM), estimate the mean-variance relationship and use this to compute appropriate observation-level weights. The data are then ready for linear mixed modelling with `dream()`. This method is the same as `limma::voom()`, except that it allows random effects in the formula

**Usage**

```
voomWithDreamWeights(
  counts,
  formula,
  data,
  lib.size = NULL,
  normalize.method = "none",
  span = 0.5,
  weights = NULL,
  prior.count = 0.5,
  prior.count.for.weights = prior.count,
  plot = FALSE,
  save.plot = TRUE,
  rescaleWeightsAfter = FALSE,
  scaledByLib = FALSE,
  priorWeightsAsCounts = FALSE,
  BPPARAM = SerialParam(),
  ...
)
```

**Arguments**

<code>counts</code>	a numeric matrix containing raw counts, or an <code>ExpressionSet</code> containing raw counts, or a <code>DGEList</code> object. Counts must be non-negative and NAs are not permitted.
<code>formula</code>	specifies variables for the linear (mixed) model. Must only specify covariates, since the rows of <code>exprObj</code> are automatically used as a response. e.g.: <code>~ a + b + (1 c)</code> Formulas with only fixed effects also work, and <code>lmFit()</code> followed by <code>contrasts.fit()</code> are run.
<code>data</code>	<code>data.frame</code> with columns corresponding to formula
<code>lib.size</code>	numeric vector containing total library sizes for each sample. Defaults to the normalized (effective) library sizes in counts if <code>counts</code> is a <code>DGEList</code> or to the columnwise count totals if <code>counts</code> is a matrix.
<code>normalize.method</code>	the microarray-style normalization method to be applied to the logCPM values (if any). Choices are as for the <code>method</code> argument of <code>normalizeBetweenArrays</code> when the data is single-channel. Any normalization factors found in counts will still be used even if <code>normalize.method="none"</code> .
<code>span</code>	width of the lowess smoothing window as a proportion. Setting <code>span="auto"</code> uses <code>fANCOVA::loess.as()</code> to estimate the tuning parameter from the data
<code>weights</code>	Can be a numeric matrix of individual weights of same dimensions as the counts, or a numeric vector of sample weights with length equal to <code>ncol(counts)</code>
<code>prior.count</code>	average count to be added to each observation to avoid taking log of zero. The count applied to each sample is normalized by library size so given equal log CPM for a gene with zero counts across multiple samples
<code>prior.count.for.weights</code>	count added to regularize weights
<code>plot</code>	logical, should a plot of the mean-variance trend be displayed?
<code>save.plot</code>	logical, should the coordinates and line of the plot be saved in the output?
<code>rescaleWeightsAfter</code>	default = FALSE, should the output weights be scaled by the input weights
<code>scaledByLib</code>	if TRUE, scale pseudocount by <code>lib.size</code> . Else to standard constant pseudocount addition
<code>priorWeightsAsCounts</code>	if <code>weights</code> is NULL, set weights to be equal to counts, following delta method for log2 CPM
<code>BPPARAM</code>	parameters for parallel evaluation
<code>...</code>	other arguments are passed to <code>lmer</code> .

**Details**

Adapted from `voom()` in `limma` v3.40.2

**Value**

An `EList` object just like the result of `limma::voom()`

**See Also**

`limma::voom()`

**Examples**

```
# library(variancePartition)
library(edgeR)
library(BiocParallel)

data(varPartDEdata)

# normalize RNA-seq counts
dge <- DGEList(counts = countMatrix)
dge <- calcNormFactors(dge)

# specify formula with random effect for Individual
form <- ~ Disease + (1 | Individual)

# compute observation weights
vobj <- voomWithDreamWeights(dge[1:20, ], form, metadata)

# fit dream model
res <- dream(vobj, form, metadata)
res <- eBayes(res)

# extract results
topTable(res, coef = "Disease1", number = 3)
```

---

[.MArrayLM2

*Subsetting for MArrayLM2*

---

**Description**

Enable subsetting on MArrayLM2 object. Same as for MArrayLM, but apply column subsetting to `df.residual` and `cov.coefficients.list`

**Arguments**

object	MArrayLM2
i	row
j	col

**Value**

subset

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